Taxonomy of the genus *Passerina* (Thymelaeaceae)

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Keywords: anatomy, cladistics, endemism, macromorphology, new species, palynology, Passerina, southern Africa, taxonomy, Thymelaeaceae

ABSTRACT

Passerina L. is mainly a southern African genus, comprising 20 species and four subspecies. A few species occur along the Great Escarpment, two extend into Zimbabwe and Mozambique, but most are concentrated in the Cape Floristic Region. Palynological, macromorphological and anatomical evidence was used in the delimitation of the genus and its infrageneric taxa. A cladistic study supports Passerina as a monophyletic genus. A genus treatment, key to species and a full species treatment are given. Each species treatment includes a taxonomic diagnosis, description and notes on taxonomy, etymology, economic value and distribution. Illustrations of representative species are provided and distribution maps are included for each species. P. esterhuyseniae Bredenk. & A.E.van Wyk is newly described. A list of excluded species names highlights the previous cosmopolitan taxonomic interpretation of Passerina, as many names are now in synonymy under other genera of the Thymelaeaceae.

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INTRODUCTION

Thymelaeaceae

Applying the sexual system of classification, Linnaeus (1754) placed the genus Passerina under Class VIII, Octandria, 1. Monogynia. This system was followed until De Jussieu (1789) instated the family Thymelaeaceae, with the following genera: Dirca L., Lagetta Juss., Daphne L., Passerina L., Stellera L., Struthiola L. Lachnaea L., Dais L., Gnidia L., Nectandra Berg. and Quisqualis L. Wikström (1818) accepted the Thymelaeaceae, but based the infrafamilial classification on the number of stamens, following Linnaeus. The most important contributions towards the infrafamilial classification of the Thymelaeaceae, based on morphological characters, were made by Endlicher (1847), Meisner (1857), Bentham & Hooker (1880) and Gilg (1894a). Using anatomical characters, further contributions were made by Van Tieghem (1893), Gilg (1894b) and Leandri (1930).

Domke (1934) proposed a widely adopted subfamilial classification for the Thymelaeaceae and divided the family into four subfamilies, namely Gonystyloideae, Aquilarioideae, Gilgiodaphnoideae and Thymelaeoideae. The genus *Passerina* is classified under the Thymelaeoideae. Based on palynological evidence Archangelsky (1971: fig. 10) added the new subfamilies Octolepidoideae, Microsem-

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matoideae and Synadrodaphnoideae and raised the Gonystyloideae to the family Gonystylaceae (also recognized by Takhtajan 1997, amongst others). Bredenkamp & Van Wyk (1996) published new evidence on the structure of the pollen wall in *Passerina* resulting in the elevation of the subtribe Passeriniae Endl. to the monogeneric tribe Passerineae (Endl.) Bredenk. & A.E.van Wyk. Evidence obtained from floral morphology, anatomy, embryology and palynology indicates that the Thymelaeaceae has a strong malvalean relationship, an affinity also supported by molecular data (APG 1998; Magallón *et al.* 1999). The possible phylogenetic relationships of the Thymelaeaceae are discussed by Bredenkamp & Van Wyk (2001b).

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The Thymelaeaceae is currently considered a family of \pm 58 genera and \pm 720 species. (Mabberley 1989; Brummitt 1992; Takhtajan 1997). It is subcosmopolitan and the distribution of the genera is listed by Mabberley (1989), as follows:

Africa: temperate southern Africa—Dais L., Englero-daphne Gilg, Gnidia L., Lachnaea L., Passerina L., Peddiea Harv., Struthiola L., Synaptolepis Oliv.; tropical Africa—Craterosiphon Engl. & Gilg, Dicranolepis Planch., Octolepis Oliv., Synandrodaphne Gilg.

Asia: Aetoxylon Airy Shaw, Amyxa Tiegh., Drapetes Lam., Eriosolena Blume, Pentathymelaea Lecomte, Rhamnoneuron Gilg, Restella Pobed., Wikstroemia Endl.

Australia: Arnhemia Airy Shaw, Drapetes Lam., Pimelea Banks & Sol., Oreodendron C.T.White.

Europe: Daphne L., Diarthron Turcz.

Japan: Daphnimorpha Nakai, Edgeworthia Meisn.

Madagascar: Stephanodaphne Baill.

Malesia: Aquilaria Lam., Enkleia Griff., Gonystylus Teijsm. & Binn., Linostoma Wall. ex Endl., Phaleria Jack.

Mediterranean region: Thymelaea Mill.

New Caledonia: Deltaria Steenis, Lethedon Spreng., Solmsia Baill.

North and South America: Daphnopsis Mart. & Zucc., Dirca L., Funifera Leandro ex C.A.Mey., Goodallia Benth., Lagetta Juss., Lasiadenia Benth., Linodendron Griseb., Lophostoma Meisn., Ovidia Meisn., Schoenobiblus Mart.

Sri Lanka: Gyrinops Gaertn.

Perhaps the economically most important character in the family is its tough fibrous bark. The bark of Wikstroemia, Daphne, Edgeworthia, Thymelaea and Daphnopsis is used for rope, and in the manufacturing of bank notes and strong paper. Flexible shoots of Dirca are used for baskets. Bark of Pimelea was used as a source of twine by early settlers in Australia.

Many genera are also known for their medicinal value. The wood of *Wikstroemia* is a source of incense and that of *W. ovata* C.A.Mey. is a strong purge. In China the bark of *Daphne* is used as an apparently safe and efficient abortifacient; it contains the glycoside daphnin and

an acrid resin (mezerein) giving plants a bitter taste. The decaying heartwood of *Aquilaria malaccensis* Lam. is saturated with a resin which is the basis of incense and when distilled it is used in perfume and medicine.

The genera *Pimelea*, *Edgeworthia* and *Daphne* are cultivated for horticultural purposes. The scent of *Daphne* flowers is carnation-like and attractive to Lepidoptera; some members are moth-pollinated. *Gonystylus bancanus* (Miq.) Kurz. is a peat swamp-forest tree, with knee-roots. Its lightweight commercial timber is used for dowelling and is much exported from Indomalesia.

In southern Africa, the bark of various genera is used for tying down thatch, for plaiting into whip thongs and for twine. *Dais cotinifolia* L. is an ornamental tree with attractive flowers, occurring mostly along the eastern regions of the country.

Passerina

In his comprehensive work on the circumscription of the Thymelaeaceae and infrafamilial taxa, Domke (1934) gave a complete historical review of the intergeneric classification of Passerina. He included the southern African genera Dais, Gnidia (= Lasiosiphon), Struthiola, Lachnaea (= Cryptadenia) and Passerina in the tribe Gnidieae, subtribe Gnidiinae of the subfamily Thymelaeoideae. Bredenkamp & Van Wyk (1996) place Passerina in the monogeneric tribe Passerineae on the basis of mainly pollen characters. Currently Passerina is considered advanced at the intergeneric level, as many of the advanced character states present in other genera of the Thymelaeoideae are all found together in this genus. The most prominent characters distinguishing Passerina are the exserted, extrorse anthers and the unique anemophilous habit (Bredenkamp & Van Wyk 1996, 2001b).

The infrageneric classification of *Passerina* is documented by Linnaeus (1753) in his *Species plantarum*, in which he described *P. filiformis*, *P. hirsuta*, *P. ciliata* and *P. uniflora*. *P. filiformis* is the only species that is currently recognized in *Passerina*. Publications mentioned in the applicable protologue and in synonymy to the various species that pre-date the nomenclatural starting point for the Spermatophyta [International Code of Botanical Nomenclature, Article 13.1 (Greuter *et al.* 2000)] are Linnaeus's *Hortus Cliffortianus* (1737), Van Royen (1740), Plukenet (1700: 180), Breyne (1678) and Burman (1739). The generic name *Passerina* appearing in *Species plantarum* (Linnaeus 1753) is associated with the subsequent description given in *Genera plantarum* (Linnaeus 1754) (Greuter *et al.* 2000, Article 13.4).

Wikström (1818) recognized 41 species of *Passerina* and the subspecies *P. filiformis* subsp. *divaricata*; of these only four species are presently recognized in *Passerina*. In the interim the subspecies was raised to species level and is presently known as *P. falcifolia*. Thunberg (1825a) recognized nine species of which only one is currently maintained. His concept of *P. glomerata*, *P. ericoides* and *Lachnaea conglomerata* were completely incorrect and caused confusion right up to the present study. Meisner (1840; 1857: 563–565) redefined the genus by clarifying 92 'species exclusae' which were mostly synonymous with

other cosmopolitan genera in the Thymelaeaceae and he retained only four species and six subspecies. The distribution of the remaining species clearly indicated that Passerina was a smaller genus, largely confined to southern Africa. At the beginning of the 20th century, Wright (1915) revised the Thymelaeaceae for the Flora capensis and his generic concept of Passerina was mostly based on that of Meisner (1857). He recognized ten species, of which three were new, as well as three subspecies. He recognized P. ericoides and Chymococca empetroides. We agree with Thoday (1924a) that *C. empetroides* is a synonym of *P. eri*coides. Although Thoday (1924a) provided a much improved classification of the group, the circumscription and identification of several species remained problematic, especially in the herbarium. Table 1 is a summary of taxa recognized in the most comprehensive works on Passerina from Linnaeus (1753) to the present study.

