



Phylogenetic climatic niche conservatism and evolution of climatic suitability in Neotropical Angraecinae (Vandae, Orchidaceae) and their closest African relatives

Marta Kolanowska^{1,2}, Elżbieta Grochocka^{1,†} and Kamil Konowalik³

¹ Department of Plant Taxonomy and Nature Conservation, University of Gdańsk, Gdańsk, Poland

² Department of Biodiversity Research, Global Change Research Institute AS CR, Brno, Czech Republic

³ Institute of Biology, Wrocław University of Environmental and Life Sciences, Wrocław, Poland

[†] Deceased.

ABSTRACT

In the present study we investigate the concept of phylogenetic niche conservatism (PNC) within the American species of angraecoid orchids (*Campylocentrum* and *Dendrophylax*) and their closest relatives in the Old World (*Angraecum*) using ecological niche modelling (ENM). The predicted niche occupancy profiles were matched with the outcomes of previous phylogenetic studies to reconstruct the evolution of climatic suitability within the orchid group studied and evaluate the role of niche differentiation in the speciation of Angraecinae. No correlation between preferred niches and taxonomic relationships within the orchid group studied was revealed. The climatic suitability of the majority of the species overlapped each other, either fully or partially. This pattern is also present in the species of other orchid genera. Our research confirms a significant level of PNC in Orchidaceae, even within taxa exhibiting a transatlantic disjunction. The analysis of the evolution of climatic suitability indicated that the adaptation to various climatic conditions is not a factor that has driven speciation within orchids studied.

Submitted 5 July 2016
Accepted 18 April 2017
Published 16 May 2017

Corresponding author
Marta Kolanowska,
martakolanowska@wp.pl

Academic editor
Sara Varela

Additional Information and
Declarations can be found on
page 15

DOI 10.7717/peerj.3328

© Copyright
2017 Kolanowska et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Subjects Biodiversity, Biogeography, Ecology, Plant Science

Keywords Angraecinae, Ecological niche modeling, Orchidaceae, Phylogenetic niche conservatism, *Angraecum*, *Campylocentrum*, *Dendrophylax*

INTRODUCTION

As defined by *Harvey & Pagel (1991)*, phylogenetic niche conservatism (PNC) is the tendency of lineages to retain their ancestral ecological niche through speciation events. In fact, some of the phylogenetic studies conducted in recent years indicate that major aspects of the niche are more preserved during evolution than expected (*Donoghue, 2008; Olalla-Tárraga et al., 2011; Khaliq et al., 2015*). However, PNC theory does not suggest that ecological barriers are insurmountable. Obviously, niche evolution occurs. Niche conservatism can be considered as a significant factor in allopatric speciation because it limits adaptation to climatic conditions at the geographic barrier (*Wiens & Graham, 2005*).

Wiens & Donoghue (2004) argue that the interaction between niche conservatism and niche evolution may be critical in the biogeographic history of many groups.

Niche conservatism in species of plants may reflect the opportunities of their ancestors during their diversification. The differences between environmental gradients could be correlated with the palaeo-environmental conditions during the radiation of phylogenetically related lineages. *Prinzing et al. (2001)* assume that the variation in the characters of species is influenced by their adaptations to their present habitats, but some traits are a legacy from their ancestor. More recent studies (*Hadly, Spaeth & Li, 2009*) suggest that whilst niche conservatism at high taxonomic levels is primarily driven by inherent life history traits, at the species level it reflects the underlying environmental controls.

PNC is used to reveal the role of ecological divergence in speciation by many authors (e.g., *Kozak & Wiens, 2006; Swenson, 2010*) and for numerous animal groups (e.g., *Cooper, Freckleton & Jetz, 2011; Wellenreuther, Larson & Svensson, 2012; Pearman et al., 2014; Kerr et al., 2015; Rato et al., 2015; Morinière et al., 2016*). Relatively little research has so far been conducted on niche conservatism in plants (*Serra-Varela et al., 2015; Hawkins, Rodríguez & Weller, 2011; Hawkins et al., 2014; Wasof et al., 2015*) and most of these studies focus on the correlation between PNC and patterns in species richness or the composition of assemblages of species. The importance of niche differentiation in the evolution of various flowering plant groups remains poorly recognized.

The great diversity of orchids is most often attributed to their adaptive radiation in response to specific pollinators (e.g., *Paulus & Gack, 1990; Cozzolino & Widmer, 2005; Schiestl, 2005*), although *Otero & Flanagan (2006)* suggest that the obligatory orchid–mycorrhizal interactions should be considered as a factor promoting speciation. *Gravendeel et al. (2004)* argue that the epiphytism rather than pollinator specialization is the reason for the high species richness in orchids. There are only a few studies on PNC and the variation of climatic suitability within the Orchidaceae (*Kolanowska et al., 2016*) and most of the previous research on this aspect was on invasive species (*Kolanowska, 2013; Kolanowska & Konowalik, 2014*).

In the present study we combine research on the evolution of climatic suitability in closely related genera with an investigation of PNC in a taxon with a trans-Atlantic disjunction. The group studied, the subtribe Angraecinae Summerh. (Orchidaceae), comprises about 49 genera and exhibits a great variation in form and habit (*Carlswald et al., 2006; Micheneau et al., 2008; Pridgeon et al., 2014*). Some of the plants in this group produce elongate stems and well-developed leaves, others are characterized by reduced stems and small, scale-like leaves. The greatest diversity of angrecoid orchids is recorded in the Paleotropics, but two genera, *Campylocentrum* Benth. and *Dendrophylax* Rchb. f., occur only in the Americas. The leafless species occur only in the Neotropics (*Carlswald, Whitten & Williams, 2003*). The evolution of leaflessness apparently has occurred at least twice in the New World (*Carlswald et al., 2006*) and whilst there are both leafy and leafless species of *Campylocentrum*, *Dendrophylax* includes only leafless species.

The aim of the present study is to provide an insight into the importance of climatic niches in orchid speciation. To investigate PNC within geographically disjunct taxa we

evaluated the similarity of the niches occupied by American angraecoid orchids and their closest relatives from Africa. It is hypothesized that species from both continents occupy different niches and that species differentiation within genera is driven by non-climatic factors. However, there are no studies on niche conservatism in this group. To explore the role of adaptation to various climatic conditions in the speciation processes in Angraecinae the evolution of their climatic suitability was reconstructed by combining ecological niche modelling with a phylogenetic analysis.

MATERIALS & METHODS

Selection of taxa

Only the species included in the phylogenetic studies of [Carlsward et al. \(2006\)](#) and [Micheneau et al. \(2008\)](#) are included in this study. The localities of the populations of the following species are listed: *Angraecum chevalieri* Summerh. *A. cultriforme* Summerh. (= *Angraecoides cultriforme* (Summerh.) Szlach., Mytnik & Grochocka), *A. eichlerianum* Kraenzl. (= *Eichlerangraecum eichlerianum* (Kraenzl.) Szlach., Mytnik & Grochocka), *A. erectum* Summerh. (= *Angraecoides erecta* (Summerh.) Szlach., Mytnik & Grochocka), *Campylocentrum fasciola* (Lindl.) Cogn., *C. lansbergii* (Rchb. f.) Schltr., *C. micranthum* (Lindl.) Rolfe, *C. pachyrrhizum* (Rchb. f.) Rolfe, *C. poeppigii* (Rchb. f.) Rolfe, *C. tyrridion* Garay & Dunst., *Dendrophylax barrettiae* Fawc. & Rendle, *D. fawcetti* Rolfe (= *Polyrrhiza fawcetti* (Rolfe) Cogn.), *D. filiformis* (Griseb.) Benth. ex Fawc. (= *Harrisella filiformis* (Sw.) Cogn.), *D. funalis* (Sw.) Fawc., *D. lindenii* (Lindl.) Benth. ex Rolfe (= *Polyradicion lindenii* (Lindl.) Garay), *D. porrectus* (Rchb. f.) Carlsward & Whitten (= *Harrisella porrecta* (Rchb. f.) Fawc. & Rendle), *D. sallei* (Rchb. f.) Benth. ex Rolfe (= *Polyradicion sallei* (Rchb. f.) Garay) and *D. varius* Urb. Based on morphological studies ([Kolanowska, Pérez Escobar & Parra Sánchez, 2012](#); [Kolanowska & Szlachetko, 2013](#); [Szlachetko & Kolanowska, 2013a](#); [Szlachetko & Kolanowska, 2013b](#)) we consider *C. sullivanii* Fawc. & Rendle as conspecific with *C. fasciola* as these taxa are morphologically indistinguishable. This is also the conclusion of [Bogarín & Pupulin \(2010\)](#).

List of localities

The occurrence data were obtained from the information recorded with the specimens deposited in herbaria AMES (Orchid Herbarium of Oakes Ames), AMO (Asociación Mexicana de Orquideología), BM (Natural History Museum, London), COL (Universidad Nacional de Colombia), F (Field Museum of Natural History), FLAS (Florida Museum of Natural History), K (Royal Botanic Gardens), MO (Missouri Botanical Garden), NY (New York Botanical Garden), and P (Muséum National d'Histoire Naturelle), as well as from the literature and the original protologues. Herbarium acronyms follow Index Herbariorum ([Thiers, 2015](#)). The georeferencing process followed is that used by [Hijmans et al. \(1999\)](#). The geographic coordinates on the herbarium sheet labels were verified. If there were no geographic coordinates on the herbarium label, we used the description of the locality where the plant was collected and assigned coordinates as precisely as possible to this location. The Google Earth (ver. 6.1.0.5001, Google Inc.) application was used to validate all the information gathered. A total of 162 localities that could be precisely located were

used in the ENM (5–46 localities per species; [Table S1](#)). *Dendrophylax filiformis* (Griseb.) Benth. ex Fawc. (= *Harrisella filiformis* (Sw.) Cogn.) and *Dendrophylax funalis* (Sw.) Fawc. were not included in this analysis because of the lack of a precise locality for them.

