

Variation of micromorphological characters of lemma and palea in the genus *Bromus* (Poaceae)

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Micromorphological features of the lemma and palea were investigated for 77 species in the genus *Bromus* (Poaceae) using scanning electron microscopy. Several micromorphological characters of both long and short cells (cork cells, crown cells, prickles and macrohairs) were observed. All were treated as separate characters, although crown cells, prickles and macrohairs are treated as a single group (exodermic cells) owing to the protrusion of their periclinal wall. The main objectives of this work are to assess the value of the micromorphological typology in systematic studies, and to characterize the six subgenera of *Bromus*, using micromorphological features of the lemmas and paleas.

Key words: *Bromus*, lemma, micromorphology, palea, Poaceae, SEM

Introduction

Micromorphological characters are valuable for systematic studies in the family Poaceae. For example, the absence of microhairs is characteristic in Pooideae (Clayton & Renvoize 1986). Micromorphological features of the leaf epidermis are also of taxonomic value (Prat 1932, 1936, Metcalfe 1960, Ellis 1979, Baum *et al.*

1989, Amarasingue & Watson 1990), being especially useful for the characterization of the larger groups, particularly subfamilies and tribes.

Micromorphological characters of the floral bracts of grasses have been used to assess systematic relationships, as well as evolutionary trends. Furthermore, microcharacters can have taxonomic significance within genera, as in the case of microhairs in *Eragrostis* (Amarasingue

& Watson 1990), or of the structure of the epicuticular waxes in *Avena* (Baum & Hadland 1975) and in *Hordeum* (Baum *et al.* 1989, Baum & Bailey 1990).

Nevertheless, the micromorphology of lemmas and paleas has not been widely studied in the Pooideae, except for some particular genera (*see* Thomasson 1986, Ortúñez Rubio & Fuente García 1995, Fuente García & Ortúñez Rubio 1996, among others). Accordingly, our objective was to carry out a micromorphological characterization of lemmas and paleas of *Bromus*, analyzing the value of their microcharacters for an infrageneric classification.

Bromus is the largest genus in the Bromeae. We estimate some 160 species worldwide for *Bromus*. They are either mesophytic or xerophytic, live in many different habitats (Watson & Dallwitz 1988), and are distributed nearly worldwide mainly in temperate zones of the Northern Hemisphere and on tropical mountains (Mabberly 1993). Bromeae is included in the Pooideae, one of the six subfamilies of the

Poaceae (Clayton & Renvoize 1986). Traditionally, the closest related species in *Bromus* have been grouped only on the basis of their morphological similarities, although recently, Acedo and Llamas (1999) have provided leaf anatomical data, allowing differentiation of some groups of species. The six subgenera of *Bromus* are: *Bromus*, *Stenobromus* Hackel, *Festucoides* Hackel, *Ceratochloa* (Beauv.) Hackel, *Neobromus* Shear and *Nevskiella* Krecz. & Vved.

Material and methods

The lemmas and paleas of 77 species (Table 1), representing the six subgenera of *Bromus*, were examined using herbarium specimens primarily from LEB. Initially, the 22 Iberian species were studied (Acedo & Llamas 1999), which belong to subgenera *Bromus*, *Stenobromus*, *Festucoides* and *Ceratochloa*. In most of these species, several specimens per population and several populations from several different locations were

Table 1. Studied species/total number of species in each subgenus of *Bromus*.

<i>Bromus</i> (29/40):	<i>B. adoensis</i> Hochst. ex Steud., <i>B. alopecuros</i> Poir., <i>B. arenarius</i> Labill., <i>B. arvensis</i> L., <i>B. brachystachys</i> Horn., <i>B. briziformis</i> Fisch. & Mey., <i>B. bromoideus</i> (Lej.) Crép., <i>B. cabrerensis</i> Acedo & Llamas, <i>B. caroli-henrici</i> Greuter, <i>B. chrysopogon</i> Viv., <i>B. danthroniae</i> Trin., <i>B. hordeaceus</i> L., <i>B. intermedius</i> Guss., <i>B. lanceolatus</i> Roth, <i>B. lepidus</i> Holmberg, <i>B. lusitanicus</i> Sales & P.M. Sm., <i>B. nervosus</i> Acedo & Llamas, <i>B. optimae</i> Scholz, <i>B. oxyodon</i> Schrenk, <i>B. pectinatus</i> Thunb., <i>B. psammofilus</i> P.M. Sm., <i>B. pseudothominii</i> P.M. Sm., <i>B. racemosus</i> L., <i>B. scoparius</i> L., <i>B. secalinus</i> L., <i>B. severtzovii</i> Regel, <i>B. squarrosum</i> L., <i>B. tythanthus</i> Newsky.
<i>Stenobromus</i> (6/6):	<i>B. diandrus</i> Roth, <i>B. fasciculatus</i> Presl, <i>B. matritensis</i> L., <i>B. rubens</i> L., <i>B. sterilis</i> L., <i>B. tectorum</i> L.
<i>Festucoides</i> (32/90):	<i>B. albidus</i> M. Bieb., <i>B. angustifolius</i> Jacq., <i>B. armenus</i> Boiss., <i>B. biebersteinii</i> Roem. & Scholz, <i>B. cappadocicus</i> Boiss. & Bal., <i>B. ciliatus</i> L., <i>B. cognatus</i> Steud., <i>B. condensatus</i> Hack., <i>B. erectus</i> Hudson, <i>B. exaltatus</i> Bernh., <i>B. firmior</i> Stapf., <i>B. frigidus</i> Boiss. & Hausskn. ex Boiss., <i>B. inermis</i> Leyss., <i>B. insignis</i> Buse, <i>B. kalmii</i> Gray, <i>B. kinabaluensis</i> (Jansen) Veldk., <i>B. kopetdagensis</i> Drobov, <i>B. mucroglumis</i> Wagnon, <i>B. natalensis</i> Stapf., <i>B. nottowayanus</i> Fernald., <i>B. pannonicus</i> Kummer & Send., <i>B. paulsenii</i> Hack. ex Paulsen, <i>B. pseudolaevipes</i> Wagnon, <i>B. pubescens</i> Muhl. ex Wild., <i>B. pumpelianus</i> Scribn., <i>B. ramosus</i> Hudson, <i>B. riparius</i> Rhem., <i>B. runssorensis</i> Schum., <i>B. speciosus</i> Ness., <i>B. timorensis</i> Veldk., <i>B. tomentellus</i> Boiss., <i>B. variegatus</i> M. Bieb.
<i>Ceratochloa</i> (8/25):	<i>B. arizonicus</i> (Shear) Stebbins, <i>B. bonariensis</i> Parodi & Camara, <i>B. breviaristatus</i> Bukl., <i>B. carinatus</i> Hock. & Arn., <i>B. catharticus</i> Valh., <i>B. lanatus</i> Kunth, <i>B. marginatus</i> Ness, <i>B. purgans</i> L.
<i>Neobromus</i> (1/1):	<i>B. berterianus</i> Dolla.
<i>Nevskiella</i> (1/1):	<i>B. gracillimus</i> Bunge.

analyzed in order to estimate intraspecific variation. An additional 55 species belonging to the previously mentioned subgenera, as well as those included in the subgenera *Neobromus* and *Nevskiyella* were also studied. As many specimens were studied as possible, within the limitations of the available material. The specimens were borrowed from the following herbaria: BCF, BM, BR, BRESA, C, COI, E, FCO, FI, G, GDA, GDAC, GH, GOET, JACA, K, L, LD, LINN, LISI, LISU, LY, MA, MAF, MICH, MO, MPU, MUB, NAP, NTM, OXF, P, PH, PRE, SALA, SALAF, SANT, SEV, UNEX, UPS, US, VAB, VALA, W, and Z. All of the identifications were confirmed using all relevant literature available.