In his treatment of *Passerina*, Meisner (1840) divided the genus into section I. *Pentamerae* and section II. *Tetramerae*. *P. polycephala* E.Mey., *P. anthylloides* L.f. and *P. calocephala* Meisn., with pentamerous flowers (section I), were eventually all placed in the genus *Gnidia* (Meisner 1857; Gilg 1894a). Meisner (1857) did not divide *Passerina* into infrageneric taxa, a pattern followed by all subsequent treatments and no further mention was made of the relevant sections. In the present genus treatment the sectional classification is not maintained.

MATERIALS AND METHODS

Material from the following herbaria was studied (acronyms according to Holmgren *et al.* 1990): BM, BOL, BREM, C, GRA, K, LINN, M, MEL, MO, NBG, P, PR, PRC, PRE, PRU, S, SBT, TCD, UPS, W, WU. A database of the specimens was compiled on the Microsoft Access Relational Database Management System for Windows, Version 2.0.

Live and preserved (dried and in liquid preservatives) material of all the species and subspecies in *Passerina* was studied. As far as possible, material was collected from at least five different localities for every taxon. Illustrations were made from herbarium material by means of a drawing tube. Measurements were taken using a dissection microscope and a calibrated eyepiece. Because the laminas of most leaves and floral bracts are cymbiform or rolled, the depth was measured, with dimensions indicated as length × depth.

Light microscopy (LM) was used for general leaf anatomy, epidermal studies and floral anatomy (Bredenkamp & Van Wyk 1999, 2001a, 2001b). Leaf and floral material was fixed and stored in a 0.1 M phosphate-buffered solution at pH 7.4, containing 2.5% formaldehyde, 0.1% glutaraldehyde and 0.5% caffeine [modified Karnovsky fixative; Karnovsky (1965)]. The material was washed in water, dehydrated and embedded in glycol methacrylate (GMA) following the methods of Feder & O'Brien (1968). Embedded material was serially sectioned. Sections were stained in toluidine blue 'O', subjected to the periodic acid-Schiff's (PAS) reaction and mounted in Entellan (Art. 7961, E. Merck, Darmstadt).

Scanning electron microscopy (SEM) was used to study the epidermal surface features (including epicuticular waxes) and to verify the structure of the cuticle (Bredenkamp & Van Wyk 2000).

Transmission electron microscopy (TEM) was used for the study of the structure of mucilaginous epidermal cell walls in *Passerina* (Bredenkamp & Van Wyk 1999).

Terminology used in the descriptions of inflorescences and flowers is mentioned in Bredenkamp & Van Wyk (2001b). General descriptive terminology follows Stearn (1973) and Radford *et al.* (1974). Author citations follow Brummitt & Powell (1992).

Passerina L. Species plantarum: 559 (1753); L.: 168 (1754); P.J.Bergius: 126 (1767); Mill.: (1768); Burm.f.: 12 (1768); L.: 236 (1771); L.: 225 (1782); L.: 374 (1784); Thunb.: 75 (1794); J.C.Wendl.: 18 (1798); Willd.: 429 (1799); Poir.: 39 (1804): Lam. & DC.: 359 (1805); Wikstr.: 319 (1818); Thunb.: 374 (1825a); Meisn.: 390 (1840); Steud.: 273 (1841); C.A.Mey.: 45 (1843); Meisn.: 561 (1857); Harv.: 325 (1868); Gand.: 418 (1913); C.H.Wright: 9 (1915); Thoday: 146 (1924a); Marloth: 214 (1925); Domke: 137 (1934); Palmer & Pitman: 1583 (1972); Coates Palgrave: 648 (1977); Bond & Goldblatt: 432 (1984); Hilliard & B.L.Burtt: 182 (1987); Goldblatt & Manning: 683 (2000). Type species: Passerina filiformis L.

Sanamunda [Clus.: 89 (1601); L.: 146 (1737)] Adans.: 258 (1763); Lam. & DC.: 359 (1805); Raf.: 104 (1836). Type species: as above.

Thymelaea [Tourn.: 594 (1719); L.: 146 (1737)] Adans.: 258 (1763); Juss.: 77 (1789); Lam. & DC.: 359 (1805). Type species: *Daphne laureola* L.

Passerine Lam. & DC.: 359 (1805) orth. var.

Balendasia Raf.: 105(1836). Type species: B. ericoides (Burm.f.) Raf.

Steiroctis Raf.: 105 (1836). Type species: not designated (Farr et al. 1979).

Trimeiandra Raf.: 105 (1836). Type species: *T. spicata* Raf. nom. illeg.

Lonchostoma obtusiflorum Wikstr. nom illeg. = Passerina pentandra Thunb. (Farr et al. 1979) ≡ Lonchostoma Wikstr.: 350 (1818) nom. cons.

Chymococca Meisn.: 565 (1857); Harv.: 325 (1868); Benth. & Hook.: 194 (1880); Thoday: 166 (1924a). Type species: *C. empetroides* Meisn.

Passerina L. Sectio Pentamerae Meisn.: 390 (1840). Type species: not designated.

Passerina L. Sectio Tetramerae Meisn.: 395 (1840). Type species: not designated.

⁼ the identity sign denoting nomenclatural synonymy for names based on the same type species.

TABLE 1.—A summary of taxa in the most comprehensive works on Passerina from Linnaeus (1753) to the present study

Linnaeus (1753)	Wikström (1818)	Thunberg (1825a)	Meisner (1857)	Wright (1915)	Thoday (1924a)	Present study
P. filiformis <i>L.</i>	P. filiformis L.	P. filiformis L.	P. filiformis L.	P. filiformis L.	P. filiformis L.	P. filiformis L. (= P. cupressina J.C.Wendl. nom. nud.)
			(= P. pectinata)	(= P. pectinata	(= P. pectinata nom. nud.)	(= P. pectinata Lodd. nom. nud.)
			nom. nud.)	nom. nud.)		subsp. filiformis subsp. glutinosa (Thoday) Bredenk. & A F yom Web.
			P. filiformis L. α vulgaris Meisn.	P. corymbosa Eckl. ex C.H.Wright	P. vulgaris Thoday	P. corymbosa Eckl. ex C.H. Wright
					(= P. filiformis L. var. vulgaris Meisn.)	(= P. corymbosa Eckl. ex Meisn)
	P. filiformis <i>L.</i> β divaricata <i>Wikstr.</i>		P. filiformis L. β falcifolia Meisn.	P. falcifolia C.H.Wright	P. falcifolia C.H.Wright	P. falcifolia (Meisn.) C.H. Wright
					(= P . filiformis L. var. divaricata Wikstr.) (= P . filiformis L. var. falcifolia Meisn.)	(= P. filiformis L. var. divaricata Wikstr.) (= P. filiformis L. var. folcifolia Meisn.)
			P. filiformis <i>L.</i> γ comosa <i>Meisn</i> .	P. comosa C.H.Wright	P. comosa C.H. Wright	P. comosa (Meisn.) C.H.Wright (= P. filiformis L. var. comosa Meisn.)
			P. filiformis L 8 squarrosa Meisn	P. rubra C.H.Wright	P. rubra C.H.Wright	P. rubra C.H.Wright
P. hirsuta L.	P. hirsuta L. (= P. metnan Forssk.)		Thymelaea hirsuta <i>Endl.</i> (= <i>P. hirsuta</i> L.) (= <i>P. metmon</i> Eoreck)		(= F. Hiljorniis L. Var. squarrosa Metsn.)	(= P. filiformis L. var. squarrosa Meisn.)
P. ciliata <i>L.</i>	P. ciliata L.	P. ciliata Thunb. (= P. ciliata L.)	Cryptachia official Meisn. (= P. ciliata Thunb.) Gnida? ciliata Meisn. (= P. ciliata L.)			
P. uniflora L.	P. uniflora L.	P. uniflora L.	Cryptadenia uniflora <i>Meisn</i> . $(=P, uniflora L.)$			
Lachnaea conglomerata L.	P. conglomerata Thumb. (= L. conglomerata L.) (= P. glomerata Thumb.)	P. glomerata Thumb. (= L. conglomerata L.) (= P. ericoides L.)	L. conglomerata L.		P. glomerata Thunb. (= L. conglomerata L.)	L. conglomerata L. nomen rejiciendum
	P. paleacea Wikstr. (= Lachnaea paleacea fide Wikstr.)		P. paleacea Wikstr. (= Lachnaea paleacea fide Wikstr.)	P. paleacea Wikstr. (= Lachnaea paleacea fide Wikstr.)	P. paleacea <i>Wikstr.</i> (= Lachnaea paleacea Wikstr.) (= P. glomerata fide Thunb.)	P. paleacea <i>Wikstr.</i> (= Lachnaea paleacea fide Wikstr.) (= P. glomerata sensu Thunb.)
	P. ericoides L.		P. ericoides L.	P. ericoides L.	P. ericoides L. (= Chymococca empetroides Meisn.) (= P. filiformis var. crassifolia Fek! & Zevh fide Meisn.)	(= P. ericoides sensu Thunb.) P ericoides L. (= Chymococca empetroides Meisn.) (= P. filljornis L. var. crassifolia E-P. & Zash Esh Meisn.)
			(= P. glomerata Thunb.) (= Lachuaea conglomerata L.)	(= P. glomerata Thunb.) (= Lachnaea conglomerata L.)		(=P. glomerata sensu Meisn.)
	P. rigida <i>Wikstr</i> .		P. rigida <i>Wikstr</i> .	P. rigida Wikstr.	P. rigida <i>Wikstr</i> .	P. rigida Wikstr. (= P. eriophora Gand.) (= P. ericoides sensu Meisn.)