Ecological niche modelling and niche similarity

Input data for the ecological niche modelling were 35 bioclimatic variables with a 10 arc minute resolution obtained from the CliMond dataset ([Kriticos et al., 2012](#)). To choose an appropriate modelling extent (i.e., the species movement limits described in the M of the BAM diagram; see [Barve et al., 2011](#)) terrestrial ecoregions where species occur were selected. Biotic regions serve as a reliable estimate of the area that is accessible to a species and the easiest way to accurately designate the distribution limits of a species ([Soberón & Peterson, 2005](#); [Barve et al., 2011](#)). For this purpose, the world map of terrestrial ecoregions was used ([Olson et al., 2001](#)). The bioclimatic maps were clipped to include only those regions where it was confirmed that the species occurred. In addition, some island archipelagos and island-like regions were removed from the study area (i.e., Cape Verde archipelago, Ascension Island, Saint Helena, Nile Delta and some smaller features in North Africa north of 19°N). To account for the multicollinearity nature of the initial variables, a principal component analysis (PCA) was performed in R ([R Core Team, 2014](#)).

To select an appropriate number of PCA derived maps (i.e., PC axes) the Kaiser-Guttman criterion, which is based on a mean of all eigenvalues was used by selecting only PC axes with eigenvalues larger than this mean. Following this criterion, maps based on the first six principal components were selected ([Table S2](#)). The ecological niche modelling was done using a maximum entropy method implemented in Maxent version 3.3.3k ([Phillips, Dudík & Schapire, 2004](#); [Phillips, Anderson & Schapire, 2006](#); [Elith et al., 2011](#)), which is based only on species presence. The maximum iterations were set to 10,000 and convergence threshold to 0.00001. Random seeds were used to keep 1,000 bootstrapping runs from using replicate test and training samples. For each run 20% of the data were set aside as test points ([Urbina-Cardona & Loyola, 2008](#)). In order to maintain a sufficient sample size for taxa with a small number of occurrences (below 10) duplicate presence records were not removed and presence points were duplicated within a given grid cell. This procedure was only used for taxa that are narrowly distributed insular endemics with a small number of occurrences for which it would be difficult carry out climatic filtering ([Varela, Anderson & García-Valdés, 2014](#)). To simplify the interpretation of the probability of a species being present the logistic Maxent output format was chosen.

The model was evaluated using the most common metrics, area under the curve (AUC; [Mason & Graham, 2002](#)) and true skill statistic (TSS; [Peirce, 1884](#)). AUC was calculated using the Maxent application automatically based on the training localities. Whilst some authors suggest that it may be misleading ([Lobo, Jiménez-Valverde & Real, 2008](#)), it seems to be a valid metric for determining the reliability of the fit of the ENM ([Warren, Glor & Turelli, 2008](#)). TSS was calculated using maximum training sensitivity plus specificity.

The similarity between the niches occupied by the species studied was measured using Schoener's D (D; [Schoener, 1968](#)) and I statistic (I; [Van der Vaart, 1998](#); [Warren, Glor & Turelli, 2008](#); [Warren, Glor & Turelli, 2010](#)) implemented in ENMTools package for R

(Warren, 2016), using the methods for calculating environmental distances proposed by Broennimann *et al.* (2012). Schoener's D statistic uses direct measures of species density, which in this study are changed to measures of densities of occurrence modelled in environmental space. 'I' statistic is based on the modified Hellinger distance that compares two probability distributions. These two metrics range from 0 (no similarity) to 1 (high similarity). To test the importance of distance the niche identity test was used and calculated by the same function that produced the D and I statistics in the ENMTools package for R (Broennimann *et al.*, 2012; Warren, 2016). To visualize the suitable climatic niche of each species, a PCA of the raw climatic conditions occurring at given locations was performed using R (R Core Team, 2014).

Operations on GIS data were carried out on ArcGis 9.3 (ESRI) and R (R Core Team, 2014).

Phylogenetic analysis

To construct the phylogenetic tree, sequences from ITS and *trnL-F* (Table S3) were aligned using Mafft 6.833b (Katoh & Toh, 2008). Gapcoder (Young & Healy, 2003) was used to code indels. The Alignments were then merged and a Bayesian phylogenetic analysis was performed using MrBayes 3.2.1 (Ronquist *et al.*, 2012). For the nucleotide part, the best model according to the AIC implemented in jModelTest 2.1.1 (Darriba *et al.*, 2012) was used. For the binary coded gaps, a Jukes–Cantor model (Jukes & Cantor, 1969) was used. 15,000,000 generations were performed in two runs, discarding the first 25% as the burning in fraction and sampling every 1,000th tree. To estimate node ages function chronos in package “ape” was used as described previously (Kolanowska *et al.*, 2016). Lambda was set to 20 and ages were estimated using a semi-parametric method based on penalized likelihood where branch lengths indicate mean numbers of substitutions (Sanderson, 2002; Paradis, Claude & Strimmer, 2004). As this calibration indicates splits between *Campylocentrum* and *Dendrophylax* (max. 12.72 Mya, min. 4.64 Mya), and *Angraecum* and *Polystachya* (max. 32.48 Mya, min. 22.2 Mya) we used published divergence times (Givnish *et al.*, 2015; Andriananjamanantsoa *et al.*, 2016).

To reconstruct ancestral climatic suitabilities the Phyloclim package was used (Heibl & Calenge, 2013) which implements the methods originally developed by Evans *et al.* (2009). Predicted niche occupancy (PNO) was reconstructed and together with the phylogenetic tree were used to infer ancestral climatic suitabilities. PNO integrates species probability distributions (derived using MaxEnt) with respect to climatic variables. Ancestral climatic suitabilities are the PNOs projected onto the phylogenetic tree. They were reconstructed based on the distribution of climatic suitabilities (PNOs) using maximum likelihood and Brownian motion assumption to plot them at each interior node of the tree.

In addition, a Mantel test was used to verify the correlation between the genetic and niche distances (Serra-Varela *et al.*, 2015). This analysis was performed using the Mantel test available in the ADE4 package of R (Dray & Dufour, 2007) and its significance was assessed by performing 9,999 replications. Genetic distances were generated in PAUP* (Swofford, 2002) using the distances predicted by the GTR model and as niche distances

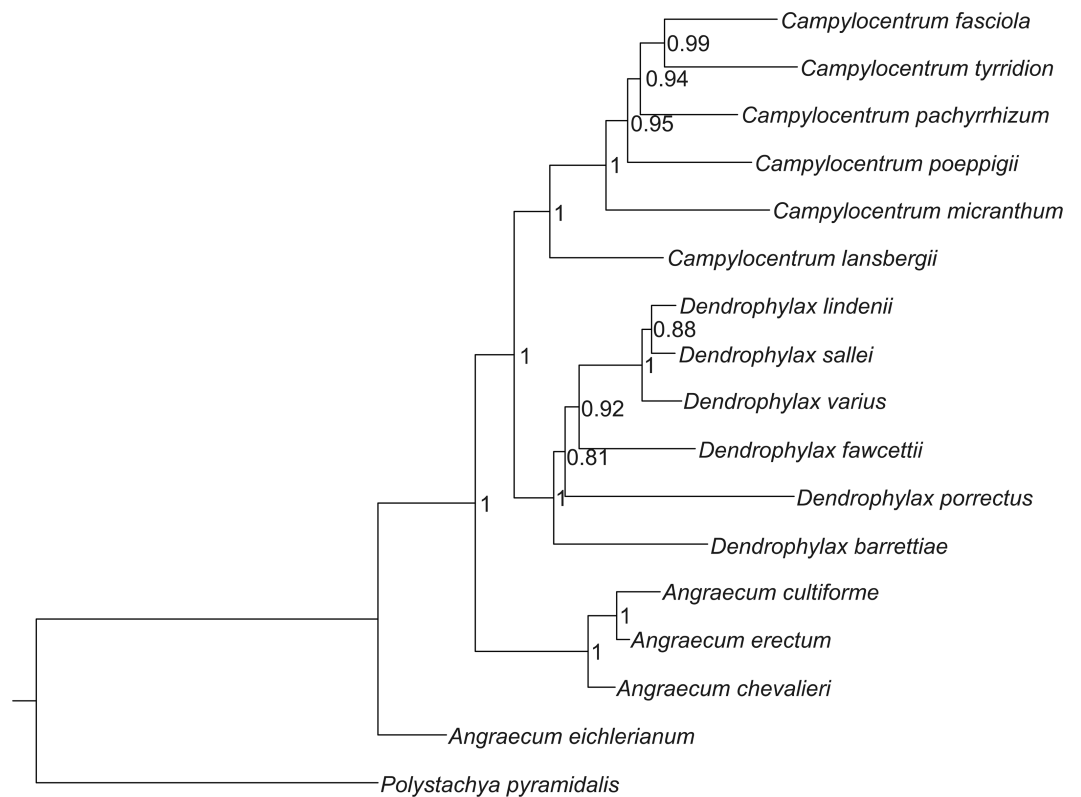


Figure 1 Bayesian inference tree of the combined nuclear ITS and chloroplast *trnL-F* intergenic spacers for 16 species of Angraecinae and one outgroup species. Numbers beside the nodes indicate posterior probability.

Schoener's D indices were used. GTR distances were chosen based on the best AIC score in the jModelTest 2.1.1 (Darriba et al., 2012).

Age range correlation (ARC, Fitzpatrick & Turelli, 2006) was performed in R using package phyloclim (Heibl & Calenge, 2013). ARC is equivalent to phylogenetic independent contrasts (PIC) and explains how niche similarity (in this case measured using D and I statistics) change over time, which is represented by the nodes of the phylogenetic tree (Fitzpatrick & Turelli, 2006). Monte Carlo resampling with 3,000 replicates was used to assess the statistical significance. This test was used to verify whether any of the observed differences could be explained by phylogeny or whether these are more probably a result of "ecological drift" than "ecological specialization".

RESULTS

Phylogenetic relationships

The Bayesian tree, based on sequences of both ITS and *trnL-F*, indicate that *Campylocentrum* and *Dendrophylax* form well-supported clades, which originated from African representatives of *Angraecum* (Fig. 1). These analyses support the results of the previous molecular studies of Carlswald et al. (2006), Micheneau et al. (2008) and Szlachetko et al. (2013).