Data were taken from the middle part of the side of the lemma, and from the middle part of the palea (Fig. 1). Observations were carried out first with a stereo microscope, a light microscope (LM), and finally a scanning electron microscope (SEM). Stains such as Sudan III were used to aid identification of cork cells using LM. Some samples were sonicated in xylene for at least thirty minutes to remove epicuticular waxes that can obscure surface features. For SEM observations, lemmas and paleas of the lowermost floret of mature spikelets were glued onto stubs and sputter-coated with 40 Å of gold-palladium under high vacuum. The samples were examined at 5–20 kV in a JEOL J.S.M. 6100 SEM.

We followed Ellis (1979) for the description of lemma micromorphology, since lemmas are homologous to leaves (Snow 1996), and we extended the terminology to paleas because they show similar epidermal characteristics. The only exception to Ellis' terminology is that referring to the crown cells (see below for definition and references).

Results

Description of micromorphological characters

Long cells

Long cells are the dominant element of the epidermis of lemmas and paleas, which, despite

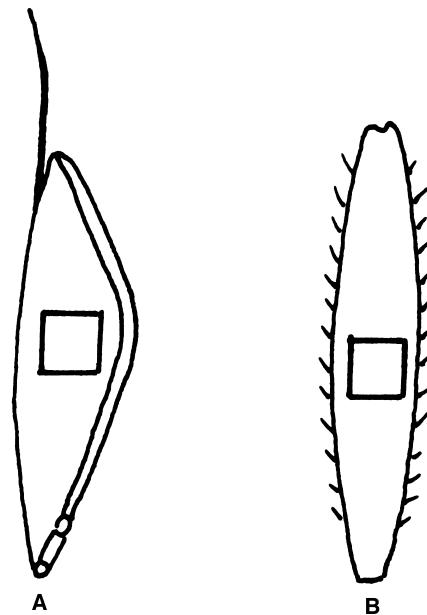


Fig. 1. Observation area. — **A:** Lemma. — **B:** Palea (abaxial surface).

their name, vary notably in length (Table 2 and Table 3). These are rectangular, long and narrow cells having convex periclinal walls. In contrast, the anticlinal walls are parallel and highly sinuous, with waves in shape of Ω. The ends are often angular, and their length ranges from 20 to 245 µm.

Short cells

Short cells are in general nearly equal-sided, although cells with a width exceeding the length are also relatively frequent (Tables 2 and 3). Short cells alternate with the long cells or occur solitary or in pairs. The frequency of short cells is resulting to some degree from the length of the long cells, and shows variations among several taxa. The anticlinal cell walls are straight or sinuous. Two types of short cells have been observed: cork cells and exodermic cells.

Cork cells

These are short cells containing solid deposits of organic substances and suberified cell walls

(Kaufmann *et al.* 1970). Their periclinal walls are usually slightly convex and collapse upon dehydration. The most frequent shape is polygonal or \pm square. In some cases they adopt different morphologies (semicircular, reniform) that are consistent in the several groups.

Exodermic cells

These cells protrude beyond the general level of the epidermis and have periclinal walls with differing degrees of convexity. Depending upon the size and shape of the process, three subtypes can be differentiated into:

— **Crown cells:** Short exodermic cells with small convex or conical protrusions, which occur frequently on the lemma and palea. This term was used by Prat (1932) and has been revived by Watson and Dallwitz (1988) in order to characterize the leaf epidermis of some genera of grasses. Many authors have neglected them, including all exodermic

cells under the name of macrohairs. Ellis (1979) referred to these cells as hooks, using the term as a synonym of crochets and crown cells, although he only represented figures with hooked protrusions, thereby excluding those with rounded protrusions. The protrusions of the periclinal wall of the crown cells vary in shape and size. In general they have crenulate walls, with pronounced waves similar to those of the long cells.

- **Prickles:** Exodermic cells with elongated tips and swollen bases arising directly from the epidermis. They occur frequently especially on the veins, although in some groups they occur between long cells with a comparable frequency. Valdes-Reyna and Hatch (1991) found a high silica concentration in this structure.
- **Macrohairs:** Unicellular structures, soft or rigid, often with a bulbous base. They occur between other epidermal cells, and, in some cases, they have an associated small cork cell.

Table 2. Micromorphological characters of lemma. Length in μm ; cell density = number per mm^2

Lemma	<i>Bromus</i>	<i>Stenobromus</i>	<i>Festucoides</i>	<i>Ceratochloa</i>	<i>Neobromus</i>	<i>Nevskiella</i>
Long cells						
length	50–150	35–100	30–90	20–150	50–100	30–130
anticlinal wall	sinuous	sinuous	sinuous	sinuous	sinuous	sinuous
periclinal wall	convex	convex	convex	convex	convex	convex
Short cells						
length	6–20	5–27	7–30	10–16	10–22	7–12
CORK CELLS						
shape	square	semicircular	reniform	square	reniform	reniform
density	40–320	40–500	40–460	100–320	400–500	340
length	6–7	5–9	7–13	10–13	10–13	7–9
EXODERMIC CELLS						
Crown cells						
cell shape	square	square	square	square	square	square
protrusion shape	spherical	spherical	spherical	pointed	pointed	pointed
density	240–920	280–1960	(5)40–1120	140–1300	1200–1500	900–1200
length	15–20	15–27	15–30	12–16	17–22	8–12
Prickles						
density	0–200	140–220	0–280	40–60(200)	0–40	200–240
Macrohairs						
type	soft	rigid	rigid	soft or rigid	rigid	rigid
density	0–400	0–80	0–300	0–200	0–240	0–20
length macrohair	40–600	50–400	100–1000	30–300	70–350	60–70

Epicuticular waxes

Epicuticular waxes are deposits, formed mainly by a large mixture of different chemical compounds.

Micromorphological description of lemmas and paleas in *Bromus*

Bromus subgen. *Bromus*

Lemma. Long cells 50–150 µm. Cork cells square, 6–7 µm long, density 40–320 cork cells mm⁻². Crown cells square, 15–20 µm long, with a spherical protrusion, density 240–920 crown cells mm⁻². Prickles absent or scarce, up to 200 prickles mm⁻², distributed mainly on the veins. Macrohairs highly variable in density, up to 400 macrohairs mm⁻², 40–600 µm long. Waxes absent, except for outer zones of the lemma. Fig. 2A.

Palea. Long cells 35–245 µm. Cork cells

square or polygonal, 15–21 µm long, density up to 360 cork cells mm⁻². Crown cells square to ovate, 10–35 µm long, with a spherical protrusion, density up to 640 crown cells mm⁻². Prickles absent. Macrohairs soft, with fine walls and bulbous base, highly variable in density, up to 120 macrohairs mm⁻², 50–160 µm long. Waxes absent. Fig. 3A.