TABLE 1.—A summary of taxa in the most comprehensive works on Passerina from Linnaeus (1753) to the present study (cont.)

P. rigida Wikstr. P. rigida Wikstr. var. transcata Meisn. P. rigida Wikstr. var. transcata Wikstr. P. rigida Wikstr. var. transcata Meisn. P. rigida Wikstr. var. transcata Meisn. P. rigida Wikstr. var. transcata Wikstr. P. rigida Wikstr. var. transcata Wikstr. var. transcata Wikstr. P. rigida Wikstr. var. transcata Wikstr. var. transcata Wikstr. P. rigida Wikstr. var. transcata Wikstr. var. transcata Wikstr. var. transcata Wikstr. var. transcata Wikstr. var. tra	t Meisn. pro parte) Meisn. pro parte) Meisn. pro parte) truncata Meisn.) tetragona Meisn.	P. pendula Eckl. & Zeyh, ex Thodav (= P. rigida Wikstr, var. comosa Meisn.) P. burchellii Thodav (= P. tetragema fide Thodav) P. truncata (Meisn.) Bredenk. & A.E.van W (= P. rigida Wikstr. var. truncata Meisn.) (= P. rigida Wikstr. var. tetragona Meisn.) (= P. glomerata sphalm. quoad L. conglomera L. sensu Thunb.) (= Lachnaea glomerata E. sensu Thunb.) (= P. glomerata L. sensu Thunb.) (= P. glomerata Thunb.)
str. P. rigida Wikstr. var. str. P. rigida Wikstr. P. rigida Wikstr. P. rigida Wikstr. B. truncata Meisn. Chymococca Chymococca Chymococca Chymococca Chymococca P. galpini C.H.Wright P. laniflora C.H.Wright P. laniflora Gand. P. hamulata Gand.		(= P. rigida Wikstr, var. comosa Meisn.) P. burchellii Thoday (= P. tetragema fide Thoday) P. truncata (Meisn.) Bredenk. & A.E.van Wyk (= P. rigida Wikstr, var. truncata Meisn.) (= P. rigida Wikstr, var. tetragona Meisn.) (= P. glomerata sphalm. quoad L. conglomerata L. sensu Thunb.) (= Lachnaea glomerata Sphalm. quoad L. conglomerata L. sensu Thunb.) (= P. glomerata L. sensu Thunb.) (= P. glomerata Thunb.) (= P. glomerata Thunb.) (= P. glomerata Thunb.) (= P. glomerata Thunb.)
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eisn B truncata Wikstr. § truncata Meisn. Chymococca Chymococca cmpetroides Meisn. P. galnini C.H.Wright P. laniflora C.H.Wright P. laniflora Gand. P. hamulata Gand.		nicata (Meisn.) Bredenk. & A.E.van W. rigida Wikstr. var. truncata Meisn.) rigida Wikstr. var. tetragona Meisn.) glomerata sphalm. quoad L. conglomera sensu Thunb.) uchnaea glomerata sphalm. quoad conglomerata L. sensu Thunb.) ericoides sensu Thunb.) glomerata Thunb.) struncata y. truncata y. monticola Bredenk. & A.E. van Wyk
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E Bartl. Chymococca empetroides Meisn. P. galpini C.H.Wright P. laniflora C.H.Wright P. laniflora Gand. P. eriophora Gand.		rigida Wikstr. var. trancata Meisn.) rigida Wikstr. var. tetragona Meisn.) glomerata sphalm. quoad L. conglomera sensu Thunb.) tchnaea glomerata sphalm. quoad conglomerata L. sensu Thunb.) ericoides sensu Thunb.) glomerata Thunb., y glomerata Thunb., renticoides sensu Thunb.) truncata truncata monticola Bredenk, & A.E. van Wyk
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Meisn. P. galpini C.H.Wright P. laniflora C.H.Wright P. laniflora Gand. P. bamulata Gand.	7	(=P, cupressina J.C. Wendl. nom. nud.)
		coides L.
	roides Meisn.)	(= Chymococca empetroides Meisn.)
		P. galpimi C.H. Wright
	naea	Lachnaca laniflora (C.H.Wright) Bond
		(= P. laniflora C.H.Wright)
		P. rigida Wikstr.
	+	(= P. eriophora Gand.)
	probably P, paleacea	P. hamulata Gand.: nom, dub.
(= <i>P. ericoides</i> C.H.Wright nor	Wright non L. pro parte)	(= P. ericoides sensu Meisn.)
D adventibility Phonless		(= F. ngada Wikstr. var. tetragona Meisn, pro parte)
P reluidora Thoday		P valudosa Thoday
		P. drakensbergensis Hilliard & B.I. Burtt
	P. quadriff	P. quadrifaria Bredenk, & A.E. van Wyk
	P, nivicola	P. nivicola Bredenk, & A.E.van Wyk
	P. esterhuv	
		P. esterhuyseniae Bredenk, & A.E.van Wyk

Shrubs or small trees. Stems greyish brown; bark tough and stringy. Leaves decussate, imbricate on young branchlets, sessile, closely appressed to stem or spreading at an angle of 5-20°(-60°), cymbiform (boat-shaped), falcate or cigar-shaped; plane shape linear, oblong, lanceolate or narrowly trullate; base sessile or cuneate; apex truncate and hump-backed, obtuse, rounded, acuminate or acute to almost spine-tipped; margins sometimes ciliate; length × depth $(1.5-)2.5-4.0(-8.0) \times (0.8-)1.2-2.0(-3.0)$ mm (leaf shape usually cymbiform, depth of lamina is distance from adaxial groove to main vein situated abaxially); lamina inversely ericoid, adaxial surface concave, tomentose, abaxial surface convex, glabrous, seldom tomentose. Inflorescences comprising polytelic synflorescences [apex of main florescence (main axis) not terminating with a flower (indeterminate), co-florescences (lateral branches) of the same structurel; main florescences as well as co-florescences spicate; spikes reduced, resembling terminal subcapitulate inflorescences, each characterized by two terminal leaves with axillary blind-ending rudimentary flowers, enveloping minute growing point, proliferating growth (inflorescence apex growing out and returning to vegetative growth) less common (Figures 1, 3, 4), or spikes mostly extended, number of spikes reduced or multiflowered, main and co-florescences present, proliferating growth common (Figures 10, 12, 13, 15, 16, 20, 23). Bracts enveloping flowers and fruits, largest after anthesis of flowers, becoming more coriaceous and rounded at fruit set, decussate, imbricate, cymbiform or helmet-shaped; plane shape oblong, lanceolate, ovate and obovate to widely ovate and obovate, rhombic and narrowly obtrullate to obtrullate; base sessile, cuneate; main vein strongly developed, often keeled, extending to form a leaf-like point in many species; apex obtuse, rounded or acute; texture mostly coriaceous; lamina with adaxial surface (inside) concave, abaxial surface (outside) convex, outside usually glabrous, inside base or midrib tomentose or completely tomentose, coriaceous or chartaceous, rugose or smooth, ± succulent or thin, sometimes \pm 3-5-ribbed, reticulately veined or ribbed and reticulately veined on each side of main vein; wings absent or bullate, coriaceous, chartaceous or membranous: margins often ciliate to setose; size variable, bracts without leaf-like point, length \times depth (2.5-)3.5-4.5(-5.5) \times (0.9-) 1.0–1.5(–2.4) mm or bracts with leaf-like point, length \times depth $(4.0-)5.1-6.3(-7.3) \times (1.4-)1.5-2.0(-2.6)$ mm.

Flowers actinomorphic, hypogynous. Floral envelope constituting hypanthium and sepals, (4.0–)5.3–7.3(–8.4) mm long, membranous during pollination and yellowish in P. rigida, P. paleacea, P. nivicola and P. esterhuyseniae, slightly succulent and greenish in P. ericoides, mostly yellow-pink in all other species, dehydrated after shedding of pollen, becoming papyraceous or coriaceous, yellow-pink tones turning red. Pedicel very short or absent. Receptacle very short. Hypanthium a membranous to coriaceous cylindric tube (fused calyx and androecium, differentiating into sepals and diplostemonous androecium arising from hypanthium rim at separation of sepals, Figure 1); indumentum variable in density, trichomes nonglandular, uniseriate, often spiralled, whitish, density of indumentum at ovary ranging from glabrous to tomentose or strigose; neck (narrowed tube between apex of ovary and sepals) (0.3-)0.6-2.6(-3.0)mm long, density of indumentum ranging from glabrous to tomentose on outside, inside often hairy, abscission tissue not macroscopically discernable, articulation plane

absent, after fruiting fragmentation of neck base caused by dehydration and torsification of tissue, sepals and androecium being shed in most species. Sepals 4, petaloid, imbricate in bud, flexed in flower, often setose with up to 5 long trichomes on outer surface and glabrous to tomentose on inner surface; outer sepals cymbiform or concave; inner sepals oblong, elliptic or obovate. Corolla absent. Disc absent. Androecium dimorphic diplostemonous, arising from hypanthium at separation point of sepals; filaments of antipetalous whorl (0.4–)0.7–1.2(–1.5) mm long, those of antisepalous whorl (1.2–)1.4–2.2(–2.4) mm long; anthers $(0.5-)0.7-0.9(-1.1)\times(0.2-)0.3-0.4(-0.7)$ mm, sub-basifixed, 2-thecous and 4-locular, exserted, extrorse. Ovary superior, $(1.6-)2.0-2.5(-2.7)\times(0.5-)0.6-1.4$ (-1.7) mm, bicarpellate during embryonic stage (Bunniger 1972), pseudomonomerous (Heinig 1951) at maturity, placentation parietal, uniloculate, with 1 pendulous ovule laterally attached near top of ovary; style separating laterally from top of ovary, maintaining lateral position in hypanthium neck, reaching beyond hypanthium rim; stigma ± globose, mop-like or penicillate (wind pollination). Fruit enveloped by persistent, loosely arranged hypanthium fragmented at neck base or, in some species, fragmenting over widest circumference of fruit, the fragmented hypanthium, sepals and androecium being shed; in P. ericoides and P. rigida a fleshy, 1-seeded berry, 5.3 × 4 mm; in all other species an achene, pericarp membranous and dry, 2.5 × 1.2 mm. Seed broadly fusiform with outgrowths at both micropylar and funicular ends, $2.2(-2.9) \times 1.2(-1.6)$ mm; tegmen black and shiny, often with white spots; endosperm formation nuclear, but later becoming cellular throughout.