Table 1 The average training AUC and TSS for the replicate runs.

Species	AUC	TSS
<i>A. chevalieri</i>	0.972	0.879
<i>A. cultriforme</i>	0.767	0.564
<i>A. eichlerianum</i>	0.952	0.795
<i>A. erectum</i>	0.990	0.937
<i>C. fasciola</i>	0.958	0.891
<i>C. lansbergii</i>	0.985	0.974
<i>C. micranthum</i>	0.886	0.743
<i>C. pachyrrhizum</i>	0.940	0.879
<i>C. poeppigii</i>	0.934	0.848
<i>C. tyrridion</i>	0.914	0.770
<i>D. barrettiae</i>	0.918	0.831
<i>D. fawcettii</i>	0.975	0.907
<i>D. lindenii</i>	0.836	0.773
<i>D. porrectus</i>	0.690	0.499
<i>D. sallei</i>	0.846	0.736
<i>D. varius</i>	0.805	0.735

Evaluation of the modelling of the ecological niche

All projected niche models had high AUC scores ranging from 0.690 to 0.990 (Table 1) indicating the model is good in distinguishing presence data from background data (Phillips, Anderson & Schapire, 2006). The TSS scores showed the same trend but were lower than the AUC scores and ranged from 0.499 to 0.974. The low scores of AUC and TSS are mainly for taxa that are narrow endemics (e.g., those occurring on Caribbean islands). As these scores are close to that of a random prediction they should be treated with caution and may indicate that climate is not a significant factor influencing their distribution (most probably they originated by allopatric speciation, which is not necessarily reinforcing climatic differences).

Distribution of suitable niches and niche similarity

The distribution of the suitable niches of all species studied is presented in Figs. 2–4. This indicates there is a relatively low cover of suitable habitats for *Dendrophylax fawcettii* and *Campylocentrum lansbergii*.

The Mantel test did not reveal a correlation between genetic and environmental distance ($r = -0.14$, $p = 0.87$) supporting the hypothesis that niche evolution (i.e., niche difference) is not related to phylogeny and that phylogenetically distant taxa can occupy similar niches. In addition, the results of this analysis (Fig. 5) did not indicate a correlation between the similarity of the niches occupied by the species studied and their phylogenetic relationships (Fig. 1). Overall, the most similar niches are occupied by *Angraecum cultriforme* and *Dendrophylax porrectus* ($I = 0.99$, $D = 0.90$), whilst the greatest differences were recorded for *Dendrophylax fawcettii* and *Angraecum erectum* ($I = 0.10$, $D = 0.03$). The general similarity of the niches occupied by Angraecinae is also visible in the PCA graph (Fig. 6). In this diagram the suitable niches of many clades overlap significantly. It is noteworthy that

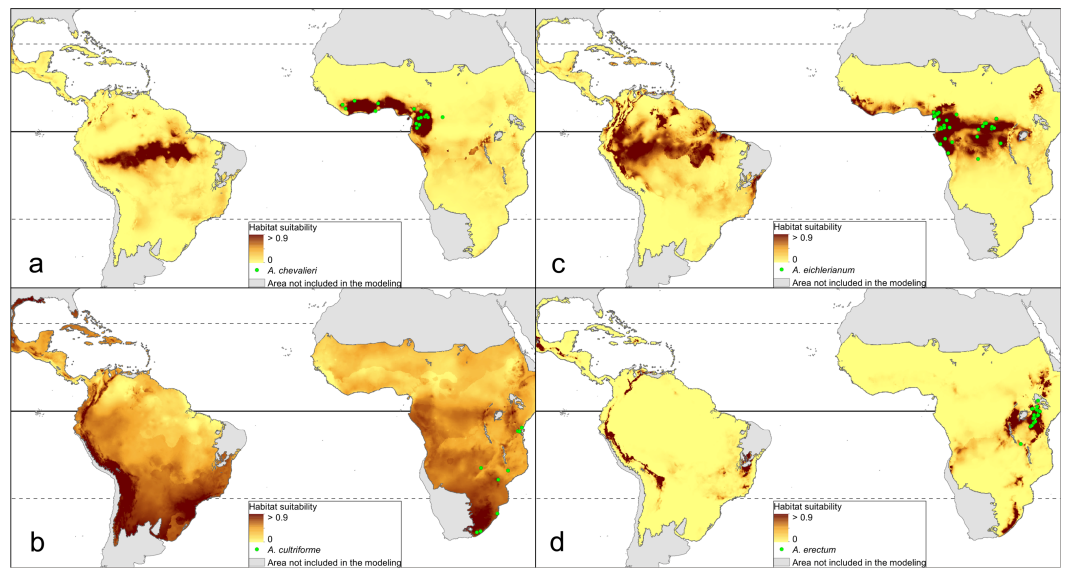


Figure 2 Distribution of suitable niches of *Angraecum* representatives. (A) *Angraecum chevalieri*; (B) *Angraecum cultriforme*; (C) *Angraecum eichlerianum*; (D) *Angraecum erectum*. Localities used in the ENM analysis marked as green spots.

the niches of all *Dendrophylax* species fall within those of *Angraecum* and *Campylocentrum*. The climatic suitabilities of these particular species also overlap each other and there are no clear boundaries between the genera (taxa) studied in terms of their preferred climatic conditions.

Ancestral climatic suitability

There are no significant differences in the evolutionary history of the taxa studied in their climatic suitabilities and tolerance of species overlap (Fig. 7). Species of both the Neotropical genera included in this study appeared to have slightly wider climatic suitabilities than their closest African relatives. The climatic suitability of *Angraecum eichlerianum* measured in terms of the climatic factors analyzed has changed very little over time. The niches of the other species of *Angraecum* studied (*A. erectum*, *A. chevalieri* and *A. cultriforme*) diverged and stabilized much later. The climatic suitability of the Neotropical species evolved in various directions and the climatic niche was apparently not highly conserved within particular genera. Climatic suitability of the African *Angraecum cultriforme* and *A. erectum* changed and became more similar to that of Neotropical Angraecinae.

Similar results were obtained from the age range correlation using both D and I for which the slope is negative and intercept is higher than 0.5, which could indicate sympatric speciation (Fitzpatrick & Turelli, 2006; Warren, Glor & Turelli, 2008). These results imply that recently diverged nodes are more similar than more ancestral nodes and recent species occupy similar niches (Fig. 8). ARC was not significant (Table 2), which indicates that climatic niche differentiation did not play a role in the diversification of Angraecinae. The

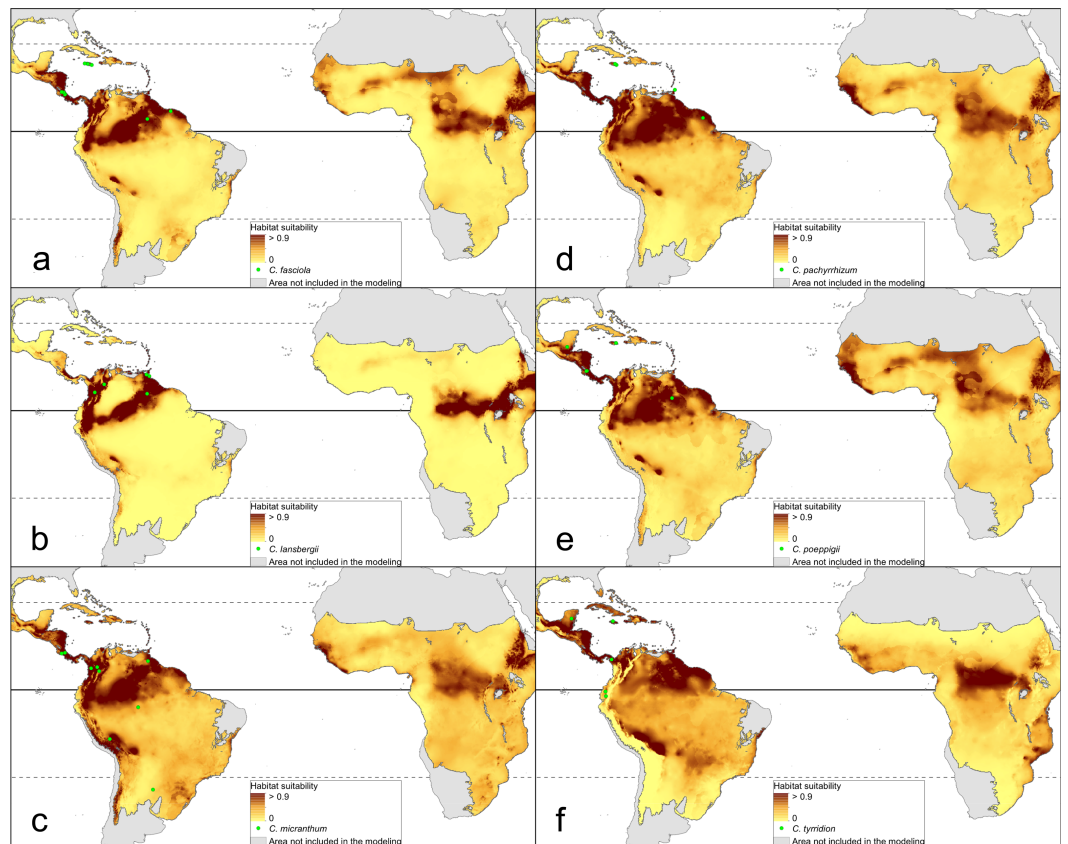


Figure 3 Distribution of suitable niches of *Campylocentrum* representatives. (A) *Campylocentrum fasciola*; (B) *Campylocentrum lansbergii*; (C) *Campylocentrum micranthum*; (D) *Campylocentrum pachyrrhizum*; (E) *Campylocentrum poeppigii*; (F) *Campylocentrum tyrridion*. Localities used in the ENM analysis marked as green spots.