Bromus subgen. *Stenobromus*

Lemma. Long cells 35–100 µm. Cork cells semicircular, 5–9 µm long, density 40–500 cork cells mm⁻². Crown cells square, 15–27 µm long, with a spherical protrusion, and density 280–1960 crown cells mm⁻². Prickles 140–220 per mm². Macrohairs absent or scarce, up to 80 macrohairs mm⁻², 50–400 µm long. Waxes absent. Fig. 2B.

Palea. Long cells 30–75 µm. Cork cells semicircular, 6–8 µm long, density 40–560 cork cells mm⁻². Crown cells rectangular, 13–18 µm

Table 3. Variation of micromorphological characters in palea. Length in µm; cell density = number per mm².

Palea	<i>Bromus</i>	<i>Stenobromus</i>	<i>Festucoides</i>	<i>Ceratochloa</i>	<i>Neobromus</i>	<i>Nevskiella</i>
Long cells						
length	35–245	30–75	30–200	30–125	65–160	130–180
anticlinal wall	sinuous	sinuous	sinuous	sinuous	sinuous	sinuous
periclinal wall	convex	convex	convex	convex	convex	convex
Short cells						
length	10–35	6–18	10–30	10–25	7–22	10–12
CORK CELLS						
shape	square, polygonal	semicircular	rounded- elliptic	rounded- elliptic	oblong	not observed
density	0–360	40–560	20–600	120–480	20–60	–
length	15–21	6–8	10–18	10–15	7–13	–
EXODERMIC CELLS						
<i>Crown cells</i>						
cell shape	square- ovate	rectangular	square- rectangular	square	rectangular	not observed
protrusion shape	spherical	pointed	pointed	pointed	pointed	–
density	0–640	210–1600	20–900	140–860	400–500	–
length	10–35	13–18	12–30	14–25	18–22	–
<i>Prickles</i>						
not observed	not observed	not observed	not observed	not observed	not observed	not observed
<i>Macrohairs</i>						
type	soft		rigid	soft		
density	0–120		0–400	60–100		
length macrohair	50–160		20–130	50–75(175)		

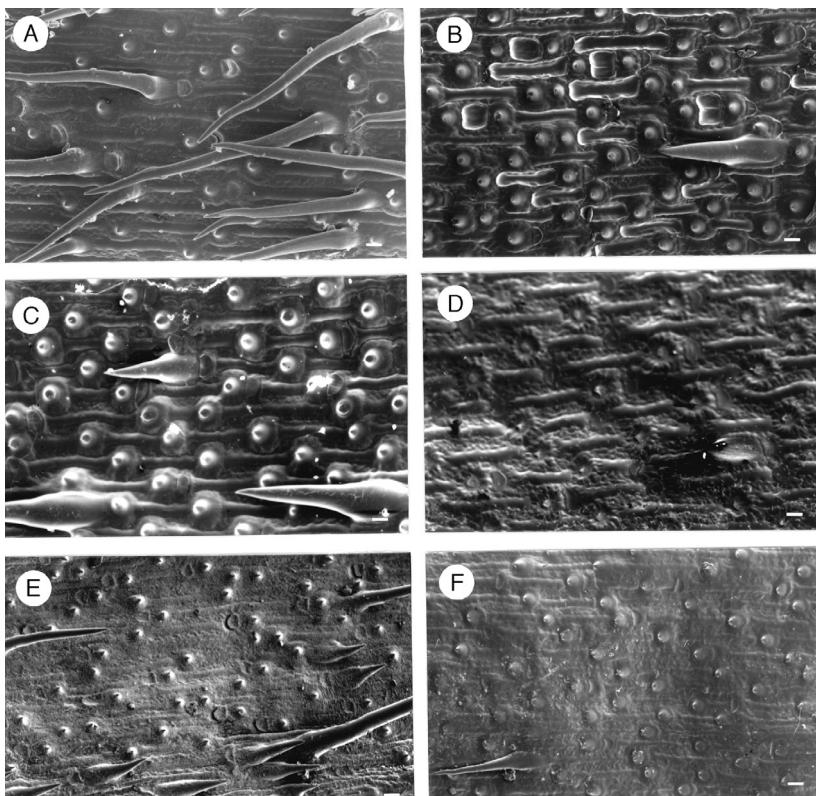


Fig. 2. Lemma surface.
— A: *Bromus tytthanthus*.
— B: *B. sterilis*. — C: *B. timorensis*. — D: *B. catharticus*. — E: *B. berterianus*. — F: *B. gracillimus*. Scale bar 10 μm .

long, with a pointed protrusion, density 210–1600 crown cells mm^{-2} . Prickles and macrohairs absent. Waxes absent. Fig. 3B.

Bromus subgen. *Festucoides*

Lemma. Long cells 30–90 μm . Cork cells reniform, 7–13 μm long, density 40–460 cork cells mm^{-2} . Crown cells square, 15–30 μm long, with a spherical protrusion, density 40–1120 crown cells mm^{-2} , seldom as few as five cells mm^{-2} . Prickles with an ovate base, density up to 280 prickles mm^{-2} . Macrohairs very variable in density, up to 300 macrohairs mm^{-2} , 100–1000 μm long. Waxes dense, covering almost all the surface. Fig. 2C.

Palea. Long cells 30–200 μm . Cork cells rounded-elliptic, 10–18 μm long, density 20–600 cork cells mm^{-2} . Crown cells square-rectangular, 12–30 μm long, with a pointed protrusion, density 20–900 crown cells mm^{-2} . Prickles absent. Macrohairs rigid, highly variable in density

up to 400 macrohairs mm^{-2} , 20–130 μm long. Waxes highly variable from absent to very abundant. Fig. 3C.

Bromus subgen. *Ceratochloa*

Lemma. Long cells 20–150 μm . Cork cells square, 10–13 μm long, density 100–320 cork cells mm^{-2} . Crown cells square, 12–16 μm long, with a pointed protrusion, density 140–1300 crown cells mm^{-2} . Prickles 40–60 per mm^2 , seldom up to 200. Macrohairs soft or rigid, highly variable in density up to 200 macrohairs mm^{-2} , 30–300 μm long. Waxes generally abundant. Fig. 2D.

Palea. Long cells 30–125 μm . Cork cells rounded-elliptic, 10–15 μm long, density 120–480 cork cells mm^{-2} . Crown cells square, 14–25 μm long, with a pointed protrusion, density 140–860 crown cells mm^{-2} . Prickles absent. Macrohairs soft, density 60–100 macrohairs mm^{-2} , 50–75 μm long. Waxes absent. Fig. 3D.