Diagnostic characters: plants of Passerina are shrubs or small trees, distinguished by the inversely ericoid leaves, that are inverse-dorsiventral in c/s. The inflorescences are few- to multiflowered, simple or compound spikes, often reduced, artificially resembling terminal subcapitulate inflorescences. Each flower is enveloped by a conspicuous bract, becoming more coriaceous and rounded at fruit set. The flowers are adapted to wind pollination. During pollination the flower colour is yellow-pink, the four petaloid sepals are flexed and the anthers are exserted and extrorse (unique for Thymelaeaceae in southern Africa). All anthers open explosively and the pollen is shed at once. The stigma is moplike. The fruit is enveloped by a persistent, loosely arranged hypanthium. Because of the absence of an articulation plane, the hypanthium fragments at neck base or, in some species, over the widest circumference of the fruit, the fragmented hypanthium, sepals and androecium being shed. P. ericoides and P. rigida are characterized by a fleshy, 1-seeded berry and all other species by an achene.

Etymology: Passerina refers to the Latin passer (= a sparrow) as the seeds resemble a sparrow's beak.

Common names: the vernacular name 'sparrow-wort' was suggested by Miller (1768) for all Passerina species and Wendland (1798) used the name fadenförmige Vogelkopf. According to Smith (1966) gonna is a collective name once used by the Khoekhoe for various members of Thymelaeaceae, e.g. several species of Passerina and Struthiola.

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Uses: many Passerina species grow on sand dunes and in sandy areas, with parts of the woody stem subterraneous, forming runners and developing an extended root system. Most of these plants are pioneers and resprouters, increasing their chances of survival in disturbed areas. These plants are excellent sand binders and are suitable for binding problematic sandy areas, especially after the clearing of invader species. Sim (1919) recommended Passerina in his list of trees and shrubs for coastal areas exposed to sea winds. Certain Passerina species such as P. falcifolia are small trees and can be used as ornamental garden plants. P. filiformis has been cultivated in Britain and Europe since the time of Linnaeus, P. obtusifolia is used in the wild flower industry in the Robertson area. The bark is exceedingly tough and is used for tying down thatch. According to Watt & Breyer-Brandwijk (1962) it is also plaited into whip thongs and used as twine. Members of the genus are not browsed by stock as the plants are apparently unpalatable (Story 1952). Ash from Passerina obtusifolia was traditionally used by the people of Genadendal in Western Cape in the home industry of soap-making. Although certain species have been recorded in cancer research, these plants are not currently known for their medicinal value.

Flowering and fruiting: most Passerina species flower profusely in spring, from September to October. During this season the Cape Floristic Region is quite windy and large amounts of pollen are produced, as Passerina is wind-pollinated. Pollen is often wafted away in clouds, causing a kind of hay-fever in sensitive persons (Marloth 1925). Fruiting time is mostly from December to January. The fleshy fruits of P. ericoides and P. rigida are dispersed by birds or rodents inhabiting the distribution ranges of these species along the South African coast. Fruits of P. truncata subsp. truncata, growing in the Karoo, passively fall to the ground, where they are probably dispersed by ants or rodents. The fruits of P. montana, occurring along the Great Escarpment,

are probably dispersed by birds as they are arranged at the tips of branchlets, exposed, red, and beak-like, possibly resembling the beaks of nestlings.

Distribution and ecology: in Passerina the highest number of species per grid (nine) occurs in each of the grids 3321 (Ladismith), 3322 (Oudtshoorn) and 3419 (Caledon). The highest diversity of species (six) occurs in the False Bay area, from Seekoeivlei, including the Cape Flats, to De Mond at the Palmiet River (3418B). After an extensive study of herbarium material in cooperation with field work, Western Cape is regarded as the centre of diversity for Passerina, from where certain species extend west, north and east.

Thoday (1925) published an account of the geographical distribution and ecology of *Passerina*, based on 15 species. Of the 20 species currently studied, 10 are endemic and 4 species are near-endemic to the Cape Floristic Region. P. obtusifolia is widespread in the Northern, Eastern and Western Cape, whereas P. corymbosa occurs in Western and Eastern Cape, with outliers in KwaZulu-Natal. P. rigida is distributed from Western Cape, along the coast to northern KwaZulu-Natal; all these species are endemic to the southern African provinces in which they occur. P. drakensbergensis is endemic to the Bergville District in KwaZulu-Natal. P. montivaga is found from Mossel Bay and Oudtshoorn to Eastern Cape and along the escarpment northwards to Zimbabwe and P. montana is distributed from the eastern mountains and Great Escarpment of southern Africa to Zimbabwe and Mozambique. P. montivaga and P. montana are near-endemic to the Great Escarpment.

Conservation status: with the exception of Passerina esterhuyseniae (from the northern Cederberg Mountains) known from herbarium material only, all other species and subspecies of Passerina were studied in the wild. Assessments were done using the guidelines of the IUCN Species Survival Commission (2000) and Victor (2002).

Key to species

1a Inflorescences comprising terminal subcapitulate spikes; proliferating growth (inflorescence apex growing out and returning to vegetative growth) uncommon (Figures 1, 3, 4): 2a Floral envelope yellow and membranous, up to 4 mm long, neck (portion of hypanthium between ovary and sepals) very short, ± 0.3 mm long; bracts widely ovate, wings membranous and obscurely veined1. P. paleacea 2b Floral envelope yellow-pink and papyraceous, 4.9-6.4 mm long, neck 0.7-1.4 mm long; bracts variously shaped, wings present or absent: 3a Leaves narrowly oblong to oblong; base sessile, dilated; apex truncate, or truncate to rounded, keeled, often 2. P. truncata appearing humped on the back (abaxially) . . . 3b Leaves linear-lanceolate; base diamond-shaped to rounded; apex rounded to acute: 4a Abaxial surface of young leaves tomentose; bracts ovate to widely ovate; lamina comose on inside, sparsely hairy to tomentose on outside, ± 3-ribbed on each side of main vein, coriaceous and rugose; length × depth $(4.5-)4.9 \times 1.5(-1.8)$ mm (leaf shape usually cymbiform, depth of lamina is distance from adaxial groove to main vein situated abaxially). 4b Abaxial surface of young leaves glabrous; bracts ovate to obovate; lamina villous on inside, glabrous on outside, obscurely ribbed on each side of main vein, thinly coriaceous; length × depth (3.2-)4.0 × 0.9(-1.6) mm 1b Inflorescences comprising extended spikes, number of spikes often reduced, or many compound, multiflowered spikes present; proliferating growth common (Figures 10, 12, 13, 15, 16, 18, 20, 21, 23): 5a Inflorescences reduced, often to solitary spikes; dwarf shrubs up to 300 mm high; bracts rhombic, dark green when fresh, dark brown in dried specimens, coriaceous, membranous wings absent 5b Inflorescences with multiflowered main and co-florescences; low shrubs, shrubs or small trees; bracts variously coloured, textured and winged: 6a Fruit fleshy; floral envelope greenish or yellow, coriaceous or membranous; leaves greyish green 7a Fruit a red berry; floral envelope greenish and coriaceous, hypanthium strigose; leaves oblong, apex obtuse