Table 2 Results of the age-range correlation (ARC) analyses using randomization tests under Monte Carlo resampling.

	Intercept	f (greater) ¹	Slope	f (greater) ¹
D	0.6747	0.055 ($P = 0.11$)	-0.2719	0.89 ($P = 0.23$)
I	0.8528	0.13 ($P = 0.25$)	-0.1727	0.79 ($P = 0.42$)

evolution of Angraecinae corresponds to Brownian motion. The species in this group do not exhibit ecological specialization within their climatic suitabilities.

DISCUSSION

Phylogenetic niche conservatism

Traditionally, natural selection and adaptation result in lineages inhabiting different environments. However, the similarity between the niches occupied by Neotropical angraecoid orchids and their African relatives indicated by this research suggests that they have retained the general climatic suitability of their common progenitor. The recorded shift in climatic suitability of two ancestral African angraecoid orchids could have occurred

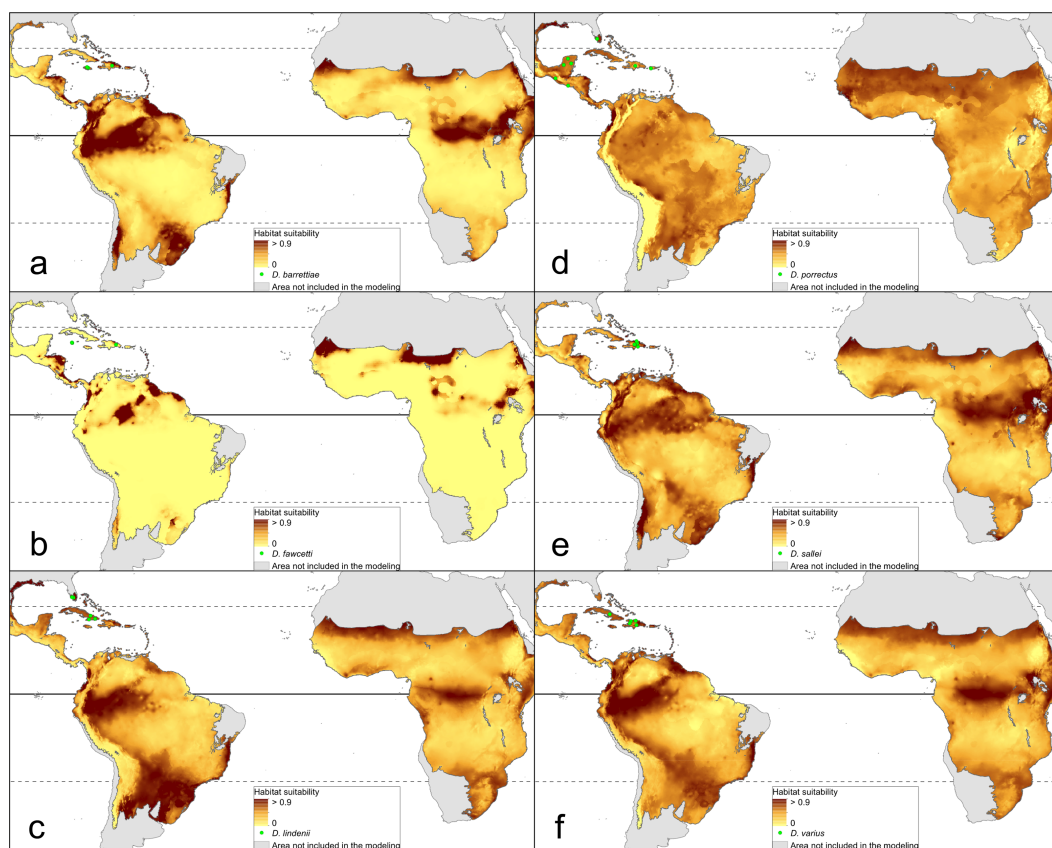


Figure 4 Distribution of suitable niches of *Dendrophylax* representatives. (A) *Dendrophylax barrettiae*; (B) *Dendrophylax fawcettii*; (C) *Dendrophylax lindenii*; (D) *Dendrophylax porrectus*; (E) *Dendrophylax sallei*; (F) *Dendrophylax varius*. Localities used in the ENM analysis marked as green spots.

relatively recently, after the stabilization of niche preferences within New World species of this subtribe. Another possible explanation is that their climatic suitabilities were derived from that of the ancestor of these taxa.

Wiens (2004) suggests that limited adaptation to environmental conditions is a crucial factor in promoting the divergence in the initial origin of lineages and less important in the subsequent divergence of these lineages. The results of this study reveal that climatic niche diversification did not significantly influence the speciation of the Neotropical angreoid orchids. This is the third study showing that PNC in Orchidaceae is very high (*Kolanowska & Szlachetko, 2014; Kolanowska et al., 2016*) and that their diversity cannot be explained by niche differentiation. Our results confirm Wiens's assumption about the low effect of niche modification in speciation.

Evolutionary implications

The divergence of Neotropical angreoid orchids from their African relatives took place ca. 11.5–16 Mya (*Micheneau et al., 2010*), in the Miocene, but the diversification within the American representatives of Angraecinae began about 6–7 Mya. The late Miocene marked the start of a paroxysm resulting in the uplift of the northeastern Andes (*Hoorn et al., 1995*)

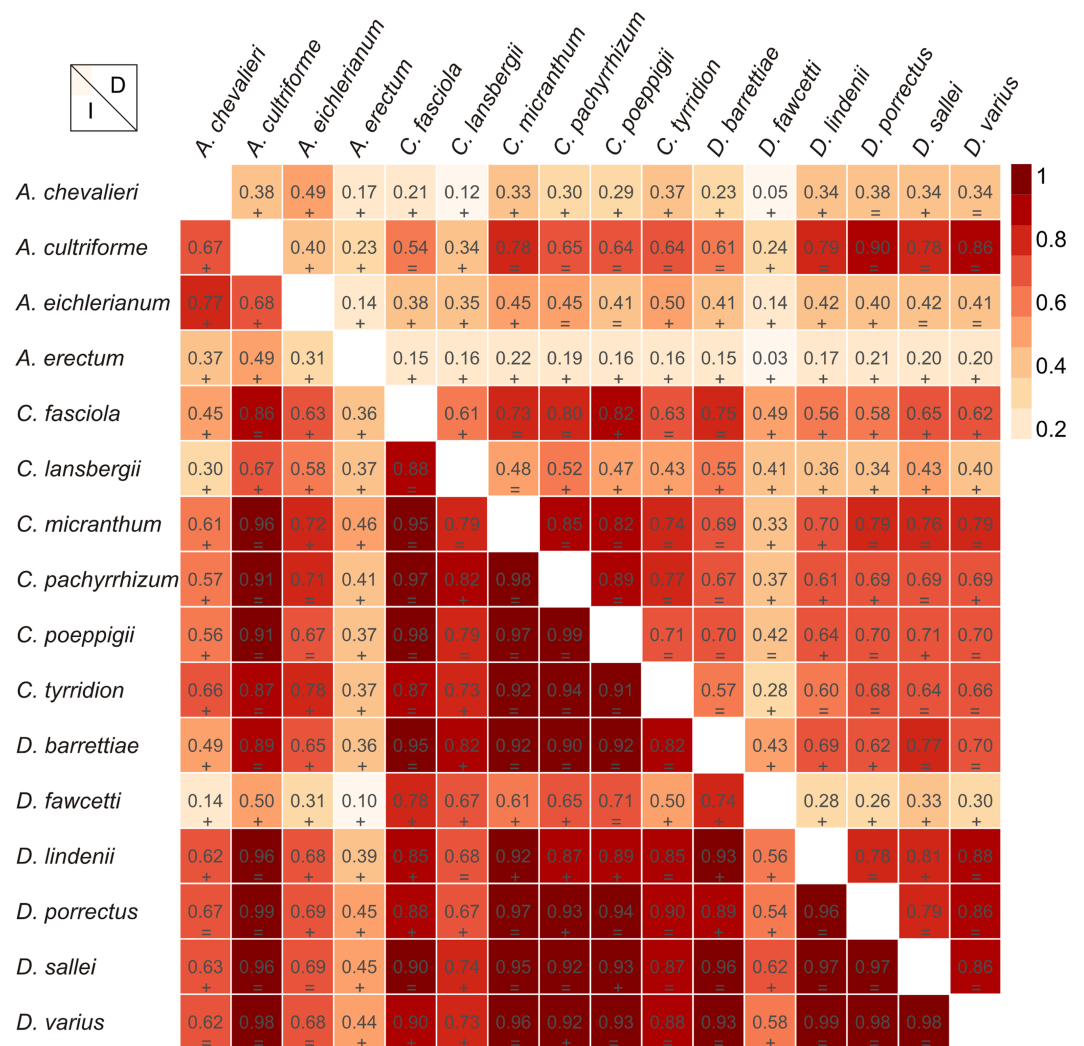


Figure 5 Niche similarities of the studied taxa based on the Schoener's D (upper triangular) and I statistic (lower triangular). Higher indexes indicate more similarity as shown by the red intensity. Results of the niche identity test are marked below each value where "+" means that the niches are different ($p = 0.01$) and "=" indicate the the niches are similar ($p > 0.01$).

and a significant increase in sediment rates between ca. 7.9 and 6 Mya (*Uba, Strecker & Schmitt, 2007*). The Andean sediments reached the Atlantic coast via the Amazon drainage system, and the Amazon River became fully established at about 7 Mya (*Hoorn et al., 2010*). The rise of the Andes influenced climatic conditions in South America and the range began to constitute a solid migration barrier that limited the dispersal of *Campylocentrum*. Uplift of the Lesser Antilles arc and the associated platform initiated during Early-Middle Miocene formed a barrier to sedimentation between the two basins by the late Miocene. After the Miocene, most deposition in the Greater Antilles was confined to present coastal and offshore areas (*Khudoley & Meyerhoff, 1971*). This heterogeneity in topography and environmental conditions probably led to rapid divergence between populations, as it reduced the dispersal and gene flow between adjacent populations. Based on the geological