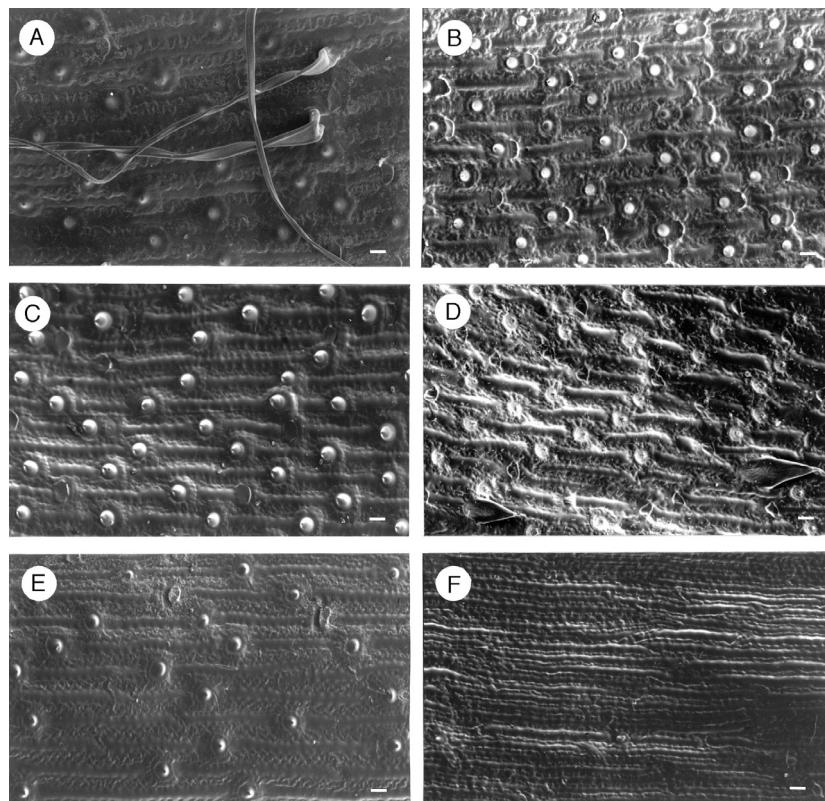


Fig. 3. Palea surface. —
A: *Bromus alopecuros*. —
B: *B. sterilis*. — **C:** *B. timorensis*. — **D:** *B. catharticus*. — **E:** *B. berterianus*. — **F:** *B. gracillimus*.
Scale bar 10 µm.

Bromus subgen. *Neobromus*

Lemma. Long cells 50–100 µm. Cork cells reniform, 10–13 µm long, density 400–500 cork cells mm⁻². Crown cells square, 17–22 µm long, with a pointed protrusion, density 1200–1500 crown cells mm⁻². Prickles up to 40 per mm². Macrohairs highly variable in density, up to 240 macrohairs mm⁻², 70–350 µm long. Waxes dense. Fig. 2E.

Palea. Long cells 65–160 µm. Cork cells oblong, 7–13 µm long, density 20–60 cork cells mm⁻². Crown cells rectangular, 18–22 µm long, with a pointed protrusion, density 400–500 crown cells mm⁻². Prickles and macrohairs absent. Waxes absent. Fig. 3E.

Bromus subgen. *Nevskiella*

Lemma. Long cells 30–130 µm. Cork cells reniform, 7–9 µm long, density 340 cork cells mm⁻². Crown cells square, 8–12 µm long, with a pointed

protrusion, density 900–1200 crown cells mm⁻². Prickles absent or scarce, 200–240 prickles mm⁻². Macrohairs absent or scarce, up to 20 macrohairs mm⁻², 60–70 µm long. Waxes absent or very scarce. Fig. 2F.

Palea. Long cells 130–180 µm. Cork cells absent. Crown cells generally absent in most of the palea, although sometimes a few crown cells can be found near the base or beside the keel. Prickles and macrohairs absent. Waxes absent. Fig. 3F.

Discussion and conclusions

One of the main problems with micromorphological studies is to determine the optimal site of observation. We have noticed variation in lemmas and paleas depending on the zone on observation. The lemma shows variations in the expression of micromorphological characters among the apex, base and margins, whereas the palea varies mostly near the apex and in some

cases in the wings. Because of this, we have selected the middle part of the lemma side, and the central part of the palea body (Fig. 1). These regions appear to be quite constant in their micromorphological characteristics among the taxa studied.

The analysis of the epidermal micromorphology of lemma and palea of *Bromus* reveals considerable variation in some subgenera, particularly regarding the presence of waxes and density of short cells. Several differences also exist as regards the presence or absence of macrohairs, but in some species this character can show a high variation even among specimens from the same population.

On the basis of the epidermal micromorphology of lemma and palea, it can be inferred that all species of the genus *Bromus* have the same epidermal cell types, as it can be expected in species of the same genus. It means that it is possible to identify a fragment of a floral bract as *Bromus*, or one of its subgenera, using the micromorphological characteristics presented here. Nevertheless, the similarities among species of the same subgenus, or lower taxonomic groups, are higher than those among species of different subgenera. The most different one is the subgenus *Nevskiella*. If these differences were as high as those on the other genera of *Bromeae* (*Boissiera* and *Littledalea*) this could support its segregation from *Bromus* as proposed by Kreczetowicz and Vvedensky (1934).

There are two characters that, in general, have a close positive correlation. These are the length of the long cells (in lemma and palea) and the number of short cells. For example, subgenus *Stenobromus* has the shortest short cells (5 µm) and the highest density of short cells (ca. 2000, up to 560 cork cells mm⁻² and up to 1600 crown cells mm⁻²). However, as the long cells increase in length, the density of short cells decreases. An extreme case is found in the palea of the subgenus *Nevskiella*, where long cells are relatively long, and short cells are lacking entirely. The lowest values for lemmatal long cells size are found in subgenus *Ceratochloa*, where longer long cells also occur, thereby making long cells size not useful for characterizing the epidermal type.

More differences occur on short cells than

long cells. Thus, the crown cells show considerable variation in density and in their type of protrusions, whereas cork cells show variation in their size and density (Table 2).

The exodermic cells, like the cork cells, are distributed in longitudinal arrays, alternating with the long cells and other epidermal cells.

The density of cork cells, in relation to the other epidermal elements is also variable, ranging from 0–600 cells mm⁻². In a given subgenus the number of cork cells is higher in the palea than in lemma, except in the subgenus *Nevskiella*, where this type of cell has not been observed on the palea. The highest values for the length of cork cells are found on the palea of subgenera *Festucoides* and *Stenobromus*. The size of the cork cells varies from 5–13 µm in the lemma and 6–21 µm in the palea. The highest values are found in the palea of subgenus *Bromus*, where cork cells are between 15–21 µm. (Table 3).

In *Bromus* all the exodermic cells are short cells. Snow (1996) reported the first evidence of macrohairs originated in short cells in *Eleusininae*. We have observed the same thing in *Bromus* where all the exodermic cells are short cells.

Crown cells vary from square to rectangular, although sometimes they have an irregular shape, as in the palea of subgenus *Stenobromus*. The protrusion or protuberance can be spherical or pointed and vary in its relative size to the cell supporting it. It can be either similar or slightly shorter than the cell, or of 1/3 or less of the diameter of the cell. In the first case the protrusion is formed by most of the external periclinal wall of the cell, or at least more than 1/3. In the second case the protrusion is formed only by the central portion of the outer periclinal cell wall (palea of *Bromus mucroglumis*, subgen. *Festucoides*).

The density of the crown cells varies notably among and within different subgenera. In the palea they vary from complete absence (subgenus *Nevskiella*) to 1600 cells mm⁻² (subgenus *Stenobromus*), where these differences are more significant. In the lemma their density always exceeds that of the palea, in species as well as in subgenera. It ranges from 40 cells mm⁻² in subgenus *Festucoides* to 1960 cells mm⁻² in *Stenobromus*.

The size of the crown cells ranges from 8 to

30 µm. Subgenus *Nevskiella* has the smallest (8–12 µm), whilst in the remaining subgenera they are 12–30 µm.