7b Fruit a bright yellow berry; floral envelope yellow and membranous, glabrous at ovary, tomentose at neck; leaves narrowly lanceolate to ovate, apex acute with main vein visible as a blunt keel; bracts widely 6b Fruit dry (an achene); floral envelope yellow, yellow-pink or red, membranous or papyraceous; leaves variously coloured: 8a Floral envelope membranous, vellow or vellow-pink: 9a Floral envelope yellow or yellow-pink; bracts chartaceous, widely obovate, wings membranous, often 9b Floral envelope yellow; bracts thinly chartaceous, smooth and helmet-shaped with membranous rims. 9. P. esterhuyseniae 8b Floral envelope papyraceous, yellow-pink or red: 10a Young leaves and bracts abaxially sparsely hairy, becoming tomentose towards apex, older leaves rugose 10b Young leaves and bracts abaxially glabrous: 11a Bracts shorter than 4.5 mm: 12a Bracts rhombic in outline, softly coriaceous, with membranous wings, margins brownish ciliate. 12b Bracts oblate in outline, chartaceous, with bullate membranous wings, margins glabrous 11b Bracts longer than 4.5 mm: 13a Floral envelope ± 5.9 mm long; outer and inner sepals concave and lanceolate; bracts larger than 13b Floral envelope 6.0–8.4 mm long; outer and inner sepals variously shaped; bracts not as above, variously shaped and coloured: 14a Midrib of bract shortly extended into an acute apex, lamina rhombic to obtrullate, distinctly angled, 4- or 5-ribbed; leaves with a distinct midrib, laterally compressed, greyish green, drying greyish 14b Midrib of bract extending beyond lamina into a leaf-like point, lamina variously shaped and ribbed; leaves with a distinct midrib, or midrib less obvious, abaxially convex or laterally compressed, variously coloured: 15a Bracts with leaf-like point, obtuse at apex, lamina closely 2-ribbed at margin; hypanthium fragments 15. P. obtusifolia 15b Bracts with leaf-like point variously shaped, but not obtuse, lamina (faintly ribbed in P. drakensbergensis) extending into a membranous margin; hypanthium fragments at neck base: 16a Adaxial (inner) surface of bracts basally to centrally setose or tomentose over entire length of midrib; wings glabrous: 17a Bracts with midrib and leaf-like point stout and strongly developed, apex acute 16. P. paludosa 17b Bracts with midrib forming a straight or filiform, leaf-like point, or midrib shortly extended: 18a Bracts with leaf-like point straight or slightly incurved; wings of bracts ovate, margins hairy in distal half, or obtrullate, narrowing abruptly into midrib 17. P. montivaga 18b Bracts with leaf-like point shortly extended or extended into a filiform, slightly falcate point; wings of bracts ovate-acuminate, gradually narrowing to a point or widely obovate, 16b Adaxial (inner) surface of bracts completely villous: 19a Bracts with midrib extended, leaf-like point falcate; wings of bracts ± 4-ribbed; hypanthium neck ± 3 mm long, tomentose, often arcuate; spikes lax, often arcuate, mottled grey-green, with up to 16 fertile, enlarged bracts 19b Bracts with midrib shortly extended into a short point, apex acute; wings of bracts ± 5-ribbed; hypanthium neck ± 2 mm long, glabrous to sparsely pubescent; spikes robust, rigid and

1. **Passerina paleacea** *Wikstr.* in Kunglinga Svenska Vetenskapsakademiens Handlingar 39: 323 (1818); Meisn.: 400 (1840); Meisn.: 562 (1857); C.H.Wright: 12 (1915); Thoday: 164 (1924a); Thoday: 388 (1924b). Type: Caput bonae Spei, Herb. Wikströmii, *Sparrman s.n.* (S!, lecto., here designated; UPS!).

Lachnaea paleacea Herb. Banks, ined., fide Wikstr.: 324 (1818); Meisn.: 562 (1857); C.H.Wright: 12 (1915); Thoday: 164 (1924a), nom. inval. in synonymy.

Passerina glomerata sensu Thunb.: 374 (1825a) pro parte quoad specim. Herb. Thunberg 9596D, 9579.

L. conglomerata L. sensu Thunb.: 374 (1825a) pro parte quoad specim. Herb. Thunberg 9596D.

P. ericoides sensu Thunb.: 374 (1825a) pro parte quoad specim. Herb. Thunberg 9596D, 9579, non L.; P. ericoides sensu Meisn.: 401 (1840) pro parte, non L.; Meisn.: 562 (1857) pro parte quoad specim. Drège s.n. (G!, K!, P!, S!).

Shrubs or shrublets 0.1–1.5 m high. *Stems* branching from base up to growing points, branchlets from previous growth persistent, arcuate, indurate; younger branchlets

ascending, densely white-tomentose, villous closer to growing points; shredded bark of older branchlets greyish brown, remains of tomentum forming lengthwise strips; leaf scars conspicuous; older stems fissured lengthwise exposing greyish white sclerenchyma fibres; internodes shorter than leaves. Leaves imbricate on young branchlets, closely appressed to stem, diverging at an angle of \pm 0–5°, cymbiform, often expanding, becoming thickly chartaceous and bract-like towards inflorescences; lamina inversely ericoid, adaxial surface concave, tomentose, abaxial surface laterally compressed and glabrous, plane shape linear to linear lanceolate, length \times depth 1.5–2.5(–4.0) \times 0.6–0.8(–1.2) mm; base sessile, dilated; apex acute, median vein forming a distinct keel incurved at apex; margins involute. Inflorescences subcapitulate, ± ellipsoid. Bracts decussate, imbricate, sessile, appressed, widely ovate in outline, length \times depth (2.5–)2.7 \times 1.2(–1.9) mm; lamina adaxially (inside) concave and villous, abaxially (outside) convex and glabrous, thickly chartaceous, smooth on each side of main vein, concolorous, greyish green, senescing to yellow-

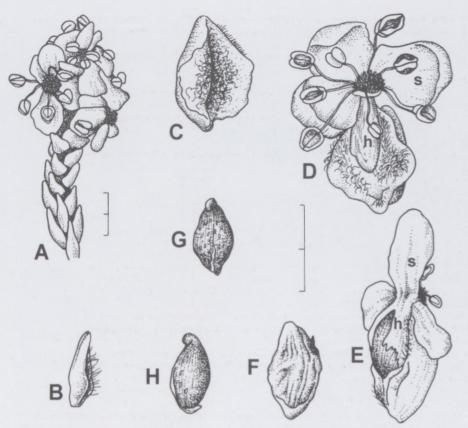


FIGURE 1.—Passerina paleacea,
Bredenkamp 960. A, spike
reduced, resembling terminal
subcapitulate inflorescence;
B, leaf; C, bract; D, flower
clasped by bract in ventral
view; E, hypanthium fragmenting at circumference of
ovary. F-H, achene: enveloped by membranous pericarp;
G, lateral view; H, ventral
view. h, hypanthium; s, sepals. Scale bars: 2 mm. Artist:
A. Stadler.

ish brown; base cuneate; main vein extending into obtuse apex; wings membranous, borders glabrous, obscurely veined. Floral envelope membranous and yellow during pollination, dehydrated after shedding of pollen, turning red to brownish, \pm 4.2 mm long. Hypanthium glabrous, neck \pm 0.3 mm long, abscission tissue and articulation plane absent. Sepals concave, elliptic or subrotund and glabrous. Androecium with filaments of antipetalous whorl \pm 0.7 mm and antisepalous whorl \pm 1.4 mm long; anthers 0.6 \times 0.5 mm. Ovary 2.4 \times 1.4 mm. Fruit an achene with pericarp membranous and dry, \pm 2.3 \times 1.2 mm, enveloped by persistent, loosely arranged hypanthium fragmenting over widest circumference of fruit, the fragmented hypanthium, sepals and androecium being shed. Figure 1.

Nomenclatural notes: in the latter half of the eighteenth century Lachnaea conglomerata L. (1753), Passerina ericoides L. (1753) and P. glomerata Thunb. (1794) were constantly confused by botanists, causing Wikström (1818: 322) to place P. glomerata and L. conglomerata in the synonymy of P. conglomerata Thunb. In the same publication Wikström delimited and described P. paleacea. However, P. paleacea is not mentioned in Thunberg's revision of 1825, in which he described P. glomerata occurring in 'Hautbay', the currently known locality of both P. paleacea and P. ericoides. This confusion is reflected on many herbarium specimens, e.g. the specimen Herb. Thunberg 9579, bearing the inscriptions P. ericoides, P. glomerata (struck out) and the word 'paleacea' written in pencil. Although Thoday (1924b) chose the specimen Herb Thunberg 9597 as the type of P. paleacea, this specimen was not chosen as lectotype in the present study, as the Sparrman specimen cited by Wikström (1818: 324) was located at S. The specimen *LINN 504.3*, positively identified as P. paleacea, bears the inscription 'sp 161', possibly referring to Sparrman. However, there

will always be doubt whether it is a duplicate of the Sparrman specimen cited by Wikström.

Diagnostic characters and relationships: Passerina paleacea may easily be confused with P. rigida, as both occur on sand dunes along the coast. The branches of P. rigida are nodding and abundantly covered by pendulous branchlets, spikes are extended and the fruits are fleshy. yellow berries. Plants of P. paleacea are less robust, reaching a maximum height of 1.5 m, and are characterized by an abundance of subcapitulate inflorescences and dry fruit. The subcapitulate inflorescences at times led to the confusion of P. paleacea with P. truncata (= P. glomerata), but, these two species are morphologically as well as geographically distinct. P. paleacea has a maritime habit and P. truncata is distributed from Vanrhynsdorp, along the Cederberg Mountains, to Malmesbury, Ceres, Tulbagh and Matjiesfontein up to Seven Weeks Poort. The earlier confusion between P. paleacea and P. ericoides was probably due to their sympatric occurrence, but these two species are morphologically quite different.

Etymology: the Latin specific epithet *paleacea* (= chaffy) probably refers to the chaff-like subcapitulate inflorescences.