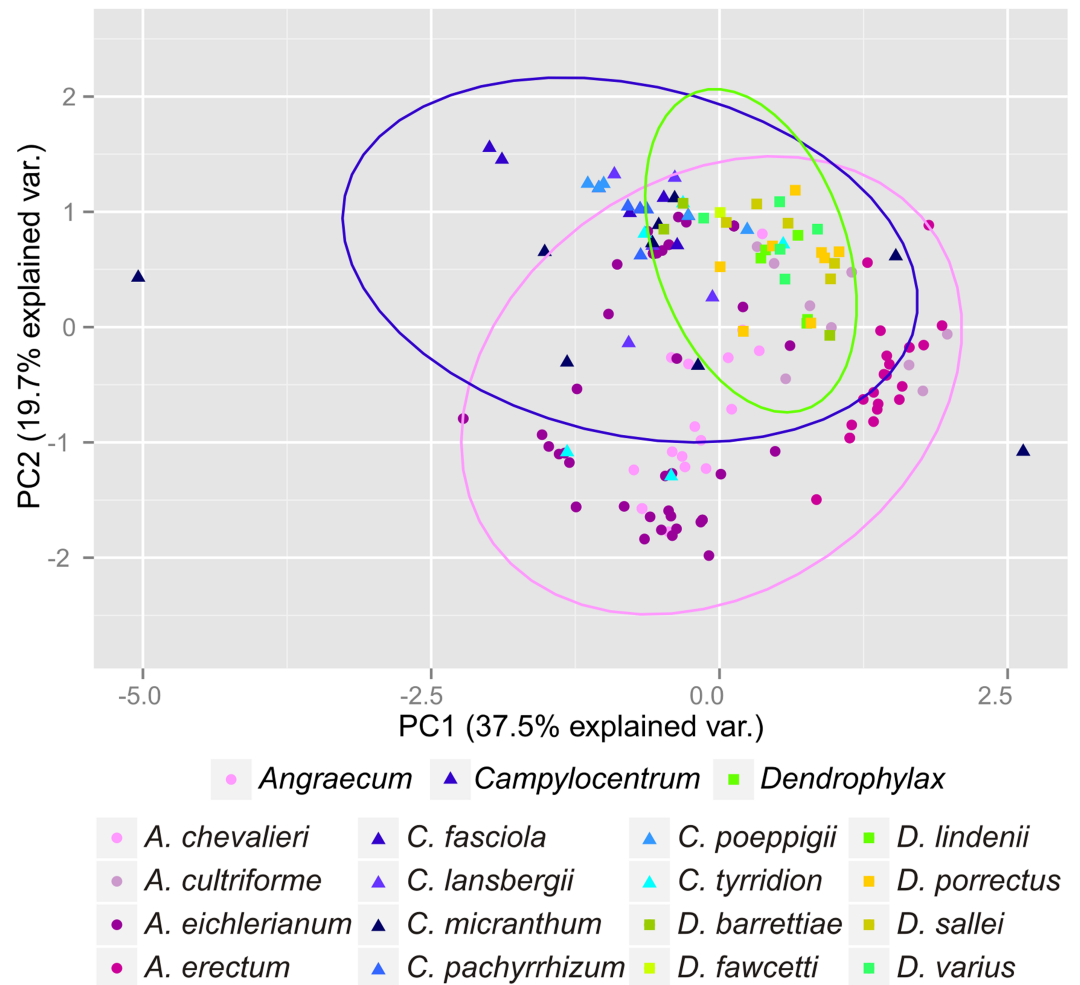


Figure 6 Principal Component Analysis of climatic variables present in the studied populations.

events that occurred at the time of diversification of Neotropical Angraecinae and the result of the ENM analysis we assume that adaptation to various niches was not the main factor determining speciation within the taxa studied.

The climatic-based potential ranges of the Neotropical species studied are considerably larger than the observed distributions of these orchids, indicating that abiotic conditions are not the main factor limiting the occurrence of *Campylocentrum* and *Dendrophylax*. Whilst the species of these two genera are characterized by similar climatic suitabilities and they share the same evolutionary and geological history, their known geographical ranges differ significantly. Populations of *Campylocentrum* occur on both sides of the Andes as well as in Mesoamerica and the West Indies. The distribution of *Dendrophylax* is restricted to Mesoamerica and the West Indies. Since there is no report of a firm relationship between any of the American angrecoid orchids and a specific phorophyte, we believe that only two factors could promote the diversification of these two genera: pollinator specificity and/or mycorrhizal specialization. Unfortunately, little is known about both of these two aspects of angrecoid orchid biology. Preliminary studies reveal that some

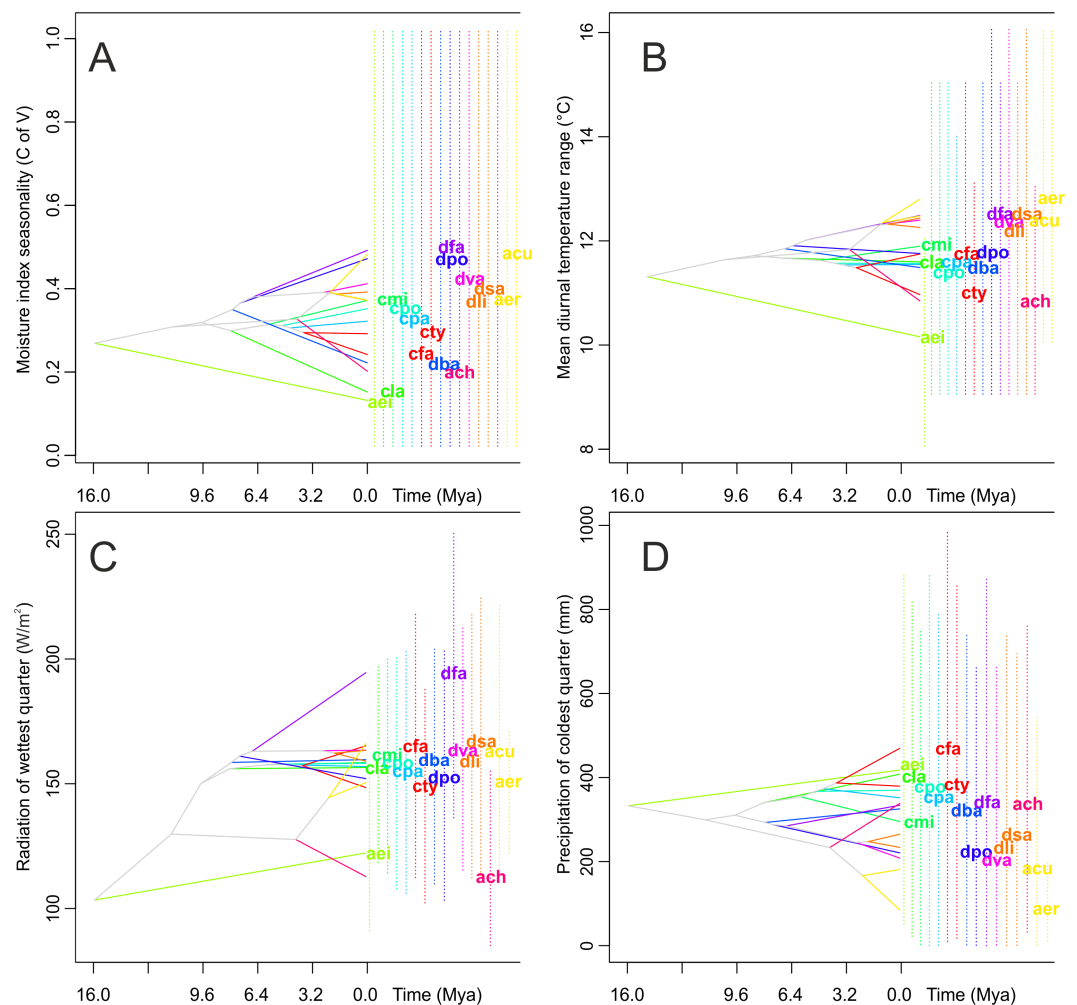


Figure 7 Examples of evolution of climatic tolerances in Angraecinae visualized on the consensus tree from Bayesian inference. (A) Moisture index seasonality (bioclim 31); (B) Mean diurnal temperature range (bioclim 2); (C) Radiation of wettest quarter (bioclim 24); and (D) Precipitation of coldest quarter (bioclim 19). The name of each studied taxon was abbreviated to the first letter of generic name and two first letters of specific epithet (e.g., *Campylocentrum fasciola* code is (“cfa”). Interior nodes represent the mean of climatic tolerances inferred for the most recent common ancestor of the extant taxa defined by that node. The 80% central density of climatic tolerance for each extant taxon is indicated by a vertical dashed line, and the mean is indicated by the taxon label, to the right of each graph. For visualization the four most important uncorrelated variables were chosen as indicated by Maxent analysis on a full set of 35 bioclimatic variables.

species of *Campylocentrum* form mycorrhizal associations with Ceratobasidiaceae (Otero, Ackerman & Bayman, 2002) and it is possible that the same symbiotic association occurs in *Dendrophylax lindenii* (Chomicki, Bidel & Jay-Allemand, 2014). Undoubtedly, leafless species rely on endomycorrhizal associations, but the diversity of their mycobionts remains unknown.