Intercostal prickles appear in variable densities, but differences are not significant, since in most cases there is high intraspecific variation. Sometimes prickles are longer than usual, and then, it is difficult to differentiate between prickles and macrohairs. This intergradation has been formerly reported by Metcalfe (1960), Ellis (1979) and Snow (1996).

Snow (1998) pointed out that these characters showing intergradation cannot be considered homologous and hence lack phylogenetic value although they may still be of local diagnostic value. The *Bromus ramosus* group (rank not defined yet, perhaps section) has macrohairs only on a marginal strip of the lemma. In this case the character is diagnostic for this group against the remaining species of the subgenus.

Since the density of macrohairs on the lemma and palea is highly variable, their presence or absence has little taxonomic significance, which confirms our experience in the genus. One exception is the disposition of the macrohairs in subgenus *Festucoides*, where many species have macrohairs restricted to the margin of the lemma. Even if they appear over much of the surface, their density in subgenus *Festucoides* increases notably in the marginal zone.

It is known that the trichomes are common in plants growing in xeric habitats (Esau 1977). We agree with Dávila and Clark (1990) that papillae, prickles and waxes could play a similar role, and, as pointed out by Stace (1965), may reduce the transpiration rate of the plant organ having them. However, we did not find a positive correlation between the density of these structures and the habitat of origin of the specimens.

The interest of the epicuticular waxes resides, according to some authors, in the fact that the configuration of the deposits of wax seems to have a genetic basis (Baum & Hadland 1975). The study of the waxes in several species could therefore be justified for taxonomic purposes. Taking this premise as valid, the epicuticular waxes have been studied in numerous taxa and for several purposes. We tried to analyze only the intrageneric variation, with the aim of knowing if this character has differential value in the

systematic of the genus *Bromus*.

The epicuticular waxes are common on the lemma, where they often appear in the zone not covered by the glume or the lemma of the lower floret. Waxes were not observed on the lemmas of the two annual subgenera *Stenobromus* and *Nevskiella*. In the palea, waxes are only frequent in some species of subgenus *Festucoides*.

Considering the studied samples, we can infer that variation in epicuticular waxes in *Bromus* (when present) is not significant. In the lemma, the absence or presence of waxes depends on the zone of observation, i.e. if it is covered or not by another bract (perhaps, there is some relationship between the presence of epicuticular waxes and the environmental conditions). The morphology of the crystals is similar in all the studied species (and therefore homogeneous in the six subgenera) being rosettes of platelets in all cases (nomenclature following Barthlott *et al.* 1998). In the palea we have only observed epicuticular waxes in subgenus *Festucoides*, although they are also lacking in some species (*B. erectus*, *B. paulsenii*, *B. pseudolaevisipes*, *B. ramosus*, *B. speciosus*, *B. timorensis*).

Thus we can remark the following conclusions: For the micromorphological studies it is necessary to make a careful selection of the site for its observation. The palea reveals itself as an important place of observation in this type of survey. The genus *Bromus*, or any of the six subgenera, can be identified using the micromorphological characteristics of palea and/or lemma.

It should be interesting to study the closely related genera in the Bromae and to compare them with the differences among the subgenera. In such a way the segregation of *Nevskiella* could be justified as an independent genus, because it is the most different one in *Bromus*. This would justify the proposal made by Kreczetovich and Vvedensky (1934).

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Appendix: Specimens examined

Bromus adoensis. France. Sementes vindas do H13 Bordeaux; Cult. na EAN, sob o n° 15749, 1947 B. Rainha (MA 284889). Germany. In Karlsruhe i. Baden kultiviert, 1909–1912, 117 m, A. Kneucker (MA 246377).

Bromus albidus. Russian Federation. An steinigen Abhängen in den Landschaften Digoria und Ossetia im Kaukasus, 1500 m, 1899 B. Markowicz (MA 246378).

Bromus alopecuroides. Greece. Rhodos, 1 km NE Salakos; Cultivated area, 1982 A. Carlström 5633 (LD). Samos. N of the town of Samos, roadside and former terraced fields, 50–100 m, 1992 S. & B. Snogerup 8832 (LD). Portugal. Alcoutim, Alamo, Rio Guadiana (Algarve), vegetação margem do rio, solo derivado de xisto, 29SPB34, 1983 M. Lousa & P. Oliveira (LISI s.n.). Alvaiazere, W da localidade de Bofinho (Beira Litoral), roca calcarea, 160 m, 29SNE51, 1970 M. Lousa (LISI 1411).

Bromus angustifolius. No data (C).

Bromus arenarius. Australia. Albury, 1916 T.H. Patterson NSW 10596 (K). New South Wales, hill E of the road from Cobar to Bourke, c. 6 km bifurcation Byrock., low-growing wood of *Eucalyptus*, *Acacia*, and grassy clearings, 200–250 m, 1988 W. Greuter, *Plants from NSW, AT, 20774* (K).

Bromus arizonicus. United States. Arizona, St. Johns Mission, next to the house Maricopa County, 1200 ft, 1961 R. J. Kramer 40 (K).

Bromus armenius. Turkey. Prov. Tunceli, Munzur dag above Ovacik, rocky limestone slopes, 2800 m, 1957 Davis & Hedge 31284 (K).

Bromus arvensis. Bulgaria. Prov. Burgas, ca. 23 km from Burgas, Aheloj-Kables Kovo, weet at side of maize field, 50 m, 35TNH5123, 1988 S. L. Jorg & S. P. Thoraton Wood 9555 (MA 452611). Norway. Bergen, Wiesbaden auf dem Bingesr., 1902 Dr. A. Schultz (MAF 26558). Spain. Cuadros (León), Borde de Camino, 30TTN83, 1988 F. Llamas (LEB 52598). Huesca, Valle de Hecho, Siresa-Oza, Gargantas del Infierno, 980 m, 30TXN8641, 1965 P. Montserrat (JACA 91665).

Bromus berterianus. Bolivia. Larecaja, Sorata, 2650 m, 1860 Mandon (W 35397). Chile. Masafuera, 1857 C. Leyb. Desv. (W 39793, Herb. E. Hackel). Valdivia, Fr. Philippi (W 7855, Herb. E. Hackel). Mexico. Lower California, 1889 Dr. E. Palmer (W 7830, Herb. E. Hackel). Mojave Desert, S.E. base of Old Dad-Ganite Mt., sun, dry fine gravel in large washes, 24 in., 1941 C. B. Wolf (W 24764). Peru. Dept. de la Libertad, Santiago de Chuca, 3000 m, L. Raimondi (W s.n.). United States. California, summit of Mustang Grade, Monterey County, 2700 ft, 1957 J. T. Howell 32106 (W 24411). Missouri, MO Bot. Garden, 1904 J. Kellogg (PH 605550).

Bromus biebersteinii. Afghanistan. Kabul, in declivibus

australibus jugi Salang, substr.gran., 3200 m, 1967, H. K. Rechinger 37092 (MA 416864).

Bromus bonariensis. Argentina. Prov. de Buenos Aires-Pdo. de Tornquist-Sa. de la Ventana, Abra de la Ventana, -78, Proyecto Ventania, M. A. Torras (K).