Distribution and ecology: Passerina paleacea occurs in both the Southwestern and the Agulhas Plain Centres of the Cape Floristic Region (CFR) (Goldblatt & Manning 2000) and is a typical fynbos element. It grows on coastal dunes and in maritime habitats from Langebaan, round the Cape Peninsula to the Cape Flats, Kogel Bay, Hermanus, Gansbaai, De Hoop, the Potberg coast, Bredasdorp, Arniston, Vermaaklikheid and Puntjie up to Stilbaai (Figure 2). The vegetation types dune fynbos and dune thicket form a mosaic along many parts

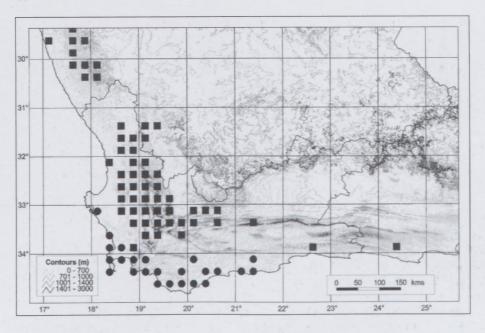


FIGURE 2.—Known distribution of Passerina paleacea, •; P. truncata subsp. truncata, •.

of the southern Cape coast (Lubke 1998a, 1998b; Lubke & Van Wijk 1998). This same distribution pattern is displayed by *P. paleacea* as it is found in the dune scrub, amongst typical fynbos species, but not in the dune thicket amongst larger shrubs or small trees with mesophytic leaves such as species of *Chrysanthemoides*, *Mimusops*, *Morella*, *Rhus* and *Sideroxylon*.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

2. **Passerina truncata** (*Meisn.*) *Bredenk.* & *A.E.van Wyk* in Bothalia 32: 66 (2002a). Type: Western Cape, near Tulbagh Waterfall, April 1865, *Zeyher 43* (K!, lecto.; MEL!, MO!, NBG!, S!, W!).

The complete description of the species and subspecies, the synonymy, relationships, etymology, distribution, habitat and key to subspecies is dealt with in *Bothalia* 32: 66–71 (2002).

2a. subsp. truncata (Figures 2, 3)

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

2b. subsp. monticola Bredenk. & A.E.van Wyk

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

3. Passerina quadrifaria Bredenk. & A.E.van Wyk in South African Journal of Botany 68: 304 (2002b). Type: Eastern Cape, 3324 (Steytlerville): Uitenhage District, Great Winterhoek Mountains, Cockscomb, (–BD), rocky ridge, 30 Nov. 1958, Esterhuysen 28006 (PRE, holo.!; BOL!, K!).

Passerina sp. nov. 3 Bredenk. & A.E.van Wyk: 70 (2000); Bredenk. & A.E.van Wyk: 56 (2001a); Bredenk. & A.E.van Wyk: 217 (2001b).

The complete description of the species, relationships, etymology, distribution and habitat appear in the *South African Journal of Botany* 68: 304–307 (2002).

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

4. **Passerina montana** *Thoday* in Bulletin of Miscellaneous Information, Kew 4: 152 (1924a); Norl. & Weim.: 630 (1958); Bond & Goldblatt: 432 (1984); Hilliard & B.L.Burtt: 182 (1987). Type: Mooi River, *Wood*

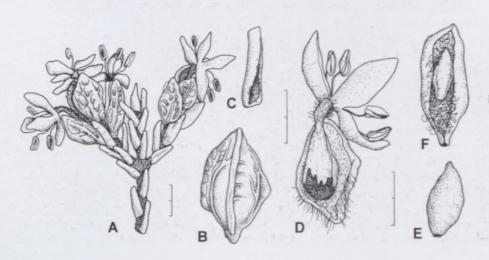


FIGURE 3.—Passerina truncata subsp. truncata, Bredenkamp 985. A, spike reduced, resembling terminal subcapitulate inflorescence; B, bract; C, leaf; D, flower clasped by bract, fragmenting at circumference of ovary. E, F, achene: E, clasped by bract; F, enveloped by membranous pericarp. Scale bars: 2 mm. Artist: A. Stadler.

4036 [K!, lecto., designated by Thoday 10: 387 (1924b); BOL!, GRA!, PRE!].

P. ericoides sensu Meisn.: 401 (1840) pro parte, non L.; Meisn.: 562 (1857) pro parte; C.H.Wright: 12 (1915), pro parte.

P. rigida Wikstr. var. tetragona Meisn.: 563 (1857) pro parte quoad specim., circa Stormberg Drège s.n.

Small trees or shrubs, (0.2-)1.0-2.0(-2.5) m high. Stems branching from base up to growing points, young stems profusely branched, young branchlets ascending; branchlets terminally leafless and woolly, with conspicuous terminal scars after dispersal of fruit; bark grevish brown. younger branchlets densely white-tomentose, villous closer to growing points; bark on older branchlets shredding, greyish brown, remains of tomentum forming lengthwise strips; leaf scars conspicuous; older stems fissured lengthwise exposing greyish white sclerenchyma fibres; internodes mostly shorter than leaves. Leaves imbricate on young branchlets, closely appressed to stem, diverging at an angle of ± 0-5°, cymbiform, lamina inversely ericoid, adaxial surface concave, tomentose, abaxial surface laterally compressed and glabrous, plane shape linear to lanceolate, length \times depth 1.5–2.5(–4.0) \times 0.6–0.8 mm; base sessile, dilated; apex acute, median vein prominent in upper third of leaf, incurved at apex; margins involute. Inflorescences subcapitulate, ± ellipsoid. Bracts decussate, imbricate, sessile, appressed, ovate to obovate in outline, length \times depth (3.2-)4.0 \times 0.9(-1.6 mm); lamina adaxially (inside) concave and villous, abaxially (outside) convex and glabrous, thinly coriaceous, obscurely ribbed, yellowish green, margins of fruiting bracts turning red; base sessile; main vein extending into subacute apex; wings membranous, brownish. Floral envelope papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brownish, ± 5.8 mm long. Hypanthium glabrous at ovary, neck tomentose, ± 0.8 mm long, fragmentation at neck base. Sepals: outer sepals cymbiform, adaxially scantilly tomentose, abaxially glabrous; inner sepals obovate, adaxially tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl \pm 0.6 mm and those of antisepalous whorl \pm 1.5 mm long; anthers \pm 0.8 \times 0.4 mm. Ovary \pm 2.1 \times 0.6 mm. Fruit an achene enveloped in beak-like, reddish, papyraceous hypanthium, fragmented at neck base; pericarp membranous and dry, $\pm 2.3 \times 1.2$ mm. Figure 4.

Nomenclatural notes: according to the concept of Wright (1915), P. ericoides is not only distributed along the southern coast of Western Cape (present interpretation), but also along the coast to Eastern Cape and further inland up to Mpumalanga. However, most of the inland specimens cited by him have been classified as P. montana by Thoday (1924a). The interpretation of P. corymbosa by Wright (1915) posed the same problem, as Wood 4036 (the lectotype of P. montana) was also placed in this taxon.

Meisner (1857) described *P. rigida* var. *tetragona* citing two Drège specimens, one from Ezelsbank and the other from Stormberg. The Ezelsbank specimen (*Drège 2971*, P, K) is *P. truncata*, but the Stormberg specimen could not be located. According to Gunn & Codd (1981), Drège crossed the Stormberg (3126BC, Queenstown) on 17 December 1832. The first author suspected that the Drège specimen would be *P. montana*, as it is common in this area. This suspicion is supported by *Sim 68* (from the Pirie Mountains in the King William's Town District), a syntype of *P. montana*, bearing the inscription '*P. rigida* Wiks-*tetragona*' and the Drège specimen from Stormberg is consequently regarded as *P. montana*.

Diagnostic characters and relationships: Hilliard & Burtt (1988) noted two rather distinct forms of P. montana in KwaZulu-Natal. The first form is characterized by plants on rock platforms that are low, rounded bushes, 0.3-1 m high, with the tips of the branches erect, whereas those of the second form inhabit valleys and are riverside bushes of up to 2 m high, with open branches and pendulous branchlets. The present study, taking the whole distribution range of P. montana into consideration, recognizes two forms. One, centred in the God's Window area of Mpumalanga, are rounded shrubs 0.5-2 m high, with many branchlets covered with smaller, decussate, imbricate leaves, bluish green in colour. The second form dominates in the Free State, Lesotho, KwaZulu-Natal and Eastern Cape. These plants are more robust, with open branches and larger, yellowish green leaves and inflorescences, which are tinged pink. However, the two forms are not geographically distinct and intermediates are common. Both forms unequivocally show the specific characters and therefore we do not propose to give them formal taxonomic recognition. Studies

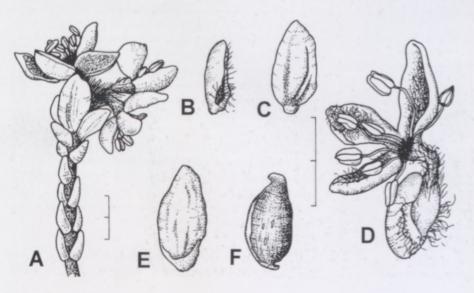


FIGURE 4.—Passerina montana, Bredenkamp 893. A, spike reduced, resembling terminal subcapitulate inflorescence; B, leaf; C, bract; D, flower clasped by bract. E, F, achene: E, enveloped by membranous pericarp; F, lateral view. Scale bars: 2 mm. Artist: A. Stadler.

of the leaf epidermis, anatomy and floral morphology (Bredenkamp & Van Wyk 2000, 2001a, 2001b) supplied no further evidence on which the two forms could be delineated.

Etymology: the specific epithet is derived from the Latin montanus (= pertaining to or growing on mountains). This is a very appropriate epithet as P. montana is distributed along the Great Escarpment from Eastern Cape to Zimbabwe.