Pollinator specificity is suggested as a promoter of evolution and speciation in angraecoid orchids by Dressler (1981). Both *Campylocentrum* and *Dendrophylax* are entomophilous, although flowers of *Dendrophylax* are reported to be pollinated by sphingid moths and

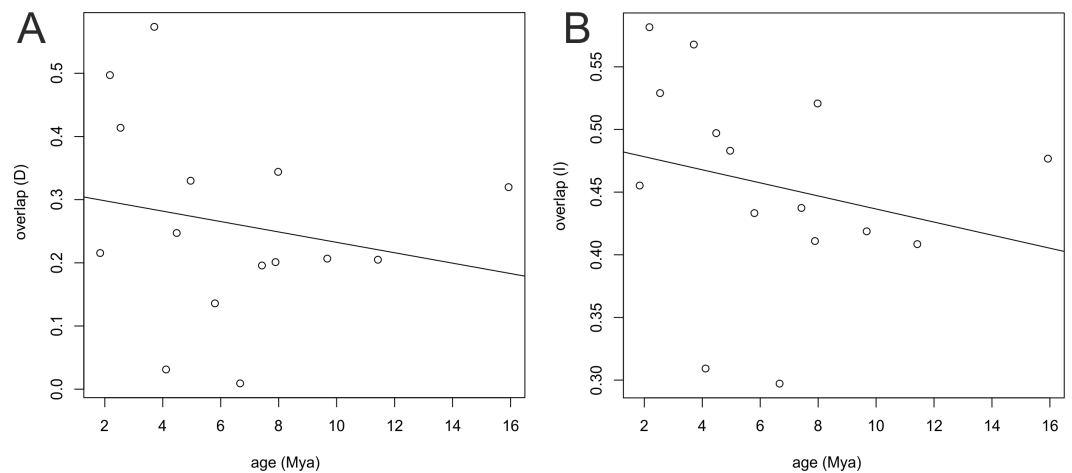


Figure 8 Result of age range correlation (ARC) analysis which presents phylogenetic signal of niche similarity for both genera using D statistic (A) or I statistic (B) in function of time (Mya). Each dot represents a node of the phylogenetic tree (Fig. 1) and the dots on the right side represent the ancestral nodes. Line is the fitted regression.

the pollen of *Campylocentrum* is also transferred by halictid and meliponini bees (Singer & Cocucci, 1999; Singer, 2003). Whilst species of *Campylocentrum* grow sympatrically (Pupulin, 1998; Pupulin, 2001; Bogarín & Pupulin, 2010), there are no records of hybrids between any the Neotropical angrecoid orchids. The lack of any significant differences in climatic suitability is also recorded in the African *Angraecum*, a close relative of Neotropical Angraecinae. The African angrecoids are highly specialized in terms of moth pollination. Most of *Angraecum* species produce long-spurred, white, nectariferous flowers. These floral characters are consistent with hawkmoth pollination (Darwin, 1862; Grant, 1985; Nilsson et al., 1985; Haber & Frankie, 1989; Micheneau et al., 2008). However, long-spurred species, especially those in Madagascar, are recognized as ancestral in the angrecoid orchid group (Nilsson et al., 1985). The studies on the pollination of species of *Angraecum* have mainly been on the Malgasy (Nilsson et al., 1985; Arditti et al., 2012) and Mascarene Islands (Micheneau, Fournel & Pailler, 2006; Micheneau et al., 2008). Moreover, the research of Micheneau et al. (2008) indicates that species of section *Hadrangis*, which have atypical short-spurred, scentless flowers are pollinated by birds and crickets. Unfortunately, the data on pollination are still very scanty and the relationship with pollinators of *Angraecum* species on Mainland Africa has not been studied. Most likely differences in pollinator composition provides a strong barrier to gene flow.

In conclusion, we found that niche conservatism in Angrecinae is very strong and that the African and Neotropical species in this group have similar climatic suitabilities. Moreover, climatic niche differentiation does not appear to be an important factor in the speciation of *Dendrophylax* and *Campylocentrum*. We suggest that pollinator specificity or restricted mycorrhizal associations played a crucial role in the development of the diversity of species in the taxa studied.

ACKNOWLEDGEMENTS

The authors thank the Curators and staff of the herbaria cited for their hospitality and assistance during our visits. We are grateful to an anonymous Reviewer for the valuable comments on this manuscript and to Jacek Wajer and Anthony Dixon for linguistic correction.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

The research was supported by The Foundation For Polish Science (Fundacja na rzecz Nauki Polskiej, FNP) and from the grant nr 14-36098G of the Grantová agentura České Republiky (GA ČR). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:
The Foundation For Polish Science (Fundacja na rzecz Nauki Polskiej, FNP).
Grantová agentura České Republiky (GA ČR): 14-36098G.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Marta Kolanowska, Elżbieta Grochocka and Kamil Konowalik conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.

Data Availability

The following information was supplied regarding data availability:
The raw data has been supplied as a [Supplementary File](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.3328#supplemental-information>.

REFERENCES

- Andriananjamanantsoa HN, Engberg S, Louis EE, Brouillet L. 2016.** Diversification of *Angraecum* (Orchidaceae, Vandaeae) in Madagascar: revised phylogeny reveals species accumulation through time rather than rapid radiation. *PLOS ONE* **11(9)**:e0163194 DOI [10.1371/journal.pone.0163194](https://doi.org/10.1371/journal.pone.0163194).

- Arditti J, Elliott J, Kitching IJ, Wasserthal LT. 2012.** ‘Good heavens what insect can suck it’—Charles Darwin, *Angraecum sesquipedale* and *Xanthopan morgani praedicta*. *Botanical Journal of the Linnean Society* **169**(3):403–432 DOI [10.1111/j.1095-8339.2012.01250.x](https://doi.org/10.1111/j.1095-8339.2012.01250.x).
- Barve N, Barve V, Jiménez-Valverde A, Lira-Noriega A, Maher SP, Peterson AT, Soberón J, Villalobos F. 2011.** The crucial role of the accessible area in ecological niche modeling and species distribution modeling. *Ecological Modelling* **222**(11):1810–1819 DOI [10.1016/j.ecolmodel.2011.02.011](https://doi.org/10.1016/j.ecolmodel.2011.02.011).
- Bogarín D, Pupulin F. 2010.** The genus *Campylocentrum* (Orchidaceae: angraecinae) in costa rica: a revision. *Harvard Papers in Botany* **15**:353–414 DOI [10.3100/025.015.0216](https://doi.org/10.3100/025.015.0216).
- Broennimann O, Fitzpatrick MC, Pearman PB, Petitpierre B, Pellissier L, Yoccoz NG, Thuiller W, Fortin MJ, Randin C, Zimmermann NE, Graham CH, Guisan A. 2012.** Measuring ecological niche overlap from occurrence and spatial environmental data. *Global Ecology and Biogeography* **21**:481–497 DOI [10.1111/j.1466-8238.2011.00698.x](https://doi.org/10.1111/j.1466-8238.2011.00698.x).
- Carlsward BS, Whitten WM, Williams NH. 2003.** Molecular phylogenetics of neotropical leafless angraecinae (Orchidaceae): reevaluation of generic concepts. *International Journal of Plant Sciences* **164**:43–51 DOI [10.1086/344757](https://doi.org/10.1086/344757).
- Carlsward BS, Whitten WM, Williams NH, Bytebier B. 2006.** Molecular phylogenetics of Vandeeae (Orchidaceae) and the evolution of leaflessness. *American Journal of Botany* **93**:770–786 DOI [10.3732/ajb.93.5.770](https://doi.org/10.3732/ajb.93.5.770).
- Chomicki G, Bidel LPR, Jay-Allemand C. 2014.** Exodermis structure controls fungal invasion in the leafless epiphytic orchid *Dendrophylax lindenii* (Lindl.) Benth. ex Rolfe. *Flora* **209**:88–94 DOI [10.1016/j.flora.2014.01.001](https://doi.org/10.1016/j.flora.2014.01.001).
- Cooper N, Freckleton RP, Jetz W. 2011.** Phylogenetic conservatism of environmental niches in mammals. *Proceedings of the Royal Society of London B: Biological Sciences* **278**:2384–2391 DOI [10.1098/rspb.2010.2207](https://doi.org/10.1098/rspb.2010.2207).
- Cozzolino S, Widmer A. 2005.** Orchid diversity: an evolutionary consequence of deception? *Trends in Ecology and Evolution* **20**:487–494 DOI [10.1016/j.tree.2005.06.004](https://doi.org/10.1016/j.tree.2005.06.004).
- Darriba D, Taboada GL, Doallo R, Posada D. 2012.** jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods* **9**(8):772 DOI [10.1038/nmeth.2109](https://doi.org/10.1038/nmeth.2109).
- Darwin C. 1862.** *On the various contrivances by which British and foreign orchids are fertilized by insects, and on the good effect of intercrossing*. London: John Murray.
- Donoghue MJ. 2008.** A phylogenetic perspective on the distribution of plant diversity. *Proceedings of the National Academy of Sciences of the United States of America* **105**(suppl. 1):11549–11555 DOI [10.1073/pnas.0801962105](https://doi.org/10.1073/pnas.0801962105).
- Dray S, Dufour AB. 2007.** The ade4 package: implementing the duality diagram for ecologists. *Journal of Statistical Software* **22**:1–20 DOI [10.18637/jss.v022.i04](https://doi.org/10.18637/jss.v022.i04).
- Dressler R. 1981.** *The orchids—natural history and classification*. Cambridge: Harvard University Press.

- Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ. 2011.** A statistical explanation of MaxEnt for ecologists. *Diversity and Distributions* 17:43–57 DOI 10.1111/j.1472-4642.2010.00725.x.
- Evans ME, Smith SA, Flynn RS, Donoghue MJ. 2009.** Climate, niche evolution, and diversification of the “bird-cage” evening primroses (*Oenothera*, sections *Anogra* and *Kleinia*). *The American Naturalist* 173(2):225–240 DOI 10.1086/595757.
- Fitzpatrick BM, Turelli M. 2006.** The geography of mammalian speciation: mixed signals from phylogenies and range maps. *Evolution* 60(3):601–615 DOI 10.1111/j.0014-3820.2006.tb01140.x.
- Givnish TJ, Spalink D, Ames M, Lyon SP, Hunter SJ, Zuluaga A, Iles WJD, Clements MA, Arroyo MTK, Leebens-Mack J, Endara L, Kriebel R, Neubig KM, Whitten WM, Williams NH, Cameron KM. 2015.** Orchid phylogenomics and multiple drivers of their extraordinary diversification. *Proceedings of the Royal Society of London Series B Biological Sciences* 282(1814):20151553+ DOI 10.1098/rspb.2015.1553.
- Grant V. 1985.** Additional observations on temperate North American hawkmoth flowers. *Botanical Gazette* 146:517–520 DOI 10.1086/337557.
- Gravendeel B, Smithson A, Slik FJW, Schuiteman A. 2004.** Epiphytism and pollinator specialization: drivers for orchid diversity?. *Philosophical Transactions of the Royal Society of London Series B Biological Sciences* 359:1523–1535 DOI 10.1098/rstb.2004.1529.
- Haber WA, Frankie GW. 1989.** A tropical hawkmoth community: costa Rican dry forest Sphingidae. *Biotropica* 21:155–172 DOI 10.2307/2388706.
- Hadly EA, Spaeth PA, Li C. 2009.** Niche conservatism above the species level. *Proceedings of the National Academy of Sciences of the United States of America* 106(suppl. 2):19707–19714 DOI 10.1073/pnas.0901648106.
- Harvey PH, Pagel M. 1991.** *The comparative method in evolutionary biology*. Oxford: Oxford University Press.
- Hawkins BA, Rodríguez MA, Weller SG. 2011.** Global angiosperm family richness revisited: linking ecology and evolution to climate. *Journal of Biogeography* 38:1253–1266 DOI 10.1111/j.1365-2699.2011.02490.x.
- Hawkins BA, Rueda M, Rangel TF, Field R, Diniz-Filho AF. 2014.** Community phylogenetics at the biogeographical scale: cold tolerance, niche conservatism and the structure of North American forests. *Journal of Biogeography* 41:23–38 DOI 10.1111/jbi.12171.
- Heibl C, Calenge C. 2013.** Phyloclim: integrating phylogenetics and climatic niche modeling. R package version 0.9-4. Available at <http://CRAN.R-project.org/package=phyloclim>.
- Hijmans RJ, Schreuder M, Cruz J, Guarino L. 1999.** Using GIS to check co-ordinates of genebank accessions. *Genetic Resources and Crop Evolution* 46:291–296 DOI 10.1023/A:1008628005016.
- Hoorn C, Guerrero J, Sarmiento GA, Lorente MA. 1995.** Andean tectonics as a cause for changing drainage patterns in Miocene northern South America. *Geology* 23:237–240 DOI 10.1130/0091-7613(1995)023<0237:ATAACF>2.3.CO;2.

- Hoorn C, Wesselingh FP, Ter Steege H, Bermudez MA, Mora A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo J, Jaramillo C, Riff D, Negri FR, Hooghiemstra H, Lundberg J, Stadler T, Sarkinen T, Antonelli A. 2010. Amazonia through time: andean uplift, climate change, landscape evolution and biodiversity. *Science* 330:927–931 DOI 10.1126/science.1194585.
- Jukes T, Cantor C. 1969. Evolution of protein molecules. In: *Mammalian protein metabolism*. New York: Academic Press, 21–132.
- Katoh K, Toh H. 2008. Recent developments in the MAFFT multiple sequence alignment program. *Briefings in Bioinformatics* 9:286–298 DOI 10.1093/bib/bbn013.
- Kerr JT, Pindar A, Galpern P, Packer L, Potts SG, Roberts SM, Rasmont P, Schweiger O, Colla SR, Richardson LL, Wagner DL, Gall LF, Sikes DS, Pantoja A. 2015. Climate change impacts on bumblebees converge across continents. *Science* 349:177–180 DOI 10.1126/science.aaa7031.
- Khalid I, Fritz SA, Prinzing R, Pfenninger M, Böhning-Gaese K, Hof C. 2015. Global variation in thermal physiology of birds and mammals: evidence for phylogenetic niche conservatism only in the tropics. *Journal of Biogeography* 42:2187–2196 DOI 10.1111/jbi.12573.
- Khudoley K, Meyerhoff AA. 1971. Paleogeography and geology history of the Greater Antilles. *Geological Society of America Memoirs* 129:1–199 DOI 10.1130/MEM129-p1.
- Kolanowska M. 2013. Niche conservatism and the future potential range of *Epipactis helleborine* (Orchidaceae). *PLOS ONE* 8:e77352 DOI 10.1371/journal.pone.0077352.
- Kolanowska M, Konowalik K. 2014. Niche conservatism and future changes in the potential area coverage of *Arundina graminifolia*, an invasive orchid species from Southeast Asia. *Biotropica* 46:157–165 DOI 10.1111/btp.12089.
- Kolanowska M, Mystkowska K, Kras M, Dudek M, Konowalik K. 2016. Evolution of the climatic tolerance and postglacial range changes of the most primitive orchids (Apostasioideae) within Sundaland, Wallacea and Sahul. *PeerJ* 4:e2384 DOI 10.7717/peerj.2384.
- Kolanowska M, Pérez Escobar O, Parra Sánchez E. 2012. A new species of *Campylocentrum* (Orchidaceae: angraecinae) from Colombia. *Lankesteriana* 12:9–11 DOI 10.15517/lank.v12i1.18269.
- Kolanowska M, Szlachetko DL. 2013. New species of the genus *Campylocentrum* (Orchidaceae, Vandeeae) from Colombia. *Phyton* 53:221–239.
- Kolanowska M, Szlachetko DL. 2014. Niche conservatism of *Eulophia alta*, a trans-Atlantic orchid species. *Acta Societatis Botanicorum Poloniae* 83:51–57 DOI 10.5586/asbp.2014.007.
- Kozak KH, Wiens JJ. 2006. Does niche conservatism promote speciation? A case study in North American salamanders. *Evolution* 60:2604–2621 DOI 10.1111/j.0014-3820.2006.tb01893.x.
- Kriticos DJ, Webber BL, Leriche A, Ota N, Macadam I, Bathols J, Scott JK. 2012. CliMond: global high resolution historical and future scenario climate surfaces for bioclimatic modelling. *Methods in Ecology and Evolution* 3:53–64 DOI 10.1111/j.2041-210X.2011.00134.x.

- Lobo JM, Jiménez-Valverde A, Real R. 2008.** AUC: a misleading measure of the performance of predictive distribution models. *Global Ecology and Biogeography* 17:145–151 DOI [10.1111/j.1466-8238.2007.00358.x](https://doi.org/10.1111/j.1466-8238.2007.00358.x).
- Mason SJ, Graham NE. 2002.** Areas beneath the relative operating characteristics (ROC) and relative operating levels (ROL) curves statistical significance and interpretation. *Quarterly Journal of the Royal Meteorological Society* 128:2145–2166 DOI [10.1256/003590002320603584](https://doi.org/10.1256/003590002320603584).
- Micheneau C, Carlswald BS, Fay MF, Bytebier B, Paillet T, Chase MW. 2008.** Phylogenetics and biogeography of Mascarene angraecoid orchids. *Molecular Phylogenetics and Evolution* 46:908–922 DOI [10.1016/j.ympev.2007.12.001](https://doi.org/10.1016/j.ympev.2007.12.001).
- Micheneau C, Fournel J, Paillet T. 2006.** Bird pollination in an angraecoid orchid on Reunion Island (Mascarene Archipelago, Indian Ocean). *Annals of Botany* 97:965–974 DOI [10.1093/aob/mcl056](https://doi.org/10.1093/aob/mcl056).
- Micheneau C, Fournel J, Warren BH, Hugel S, Gauvin-Bialecki A, Paillet T, Strasberg D, Chase MW. 2010.** Orthoptera, a new order of pollinator. *Annals of Botany* 105:355–364 DOI [10.1093/aob/mcp299](https://doi.org/10.1093/aob/mcp299).
- Morinière J, Van Dam MH, Hawlitschek O, Bergsten J, Michat MC, Hendrich L, Ribera I, Toussaint EFA, Balke M. 2016.** Phylogenetic niche conservatism explains an inverse latitudinal diversity gradient in freshwater arthropods. *Scientific Reports* 6:26340 DOI [10.1038/srep26340](https://doi.org/10.1038/srep26340).
- Nilsson LA, Jonsson L, Rason L, Randrianjohany E. 1985.** Monophily and pollination mechanisms in *Angraecum arachnites* Schltr. (Orchidaceae) in a guild of long-tongued hawk-moths (Sphingidae) in Madagascar. *Biological Journal of the Linnean Society* 91:1–19 DOI [10.1111/j.1095-8339.1985.tb01130.x](https://doi.org/10.1111/j.1095-8339.1985.tb01130.x).
- Olalla-Tárraga MA, McInnes L, Bini LM, Diniz-Filho JAF, Fritz SA, Hawkins BA, Hortal J, Orme CDL, Rahbek C, Rodríguez MA, Purvis A. 2011.** Climatic niche conservatism and the evolutionary dynamics in species range boundaries: global congruence across mammals and amphibians. *Journal of Biogeography* 38:2237–2247 DOI [10.1111/j.1365-2699.2011.02570.x](https://doi.org/10.1111/j.1365-2699.2011.02570.x).
- Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GVN, Underwood EC, D'Amico JA, Itoua I, Strand HE, Morrison JC, Loucks CJ, Allnutt TF, Ricketts TH, Kura Y, Lamoreux JF, Wettengel WW, Hedao P, Kassem KR. 2001.** Terrestrial ecoregions of the world: a new map of life on Earth. *Bioscience* 51(11):933–938 DOI [10.1641/0006-3568\(2001\)051\[0933:TEOTWA\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2001)051[0933:TEOTWA]2.0.CO;2).
- Otero JT, Ackerman JD, Bayman P. 2002.** Diversity and host specificity of endophytic *Rhizoctonia*-like fungi from tropical orchids. *American Journal of Botany* 89:1852–1858 DOI [10.3732/ajb.89.11.1852](https://doi.org/10.3732/ajb.89.11.1852).
- Otero JT, Flanagan NS. 2006.** Orchid diversity: beyond deception. *Trends in Ecology and Evolution* 21:64–65 DOI [10.1016/j.tree.2005.11.016](https://doi.org/10.1016/j.tree.2005.11.016).
- Paradis E, Claude J, Strimmer K. 2004.** APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* 20:289–290 DOI [10.1093/bioinformatics/btg412](https://doi.org/10.1093/bioinformatics/btg412).
- Paulus HF, Gack C. 1990.** Pollinators as prepollinating isolation factors: evolution and speciation in *Ophrys* (Orchidaceae). *Israel Journal of Botany* 39:43–79.