Bromus brachystachys. Afghanistan. Kabul, Guzar Gah., 1950 H. F. Neubauer (W 2451). Germany. Aschersleben, Silesia, Von Hornung (C, conf. H. C. Watson). Proving Sachaen, unter den Westerbergen, Aschersleben selten, 1905 Prof. Preusse (W 9362). Iraq. Khalis, 1960 R. Whule Haines (W 11955).

Bromus breviristatus. United States. Arizona, Santa Cruz Valley, Tucson, 1884 C. G. Pringle (PH 25786). Bozeman, uplands, 1905 J. W. Blankinship 593 (PH 519769).

Bromus briziformis. Germany. Bei Gesain Thünungen (Rhön), kultiviert, 1908 M. Goldschmidt (MA 246388, ex. herb. B). Erpert, An Ackerrandern in Loberfelte, 1894 C. Reineche (MA 13795).

Bromus bromoideus. Belgium. 1823 Lejeune (holotype?). 1963 R. Tournay (BR). Liège, cultures experimentales, 1971 P. Auquier & W. Bellotte (MAF 95554).

Bromus cabrerensis. Spain. Campus de Vegazana (León), Zona nitrificada, 30TTN81, 1994 C. Acedo (LEB 51652). Truchas (La Cabrera, León), borde de reguero con *Holcus lanatus*, *Poa trivialis* ..., 29TQG07, 1992 F. Llamas & C. Acedo (LEB 52595).

Bromus cappadocicus. Bulgaria. Balkan occid., Sophia, Kamiko supra vic Belidiehan, in pascuis saxosis, solo calcario, 1954 V. Velcev (MA 183121).

Bromus carinatus. Costa Rica. Alajuela, Volcán Poás-Vara Blanca, grassy roadsides 1968 R. W. Pohl, G. Davidse & N. S. F. Grant (K). Mexico. SE de San Pedro Nexapa, a 2 km del pueblo, 2650 m, 1964 Marina Villegas 453 (MA 284884). United States. California, San Francisco, Twin Peaks, very common on open S-facing slope, 1985 R. A. Norris 4823 (MA 376369). California, Hamilton Field-Ignacio, Marin Co., 1945 J. T. Howell (K).

Bromus caroli-henrici. Greece. Crete, Eparchie Rethimni, Tal und Schlucht N von Methoia, Kalkfelshänge, 180 m, 1982 W. Greuter & H. Risse 19456 (LD).

Bromus catharticus. Portugal. Coimbra, 29TNE45, 1879 A. Möller (LISU-P/6124). Spain. Puerto de la Cruz, 1983 E. Hernández & T. E. Díaz Gonzalez (LEB 24728). Sagunto (Valencia), 30SYJ39, 1979 no collector (LEB 52620). Torno, Onón, Naturalizado en lugares nitrificados, 20 m, 30TUP0619, 1992 F. Llamas, R. Mª Valencia & C. Acedo (LEB 52564). United States. Columbia, Dr. Schouler (E).

Bromus chrysopogon. Cyprus. Entre Xylophagou et Ayia Thékla (Larnaca), Cham (blé) abandonné et pseudosteppe

à Sarcopote, 0–5 m, 1991 *Iter Mediterraneum IV (Chipre) 104* (MA 496422). Larnaca, lac salé, 1991 *Iter Mediterraneum IV (Chipre) 197* (MA 495585).

Bromus ciliatus. **Canada.** Québec, St-Rémi d'Amherst, Comté de Papineau, 1954 G. Lamarre (MA 166162-2). **United States.** Auf flachem Ufergelände längs des Susquehanna-Flusses bei Sayre, in Pennsylvania, 1901 W. C. Barbour (MA 246389).

Bromus cognatus. **Kenya.** Tanganjica, Rongai, Kilimandscharo Seite 2400 m, 1934 H. J. Schlieben (MA 175639). Tanganjica, Kilimandscharo, Güterwald(obere Grenze), 1934 H. J. Schlieben (MA 175640).

Bromus condensatus. **Austria.** Tirolia australis, Bozen, ad saxa porphyrica monti Kalvarienberg, -98 Sauter (C).

Bromus danthoniae. **Iran.** Gotvend (Rio Karvum), 400 m, C. Pau & C. Vicioso (MA 14042). No data (Willkomm-COI).

Bromus diandrus. **France.** Mauguio (Dép. de l'Hérault), sables littoraux, Carac, ammophiletalia, 1977 A. Dubuis (MAF 105093). **Portugal.** Aveiro, Oliveira do Bairro, estrada Ilhavo-Vagos-P (Beira Litoral), solo arenoso, cultivado (Vinha), 1977 A. Marques (HVR 438). Meco (Tras-os-Montes), Ruderal viaria em areias e em mata-gais de Artemisi, 1995 M. Sequeira & L. Crespi (HVR 4632). Setubal, Mouriscos, 29SNC06, 1953 J. C. Sousa Velasco (LISI s.n.). **Spain.** Chiclana de la Frontera, Arenas viarias, 29SQA53, 1967 J. Borja Carbonell (MAF 69130). Fazouro, Borde de camino, 15 m, 29TPJ3628, 1992 F. Llamas, R. M^a Valencia & C. Acedo (LEB 52569). Puerto Lope (Granada), 700 m, 30SVG2826, 1984 Aroza & Socoro (GDA 17030).

Bromus erectus. **France.** Bayonne, cerca de Grignan, 1992 R. M^a Valencia Barrera (LEB 52523). **Spain.** Peñalba de Santiago (Bierzo, León), pastizal sobre roca calcárea, 29TQH00, 1989 F. Llamas & C. Acedo (LEB 52535). Puerto de Tarna (León), pastizales sobre sustrato calizo, 1490 m, 30TUN17, 1989 F. Llamas & C. Acedo (LEB 52534). Puerto Ventana (León), 1520 m, 29TQH47, 1989 F. Llamas & C. Acedo (LEB 51519). **United Kingdom.** Oxfordshire, Oxford, University parks, 1942 C. E. Hubbard (K).

Bromus exaltatus. **France.** Paris, cogida en la siembra, 1850 no collector (MA 246405, MA 246405-D).

Bromus fasciculatus. **Egypt.** Alexandrien, 1876 Ernest Sickenberger, *Thellung* (Z). **Greece.** Crète, Archánes, sud de Knossós, sur débris de rochers, 1982 G. Van Buggenhout (MAF 126913). **Morocco.** Marsa Saguiria, lithorrhaphaceo, in saxosis calc., 50 m, 1927 Dr. Font Quer (BCF 2351).

Bromus firmior. **Lesotho.** Maluti Mt., 6 km from Oxbow Lodge on rd. to Mokhotlo, aspect W, poorly-drained clay soil, 2700 m, 1987 M. D. Panagos 48 (PRE 711567).

South Africa. Natal, Mount Aux Sources, Sentinel Path,

common in grasslands above 8000', 1955 D. Edwards 651 (PRE 24359). Wakkerstroom, Oshoek, protected veld at vlei edgas, 1981 N. J. Devenish 777 (PRE 107034).

Bromus frigidus. **Iran.** Zardeh Kuh Mts., Bakhtiari, dry limestone rock, 12 000 ft, 1965 R. Timmis 119 (K).

Bromus gracillimus. **Afghanistan.** Sirotaí, in forest, 10000 ft, 1937 W. Koelz 11954 (K). **Iran.** Kerman, Mashig-Khaneh Soreh, 1948 Rechenjar, Aellen & Efandiari (K). **Russian Federation.** Mair, 1956 [unreadable] (K). **Tadzhikistan.** Montes Meridionales, Pamir, in valle fl. Pamir pr. Kosch-dshil, 1927 Rajkova (MA 14053 det. Nevski).