Common names: Cooper 2302 (K), from Lesotho, reported the vernacular name Likhabei and Staples 17 (PRE), from the Maluti Mountains in Lesotho, recorded the name Lekaphu. Story (1952) mentions the name pakaan. Von Breitenbach et al. (2001) used the names berg-gonna and mountain gonna.

Uses: information on the specimen Watt & Breyer-Brandwijk 1851, collected at Thabaneng, states that the plants are used medicinally. However, Watt & Breyer-Brandwijk (1962) supplied no further details.

Distribution and ecology: Passerina montana is a near-endemic to the Great Escarpment of southern Africa, with distant satellite populations in high mountain areas of Angola, Namibia and Limpopo [Northern Province], South Africa. It is distributed from Nyanga in Zimbabwe, along the escarpment to Manica and Sofala in Mozambique, Limpopo, Mpumalanga, Swaziland, KwaZulu-Natal, Free State, Lesotho and the Eastern Cape (Figure 5). Outliers in Angola have been found on the escarpment of the Huilla Plateau near Lubango and the Cheila Mountains. Several specimens of this species have been collected at Moltkeblick on the Auas Mountains in Namibia. In Limpopo, P. montana is found in the Soutpansberg area and on the Blouberg, as well as on the summit of Krantzberg in the Waterberg Mountains. A single specimen (Goossens 375) was collected in the Pretoria District, but the species is currently probably extinct in this area, due to human impact.

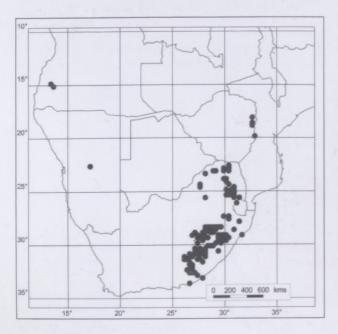


FIGURE 5.—Known distribution of Passerina montana.

This species grows at altitudes of (900–)1 200–3 000 m. At Nyanga, *P. montana* is associated with *Erica mannii* and *E. hexandra*, bordering on *Brachystegia* woodland and montane forest. In Mozambique and South Africa it has been found with *Widdringtonia nodiflora* and *Erica* species, bordering on montane forest. It is common amongst rocks on hills, mountain slopes, mountain tops, cliff ledges and rocky ridges. It also frequents stream courses and banks as well as riverbeds and banks, where the growth form has been reported as a shrub amongst rocks, a drooping bush over running water, a limply spreading bush in sand or dense bushes. These plants also grow in river valley forests and along plantations.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

5. **Passerina burchellii** *Thoday* in Bulletin of Miscellaneous Information, Kew 4: 155 (1924a); Bond & Goldblatt: 432 (1984); Goldblatt & Manning: 683 (2000). Type: Cape, Caledon Div., mountain tops of Baviaanskloof near Genadendal, *Burchell* 7761 [K, lecto.!, designated by Thoday: 387 (1924b); M!, P!].

P. tetragona Burch. MS. in Herb. Kew, non Steud. fide Thoday: 156 (1924a).

Low, erect, many-stemmed shrublets, branching mostly on new growth, ± 0.3 m high, from a common rootstock. Stems greyish brown, cork fissured, grey-brown, scabrous, surrounding prominent leaf scars; indumentum at growing point densely white-tomentose, flaking off with cork on older branchlets, which become glabrous. Leaves imbricate, overlapping ± 50%, diverging at an angle of 30°, plane shape rhombic, length \times depth 2.8(-3.5) \times 1.5 mm, adaxial surface concave, villous, abaxial surface convex, glabrous; base sessile, cuneate; apex subacute, bearded; margins brownish setose. Inflorescences with spikes extended, number of spikes often reduced, spikes sometimes solitary, 6-12-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. *Bracts* appressed, rhombic, length \times depth (3.2–)3.5 \times 1.5 mm; lamina adaxially concave (inside), abaxially convex (outside), villous inside, glabrous outside, coriaceous and smooth, extending into a smooth wing, dark green when fresh, dark brown in dried specimens; base cuneate; main vein extending into acute, bearded apex; margins brownish setose, involute. Floral envelope ± 4.7 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium glabrous at ovary, neck tomentose, ± 0.8 mm long. Sepals: outer sepals cymbiform, midrib adaxially and apex abaxially setose; inner sepals obovate, adaxially tomentose, apex abaxially setose. Androecium with filaments of antipetalous whorl ± 0.7 mm and those of antisepalous whorl ± 1.5 mm long; anthers 0.5×0.3 mm, subbasifixed, 2-thecous and 4-locular. Ovary $\pm 1.6 \times 0.6$ mm. Fruit an achene, pericarp membranous and dry, $\pm 2.5 \times 1.2$ mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in sepals and androecium being shed. Figure 6.

Nomenclatural notes: although Thoday (1924a) cited P. rigida var. comosa Meisn. partly (ex MS. in Herb.

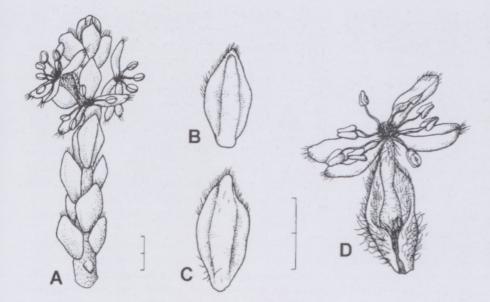


FIGURE 6.— Passerina burchellii, H. Bolus 687. A, flowering inflorescence; B, leaf; C, bract; D, flower clasped by bract. Scale bars: 2 mm. Artist: A. Stadler.

Kew) in synonymy under *P. burchellii*, the specimens cited by Meisner (1857) in the description of the var. *comosa* all belong to *P. pendula* (Eckl. & Zeyh.) Thoday.

Diagnostic characters and relationships: bearded sepals, leaves and bracts distinguish this species from *P. pendula*.

Eponymy: this plant was named in honour of the explorer and botanist W.J. Burchell, who collected in Caledon and as far north as Tulbagh between 1810 and 1811. During this trip *Burchell 7761*, the lectotype of *P. burchellii*, was collected on the summit of the mountains of Baviaanskloof near Genadendal.

Distribution and ecology: Passerina burchellii is endemic to the Southwestern and Langeberg Centres within the CFR. It is common on mountain summits of the Villiersdorp and Genadendal Districts (Figure 7), with outliers on southeastern rocky slopes of Towerkop in the Swartberg Mountains at Ladismith. This species occurs at altitudes of 1 333–2 167 m, often covered in mist. It is found in small groups on sandy loam, between boulders and rocks on upper south- or southeast-facing slopes.

Conservation status: Vulnerable (VU D2) (Victor 2002) because of small population size.

6. Passerina ericoides *L.*, Systema naturae 12,2: 733 (1767); Burm.f.: 12 (1768); L.: 236 (1771); L.: 374 (1784); Willd.: 430 (1799); Poir.: 41 (1804); Wikstr.: 325 (1818); Meisn.: 401 (1840) pro parte minore; Steud.: 274 (1841); C.A.Mey.: 49 (1843); Meisn.: 562 (1857) pro parte minore; C.H.Wright: 12 (1915) pro parte; Thoday: 166 (1924a); Bond & Goldblatt: 432 (1984); Hilliard & B.L.Burtt: 182 (1987); Goldblatt & Manning: 683 (2000). Type: *Passerina ericoides, Linnean Herbarium 504.5* [LINN, lecto.!, designated by Thoday: 148 (1924a)].

Chymococca empetroides Meisn.: 565 (1857); Harv.: 325 (1868); Bolus & Wolley-Dod: 315 (1904); C.H.Wright: 15 (1915); Thoday: 166 (1924a). P. filiformis L. var. crassifolia Eckl. & Zeyh. fide Meisn.: 565 (1857). Type: Eckl. & Zeyh. herb. no. 39 (G!, lecto., here designated; BOL!, MO!, P!, W!).

P. glomerata sensu Meisn.: 562 (1857), non Thunb.: 75 (1794). Lachnaea conglomerata L. sensu Meisn.: 562 (1857).

Low, rounded, many-stemmed shrublets, branching profusely on new growth, 0.3–1.2 m high, older branch-

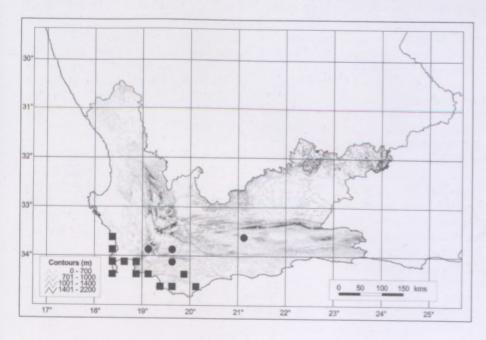


FIGURE 7.—Known distribution of Passerina burchellii, •; Passerina ericoides, •.

lets indurate, young branchlets lax, often arcuate. Stems light greyish brown, cork fissured lengthwise, greybrown, whitish scabrous, surrounding prominent leaf scars; indumentum at growing point densely white strigose, flaking off with cork on older branchlets, which become glabrous. Leaves slightly succulent, imbricate, overlapping ± 50%, diverging at an angle of 45°, plane shape oblong, length \times depth 2.5–2.8 \times 0.6–0.7 mm, adaxial surface concave, villous, abaxial surface convex, glabrous, greyish green, smooth; base sessile, dilated; apex obtuse to subacute; margins glabrous, basally sparsely setose. Inflorescences with spikes usually extended, 6-12-flowered, arrangement subterminal, axis white strigose, proliferating growth common. Bracts spreading at an angle of 60° (flowering) or 90° (fruiting), oblong to lanceolate, length \times depth \pm 3.6 \times 1.5 mm; lamina adaxially concave (inside), abaxially convex (outside), villous inside, glabrous outside, smooth, wings absent, greyish green, slightly succulent; base dilated; apex obtuse to subacute; margins glabrous, basally sparsely setose, involute. Floral envelope ± 5 mm long, coriaceous and greenish during pollination, dehydrated after shedding of pollen, turning red. Hypanthium ovateoblong at ovary, strigose, neck strigose, ± 0.6 mm long. Sepals globose when young, concave, widely obovate, outer and inner sepals adaxially puberulent, abaxially glabrous. Androecium with filaments of antipetalous whorl \pm 0.5 mm and those of antisepalous whorl \pm 1.3 mm long; anthers large, $\pm 0.9 \times 0.7$ mm, subbasifixed, 2thecous and 4-locular. Ovary \pm 2.1 \times 1.7 mm. Fruit a fleshy red berry, $\pm 5.3 \times 4$ mm, enveloped by persistent, loosely arranged hypanthium, fragmenting over widest circumference of fruit, the fragmented hypanthium, sepals and androecium being shed. Seed $\pm 2.9 \times 1.6$ mm.