- Pearman PB, Lavergne S, Roquet C, Wüest R, Zimmermann NE, Thuiller W. 2014.** Phylogenetic patterns of climatic, habitat and trophic niches in a European avian assemblage. *Global Ecology and Biogeography* **23**(4):414–424 DOI [10.1111/geb.12127](https://doi.org/10.1111/geb.12127).
- Peirce CS. 1884.** The numerical measure of the success of predictions. *Science* **4**:453–454.
- Phillips SJ, Anderson R, Schapire RE. 2006.** Maximum entropy modeling of species geographic distributions. *Ecological Modelling* **190**:231–259 DOI [10.1016/j.ecolmodel.2005.03.026](https://doi.org/10.1016/j.ecolmodel.2005.03.026).
- Phillips SJ, Dudík M, Schapire RE. 2004.** A maximum entropy approach to species distribution modeling. In: *ICML '04 proceedings of the twenty-first international conference on machine learning*. New York: ACM, pp 655–662.
- Pridgeon AM, Cribb PJ, Chase MW, Rasmussen FN. 2014.** Genera orchidacearum. In: *Epidendroideae (Part three)*. Vol. 6. Oxford: Oxford University Press.
- Prinzing A, Durka W, Klotz S, Brandl R. 2001.** The niche of higher plants: evidence for phylogenetic conservatism. *Proceedings of the Royal Society B: Biological Sciences* **268**(1483):2383–2389 DOI [10.1098/rspb.2001.1801](https://doi.org/10.1098/rspb.2001.1801).
- Pupulin F. 1998.** Orchid florula of Parque Nacional Manuel Antonio, Quepos, Costa Rica. *Revista de Biología Tropical* **46**:961–1031.
- Pupulin F. 2001.** Addenda orchidaceis quepoanis. *Lankesteriana* **1**:1–28 DOI [10.15517/lank.v1i1.23149](https://doi.org/10.15517/lank.v1i1.23149).
- R Core Team. 2014.** *R: a language and environment for statistical computing*. Vienna: R Foundation for Statistical Computing.
- Rato C, Harris DJ, Perera A, Carvalho SB, Carretero MA, Rödder D. 2015.** A combination of divergence and conservatism in the niche evolution of the Moorish Gecko, *Tarentola mauritanica* (Gekkota: Phyllodactylidae). *PLOS ONE* **10**(5):e0127980 DOI [10.1371/journal.pone.0127980](https://doi.org/10.1371/journal.pone.0127980).
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012.** MrBayes 3.2: efficient bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* **61**(3):539–542 DOI [10.1093/sysbio/sys029](https://doi.org/10.1093/sysbio/sys029).
- Sanderson MJ. 2002.** Estimating absolute rates of molecular evolution and divergence times: a penalized likelihood approach. *Molecular Biology and Evolution* **19**:101–109 DOI [10.1093/oxfordjournals.molbev.a003974](https://doi.org/10.1093/oxfordjournals.molbev.a003974).
- Schiestl FP. 2005.** On the success of a swindle: pollination by deception in orchids. *Naturwissenschaften* **92**:255–264 DOI [10.1007/s00114-005-0636-y](https://doi.org/10.1007/s00114-005-0636-y).
- Schoener TW. 1968.** The anolis lizards of bimini: resource partitioning in a complex fauna. *Ecology* **49**:704–726 DOI [10.2307/1935534](https://doi.org/10.2307/1935534).
- Serra-Varela MJ, Grivet D, Vincenot L, Broennimann O, Gonzalo-Jiménez J, Zimmermann NE. 2015.** Does phylogeographical structure relate to climatic niche divergence? A test using maritime pine (*Pinus pinaster* Ait.). *Global Ecology and Biogeography* **24**:1302–1313 DOI [10.1111/geb.12369](https://doi.org/10.1111/geb.12369).
- Singer RB. 2003.** Orchid pollination: recent developments from Brazil. *Lankesteriana* **7**:111–114.

- Singer RB, Cocucci AA. 1999.** Pollination mechanism in southern Brazilian orchids which are exclusively or mainly pollinated by halictid bees. *Plant Systematics and Evolution* **217**:101–117 DOI [10.1007/BF00984924](https://doi.org/10.1007/BF00984924).
- Soberón J, Peterson AT. 2005.** Interpretation of models of fundamental ecological niches and species' distributional areas. *Biodiversity Informatics* **2**:1–10 DOI [10.17161/bi.v2i0.4](https://doi.org/10.17161/bi.v2i0.4).
- Swenson NG. 2010.** The role of evolutionary processes in producing biodiversity patterns, and the interrelationships between taxonomic, functional and phylogenetic biodiversity. *American Journal of Botany* **98**(3):472–480 DOI [10.3732/ajb.1000289](https://doi.org/10.3732/ajb.1000289).
- Swofford DL. 2002.** PAUP* Phylogenetic analysis using parsimony (*and other methods). Version 4. Sunderland: Sinauer Associates..
- Szlachetko DL, Kolanowska M. 2013a.** *Campylocentrum guarinae* sp. nov. (Vandaeae, Orchidaceae) from Colombia. *Nordic Journal of Botany* **32**:285–288.
- Szlachetko DL, Kolanowska M. 2013b.** Two new species and a new combination in *Campylocentrum* (Orchidaceae, Vandaeae) from Guyana. *Annales Botanici Fennici* **50**:263–268 DOI [10.5735/086.050.0409](https://doi.org/10.5735/086.050.0409).
- Szlachetko DL, Tukałło P, Mytnik-Ejsmont J, Grochocka E. 2013.** Reclassification of the *Angraecum*-alliance (Orchidaceae, Vandoideae) based on molecular and morphological data. *Biodiversity Research and Conservation* **29**:1–23 DOI [10.2478/biorc-2013-0004](https://doi.org/10.2478/biorc-2013-0004).
- Thiers B. 2015.** *Index herbariorum: a global directory of public herbaria and associated staff*. New York Botanical Garden's Virtual Herbarium. Available at <http://sweetgum.nybg.org/ih/>.
- Uba CE, Strecker MR, Schmitt AK. 2007.** Increased sediment accumulation rates and climatic forcing in the central Andes during the late Miocene. *Geology* **35**:979–982 DOI [10.1130/G224025A.1](https://doi.org/10.1130/G224025A.1).
- Urbina-Cardona JN, Loyola RD. 2008.** Applying niche-based models to predict endangered-hyloid potential distributions: are neotropical protected areas effective enough? *Tropical Conservation Science* **1**:417–445 DOI [10.1177/194008290800100408](https://doi.org/10.1177/194008290800100408).
- Van der Vaart AW. 1998.** *Asymptotic statistics*. Cambridge: Cambridge Univ. Press.
- Varela S, Anderson RP, García-Valdés R. 2014.** Environmental filters reduce the effects of sampling bias and improve predictions of ecological niche models. *Ecography* **37**:1084–1091 DOI [10.1111/j.1600-0587.2013.00441.x](https://doi.org/10.1111/j.1600-0587.2013.00441.x).
- Warren DL. 2016.** ENMTools: analysis of niche evolution using niche and distribution models. R package version 01.
- Warren DL, Glor RE, Turelli M. 2008.** Environmental niche equivalency versus conservatism: quantitative approaches to niche evolution. *Evolution* **62**:2868–2883 DOI [10.1111/j.1558-5646.2008.00482.x](https://doi.org/10.1111/j.1558-5646.2008.00482.x).
- Warren DL, Glor RE, Turelli M. 2010.** ENMTools: a toolbox for comparative studies of environmental niche models. *Ecography* **33**:607–611 DOI [10.1111/j.1600-0587.2009.06142.x](https://doi.org/10.1111/j.1600-0587.2009.06142.x).

- Wasof S, Lenoir J, Aarrestad PA, Alsos IG, Armbruster WS, Austrheim G, Bakkestuen V, Birks HJB, Bråthen KA, Broennimann O, Brunet J, Bruun HH, Dahlberg CJ, Diekmann M, Dullinger S, Dynesius M, Ejrnæs R, Gégout J-C, Graae BJ, Grytnes J-A, Guisan A, Hylander K, Jónsdóttir IS, Kapfer J, Klanderud K, Luoto M, Milbau A, Moora M, Nygaard B, Odland A, Pauli H, Ravolainen V, Reinhardt S, Sandvik SM, Schei FH, Speed JDM, Svenning J-C, Thuiller W, Tveraabak LU, Vandvik V, Velle LG, Virtanen R, Vittoz P, Willner W, Wohlgemuth T, Zimmermann NE, Zobel M, Decocq G. 2015. Disjunct populations of European vascular plant species keep the same climatic niches. *Global Ecology and Biogeography* 24:1401–1412 DOI 10.1111/geb.12375.
- Wellenreuther M, Larson KW, Svensson EI. 2012. Climatic niche divergence or conservatism? Environmental niches and range limits in ecologically similar damselflies. *Ecology* 93:1353–1366 DOI 10.1890/11-1181.1.
- Wiens JJ. 2004. Speciation and ecology revisited: phylogenetic niche conservatism and the origin of species. *Evolution* 58:193–197 DOI 10.1111/j.0014-3820.2004.tb01586.x.
- Wiens JJ, Donoghue MJ. 2004. Historical biogeography, ecology, and species richness. *Trends in Ecology & Evolution* 19:639–644 DOI 10.1016/j.tree.2004.09.011.
- Wiens JJ, Graham CH. 2005. Niche conservatism: integrating evolution, ecology, and conservation biology. *Annual Review of Ecology, Evolution, and Systematics* 36:519–539 DOI 10.1146/annurev.ecolsys.36.102803.095431.
- Young ND, Healy J. 2003. GapCoder automates the use of indel characters in phylogenetic analysis. *BMC Bioinformatics* 4:6 DOI 10.1186/1471-2105-4-6.