Bromus hordeaceus. **Portugal.** Miranda do Douro, vertente para o Fresno (Tras-os-Montes), em tufos em terreno granítico, 29TQF29, 1944 G. Barbosa & F. Garça (LISI 6481). Pinhaes, Alferrarede (Ribateixo), 29SND79, 1910 R. Palinha, H. Navel & F. Mendes (LISU-P/6137). Vila Nova de Fozcoa, proximidades, estrada V. V. D (Tras-os-Montes), Marginando moitas de herbetum, 1942 G. Barbosa & M. Myre (HVR 436). **Spain.** El Caurel, Villasibil (Lugo), 1980 J. Izco, J. Amigo & J. Guitián (SANT 15583). Llombera (León), borde de camino, 30TTN84, 1992 F. Llamas & C. Acedo (LEB 52356). Santibañez el Alto, Barrio de la Calzada (Cáceres), pastizales de diente, 29TQE05, 1982 A. Valdés Franzi (SALAF 21116).

Bromus inermis. **France.** Geishouse, massif du Grand-Ballon dép. Haut-Rhin, bords d'une rue, sur le graniteb 660 m, 1982 V. Rastetter (MAF 136912). **Germany.** Würzburg in Bayern, auf Kalkhaltigem Alluvium an Ackerrändern südöst., 1903 L. Gross (K).

Bromus insignis. **Malaysia.** Java, Marakkan, Junghuhn s.n. (L det. J. F. Veldkamp).

Bromus intermedius. **Spain.** Santa Cruz de Mudela (Ciudad Real), borde de camino en la Encomienda, 30SVH57, 1951 J. Borja (MA 199370). Sierra de Cábulas (Granada), 30SVF37, 1980 A. T. Romero (GDAC 24303). **Germany.** In Karlsruhe i. Baden, kultiviert, 117 m, 1903 A. Kneucker (MA 246436).

Bromus kalmi. **United States.** Indiana, nr. Clark Road in NW Gary Lake County, in sandy swale between low dune beach ridges, 1955 H. R. Bennett (MA 167409-D).

Bromus kinabaluensis. **Malaysia.** Borneo, Mount Kinabalu, 6000–13 500 ft, 1931–32 J. & M. S. Clemens 29174 (L).

Bromus kopetdagensis. **Turkmenistan.** Turkmeniya, zentralnii Kopet-dag, 1953 V. V. Nikitin (MA 364670).

Bromus lanatus. **Colombia.** Andes Cordillera Central, Dep. Tolima, Nevado del Tolima, arenas, 1932 J. Cuatrecasas (MA 175644).

Bromus lanceolatus. **Italy.** Naples, Genijoala Porrentina,

terminis nei germinalis, 1913 without collector (MA 14040). **Morocco.** Sidi Allal Lebsar, cultures irriguées, 1500 m, 1948 J. Vindt (SANT 7444). **Spain.** Aibar-Olatz (Navarra), pasto sobre cultivos abandonados, 760 m, 30TXN3419, 1986 A. Izpuru & P. Catalán (MA 364828). Cervera, 3 km al W (Lérida), 31TCG51, 1981 J. A. Devesa, T. Luque, & C. Romero (SEV 97261). Guareña (Badajoz), 29SQD50, 1988 T. Ruiz & M. C. Viera (UNEX 8460).

Bromus lepidus. **France.** Millau (Dep. Aveyron), Culture, 1985 C. Bernard & G. Fabre (MA 367337). **Spain.** Sanabria (Zamora), 1944 Losa & Vieitez (SANT 2415). **United Kingdom.** Cothill, Berks, 1933 J. Thapple (MAF 26595, MAF 26595).

Bromus lusitanicus. **Portugal.** Souzelas, near Coimbra (Beira Litoral), growing in damp soil between a road and a stream, 70 m, 29TNE45, 1983 F. Sales (E 146/93 2).

Bromus marginatus. **United States.** Moist meadow in open valley above Crazy Head Spring 8 miles E of Lame Deer, Rosebud County, Montana, 4300 ft, 1955 H. R. Benett (MA 167407).

Bromus matritensis. **Spain.** Puzol (Valencia), en terreno labrado, 30TYK37, 1982 J. L. Carretero (VALA 4109). Sorriba (León), 30TUN23, 1988 F. Llamas & C. Acedo (LEB 52604).

Bromus mucroglumis. **United States.** Grown at the Botanical Gardens, University of Michigan, 1948 H. K. Wagnon 1520 (MICH).

Bromus natalensis. **South Africa.** Natal, Escourt, Tabam-hlope mountain, 5–6000', 1939 O. West 1380 (PRE 24340). Transvaal, Oshoek. Wakkerstroom Distr. Vleis., often in muddy wet sites, 6400', 1974 N. J. Devenish 1520 (PRE 92069).

Bromus nervosus. **Portugal.** Reguengos de Monsaraz (Estremadura), 29SPC25, 1949 J. Tapum (LISI s.n.).

Bromus nottowayanus. **United States.** Virginia, Sussex County, Nottoway River, east of Huske, bottomland woods, 1940 M. L. Fernald & Bayard 12239 (GH 23249), 12239 (GH 23248).

Bromus optimae. **Cyprus.** Chipre, Cape Greco (Larnaca), calcaires coralliens du Miocène in serieux et sabl., 10–20 m, 1991 (MA 495913, Iter Mediterraneum IV, Chipre 157). Larnaca, lac salé, ± 5 m, 1991 H. Scholz (MA 495684, Iter Mediterraneum IV).

Bromus oxyodon. **Tadzhikistan.** Montes meridionales, Pamir-alaj exterior, ad declibia herbosa montium Turkestanicorum, 1927 Drobov (MA 14038).

Bromus pannonicus. **Belgium.** Montagne d'Auronse, bas de la combe d'Auronse, a l'Est du Roc des Hirondelles, 1650 m, 1991 M. Kerguelen (MA 526523).

Bromus paulsenii. **Tadzhikistan.** Monte Pamir, prope

Iacus Jashilkul, 3900 m, 1998 O. Paulsen 1108 (C).

Bromus pectinatus. **India.** Jambatri, 4000 ft, Surq. Li-Harrin, 1895 T.A. Cope 16825 (K). Lahul, Manali, 1941 N. L. Bor (K).

Bromus psammophilus. **Turkey.** İçel, Tarsus, Dunes, 1973 T. Zulú (E).

Bromus pseudolaevipes. **United States.** Grown at the Botanical Gardens, University of Michigan, 2850 ft, 1948 H. K. Wagnon 1507 (MICH).

Bromus pseudothominii. **Spain.** Muñorodero (Santander), Borde de carretera, 5 m, 30TUP80, 1992 F. Llamas, R. M^a Valencia & C. Acedo (LEB 52562). **United Kingdom.** Haddington, Aberlady, sandy grass, VC82, 1984 O. M. Stewart 037684 & P. M. Smith (E).

Bromus pubescens. **United States.** Muhlenberg Herbarium, "M. 154"(PH).

Bromus pumpellianus. **Alaska.** Seward highway about 18 miles from Anchorage, steep hillside meadow, 1965 M. Williams 1080 (K). **Russian Federation.** Prov. Magadan, regio Tschukotsky, distr. Bilibinsky, montes Anjui, fontes fl. Pogyden, terrassa rivulis, Jagodny, 1974 T. Koroleva & I. Brysgalova (K). Zapadnaia Chucotka, Anjuiskoe Nagore, Leboderezhe, r. Malvi Anjui, r. Topolepka, 1976 B. B. Petroskij & T. M. Koroleva (K).

Bromus purgans. **United States.** Felsiger Waldran, Cabin John, Maryland, 1907 A. Chase (MA 247117).

Bromus racemosus. **Norway.** Welgesheim-Horn-Napoleoshöne-Sprendlingen, 1905 Touton (MAF 26681). **Portugal.** San Pedro do Sul, Arcozelo (Beira Litoral), solo granodiorítico, 240 m, 29TNF60, 1980 J. Franco & M. Louza (LISI 6202). **Spain.** Albeos, Creciente (Pontevedra), Mala hierba en viñedos, 1987 M. Buján (SANT 21963). Cuadros (León), borde de camino, 30TTN83, 1988 F. Llamas (LEB 52613). Llombera (León), borde de regato con *Juncus inflexus*, 30TTN8746, 1992 F. Llamas & C. Acedo (LEB 52639).

Bromus ramosus. **Spain.** Hervás (Cáceres), 30TTK56, 1975 Bote, Ladero & Pérez Chiscano (MAF 93281). Posada de Valdeón (León), borde de cursos de agua, 30TUN47, 1968 J. Borja (MAF 76329). **United Kingdom.** Dorsetshire, Corfe Castle, 1944 A. Dunston (K).

Bromus riparius. **Russian Federation.** Prov. Kursk, reservatio "Streletskai Step", 1968 V. V. Makarov (MA 284800). **Yugoslavia.** Serbia, in subalpinis Mte. Bassora, 1903 Bierbach (MA 14052).

Bromus rubens. **Spain.** Castropepe (Zamora), Comunidades nitrófilas de plantas anuales, 30TTM75, 1988 J. M. Fernández (LEB 51013). León, Campus Universitario de Vegazana (León), comunidades nitrófilas de plantas anuales, 1994 C. Acedo (LEB 52457). Sierra de Mahimon (Almería), borde de camino, 1250 m, 30SWG77, 1986 F. Gómez (LEB 31787).

Bromus runssorensis. **Kenya**. Tanganjika, Kilimandscharo, Gürtelwald, 1620 m.ü.M., 1934 *H. J. Schlieben* 4532 (MA 175638-2). Tanganyika Terr. near Mbeye, belns Mporoto, Porto Mts., 6200 ft, 1932 *S. W. St. Blair-Thomson* (MA 386787).

Bromus sclerophyllus. (G-BOISS).

Bromus scoparius. **Portugal**. Bragança. Quinta de Sta. Apolonia (Tras-os-Montes), comunidade de terófitos, rochas ultrabásicas, 1993 *C. Aguiar* (BRESA 2567). **Spain**. Atarfe, vega del Genil (Granada), borde de camino, 30SVG32, 1980 *Ladero & J. Hurtado* (MAF 109075). Peraleda de la Mata (Cáceres), pastos sobre calizas del mioceno, 30STK81, 1984 *Ladero, Valle & Ruiz* (SALAF 10380).

Bromus secalinus. **Belgium**. Moissons, Limerlé (pr. du Luxemb.), 1918 *A. Dolisy & R. Tournay* 16-1-1963 (BR s.n., “B. pr. gro. var. gro. f. gro”). **France**. Bellerive-sur-Allier, champ cultivé, 260 m, 1983 *R. Deschâtres* (SALAF 18531). **Norway**. Bergen, 1925 *Touton* (MAF 26716).

Bromus severtzovii. **Turkmenistan**. Turkmeniya. Ashabad-skii Raion, Vostochni Kopetdag Shamli, Na blazhnix mestah po beregu reki Sherloh, 1975 *I. A. Ivanov* (MA 247149). **Uzbekistan**. Montes Meridionales, Sogdiano-transoxani, in cultis prope urbem Katta-Kurgan, 1925 *Jakimova* (MA 13802).

Bromus speciosus. **South Africa**. Eastern Cape, 3226 DB, Amatole Mountains, below Gaikas Kop, waterfall marsh, confluence of streams, ± 200 m N, 1440 m, 1982 *H.D. Furness & P. B. Phillipson* 132 (PRE 692921). Natal, Underberg distr., 2929 CB, Cobham Forest Reserve, lakes, tufted in stream valley grassland, 7000 ft, 1982 *J. Manning, O. M. Hilliard & B. L. Burtt* 15977 (PRE 636321).

Bromus squarrosum. **Spain**. Cistierna (León), sobre rocas calcáreas, 30TUN24, 1988 *F. Llamas & C. Acedo* (LEB 52582). Embalse del Porma (León), repisas de rocas

calcáreas, 30TUN15, 1994 *F. Llamas* (LEB 50980). **Switzerland**. Niesch-Brieg (Wallis), 1869 *E. Siechenberges* (MAF 26722).

Bromus sterilis. **Spain**. Fresnedo (León), 29TPG98, 1988 *F. Llamas & C. Acedo* (LEB 52585). Lebeña (Santander), borde de carretera, 180 m, 30TUN78, 1991 *F. Llamas & C. Acedo* (LEB 51065). Sierra de Parapanda (Granada), Ruderal Viaria, 30SVG30, 1980 *A. T. Romero & C. Morales* (GDAC 24179).

Bromus tectorum. **France**. Nantes, 1879 without collector (MAF 26764). **Spain**. Cazorla, c. Ermita de la Virgen de la Cabeza (Jaen), en rellanos terrosos, 30SWG0096, 1983 *A. M. Hernández* (GDA 15411). León, Campus Universitario de Vegazana (León), comunidades nitrófilas de plantas anuales, 30TTN81, 1994 *C. Acedo* (LEB 52455).

Bromus timorensis. **Indonesia**. Timor, Mt. Tatamailau, in eucalypt forest, 2800 m, 1954 *Van Steenis* 18467 (L).

Bromus tomentellus. **Iran**. Kurdistania, (Assyria orient.) in montis Kuh-Sefin, (ditionis Erbil) regione superiore, 1600, 1893 *J. Bornmüller* (MA 14051). Mazanderan, in valle fluvii Calus, Pol-e Zanguleh, in saxosis schist. 2200–2600 m, 1948 without collector (K). 1963 *H. & F. Rechinger, Bor* (MA 418628).

Bromus turkestanicus. **Uzbekistan**. Montes Meridionales, Tadzhikorum, ad declivia saxosa (inter rupes)montium Tschulbair, ca. 3600, 1930, *Botschantzev & Vvedensky* (MA 13358).

Bromus tytthanthus. **Uzbekistan**. Montes Meridionales, Tian-schan occidentalis, inter rupes graníticas in montibus Mongol-tau prope, 1924 *Popov & Vvedennsky* (MA 14044).

Bromus variegatus. **Lebanon**. Auf den Gipfeln und obersten Abhängen des Dschebel Sânnin im, Libenom in Syrien, Kreidekal 2300–2600 m, 1903 *E. Hartmann* (MA 247178).