Nomenclatural notes: in the Catalogue of the Linnaean Herbarium, Savage (1945) made the following inscription 'Tulb. list c. 1769. n.1. det. L.—Blaeria ericoides'. This refers to consignments of bulbs, seeds and herbarium specimens that Rijk Tulbagh sent to Van Royen, the Burmans at Amsterdam and Linnaeus at Uppsala (Gunn & Codd 1981). Jackson (1917, 1918) published a list of 203 of the specimens sent to Linnaeus around 1769 and identified by him. The first inscription on the list is the provisional name Blaeria ericoides, which Savage (1945) believed to be the P. ericoides specimen at LINN, but there is no numbering or any other indication on the specimen to link it with Tulbagh's list (Jackson 1917–1918). As Linnaeus had already described P. ericoides in 1767, the specimen at LINN is probably not part of the Tulbagh collection. Thoday (1924a) clearly regarded the specimen at LINN, named by Linnaeus, as the type of *P. ericoides*. As no other original elements exist, P. ericoides LINN 504.5 is regarded as a lectotype designated by Thoday (1924a).

Thunberg (1825a) accepted Wikström's concept of *P. glomerata*, occurring at Hout Bay in the Cape, and cited *P. ericoides* in synonymy, causing confusion about the identity of the latter taxon. Meisner (1840) reinstated *P. ericoides*, but the concept of this taxon became even more doubtful in the light of the cited distribution. In 1857 Meisner retained his concept of *P. ericoides*, occurring at Uitenhage, Port Elizabeth, Witbergen and Onderbokkeveld, and placed *P. glomerata* and *Lachnaea*

conglomerata in synonymy under *P. ericoides*. This revision by Meisner (1857) was largely followed by Wright (1915). Because of his incorrect concept of the taxon, Meisner (1857) was confronted with material from Table Bay and Standvallei with red berries, which he then named *Chymococca empetroides*, based especially on the fleshy fruit. Thoday (1924a) was justified in placing this name in synonymy under *P. ericoides*, as the descriptions of these taxa coincide and as the fleshy fruit of *C. empetroides* is not unique, but is also found in *P. rigida*. The concept of *P. ericoides*, occurring along coastal dunes mainly in the Cape Peninsula and adjacent coastal areas of the Western Cape, was clarified by Thoday (1924a) and is also accepted in the present study.

Diagnostic characters: Passerina ericoides is characterized by greenish flowers, with a coriaceous, strigose hypanthium and the fruits are fleshy red berries. The leaves are greyish green and oblong, with an obtuse apex. The bracts are leaf-like, larger and lanceolate.

Etymology: the specific epithet ericoides refers to the ericoid appearance of this species indicated by the phrase 'corollae tubus globosus, inflatus—unde et Ericam refert flore', which was used by Linnaeus (1767) in his original description of the species.

Common names: Willdenow (1799) introduced the vernacular name heideartiger Vogelkopf, and the common names 'Christmas berry' or dronkbessie were documented by Smith (1966).

Uses: Marloth (1925) remarked that P. ericoides was laden with bright, scarlet fruits and that it was often employed as a Christmas decoration. The juicy pulp has a somewhat unpleasant taste, but appears to be harmless (dronkbessie). As early as 1919, Sim recommended P. ericoides as a useful shrub for planting in coastal areas exposed to sea winds. This species occurs on coastal dunes and on the banks of lagoons in the Cape Peninsula and adjacent coastal areas of Western Cape. The plants are excellent sand binders as they have an extensive root system from which resprouting often takes place. Because human impact and invasion of alien vegetation along the coast of the Cape Peninsula are very high, rehabilitation and conservation of coastal dunes is of vital importance. P. ericoides plants are ideally suited to combat erosion of coastal dunes and can be used as a substitute in coastal areas where alien vegetation is cleared. In their research on the coastal erosion of the Milnerton beaches, Biggs et al. (2001) made use of P. ericoides, occurring on the mobile dunes of this area as a natural monitor to indicate coastal erosion.

Distribution and ecology: Passerina ericoides ranges from Melkbosstrand along the coast of the Cape Peninsula to De Mond in the Bredasdorp District (Figure 7). It is endemic to the Southwestern and Agulhas Plain Centres within the CFR. This species occurs on littoral sand between rocks, or in dune valleys between the primary and secondary dunes.

According to a note on the specimen *Taylor 4042*, *P. ericoides* forms part of the Coastal Fynbos (Acocks 1988), currently divided by Rebelo (1998) into the

Laterite Fynbos of the Elim Flats, the Limestone Fynbos on calcareous sands overlying the limestone and associated calcretes of the Bredasdorp Formation, and the Sand Plain Fynbos from the Olifants River Mouth to Muizenberg on the West Coast lowlands. According to Rebelo (1998) the southernmost centre of the Sand Plain Fynbos is almost engulfed by the Cape Town Metropolitan Area and the area between Milnerton and Malmesbury must rank as one of the world's hottest spots for the loss of plant biodiversity.

Conservation status: Lower Risk (LR-nt) (Victor 2002), as a large portion of the coast along the Cape Peninsula is affected by human impact and invasion by alien vegetation associated with the Cape Town Metropolitan Area.

7. Passerina rigida Wikstr. in Kunglinga Svenska Vetenskapsakademiens Handlingar 39: 326 (1818); Meisn.: 402 (1840); Steud.: 274 (1841); Drège: 208 (1843); C.A.Mey.: 49 (1843); Meisn.: 563 (1857); C.H.Wright: 13 (1915); Thoday: 165 (1924a); Palmer & Pitman: 1585 (1972); Coates Palgrave: 649 (1977); Bond & Goldblatt: 433 (1984); A.E.van Wyk & P.van Wyk: 68 (1997). Type: Hab. ad Promont bonae spei [Cape Peninsula], Prof. Sparrman s.n., Herb. Thunb. 9578 (UPS, holo.!; M!, S!).

P. eriophora Gand.: 418 (1913); Thoday: 165 (1924a). Types: Natal [KwaZulu-Natal], Stanger; Natal, in ora prope Durban [coast near Durban], Wood 1712 (K!, lecto., designated by Thoday: 165 (1924a), BM!, BOL!, M!, W!); Wood 6592 (S!).

P. ericoides sensu Meisn.: 562 (1857) pro parte quoad specim. Drège s.n. (MO!), non L.

Many-stemmed, much-branched, robust shrubs of (0.6–)1.0–2.0(–3.04) m tall on coastal dunes; secondary and tertiary branches ascending, conical in appearance, formation of branchlets profuse, decussate, older branchlets self-pruning, lax or arcuate, 60–100 mm long, progressively shortening towards growing point, young branchlets ascending, 5–60 mm long, growing point nodding, fertile branchlets often pendulous and secund. *Stems* greyish brown, bark stringy; cork grey-brown; branchlets and growing points densely white-tomentose,

tomentum forming lengthwise patterns with cork on older branchlets, which later become glabrous. Leaves imbricate, overlapping ± 50%, appressed, plane shape lanceolate to ovate, length \times depth 1.6-2.5 \times 0.1-1.1 mm, adaxial surface concave, villous, abaxial surface convex, glabrous, greyish green, smooth, often covered by salt crystals; base sessile, dilated; median vein in distal half visible as a keel, forming acute apex; margins villous. Inflorescences with spikes usually extended, 6-10flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts appressed, ascending in fruit, widely ovate, length × depth (2.6–)3.4 \times 1.4(-1.9) mm; lamina adaxially concave (inside), abaxially convex (outside), villous inside, glabrous outside, smooth, with 2 or 3 shallow folds on each side of main vein, wings absent, greyish green, coriaceous; base dilated; apex with distinct, short, acute point; margins tomentose, involute. Floral envelope ± 4 mm long, membranous and yellow during pollination, dehydrated after shedding of pollen, turning red. Hypanthium glabrous at ovary, neck tomentose, ± 0.8 mm long. Sepals: outer sepals cymbiform, midrib adaxially tomentose, abaxial surface glabrous, inner sepals obovate, adaxially tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl ± 0.5 mm and those of antisepalous whorl ± 1.5 mm long; anthers, $\pm 0.8 \times 0.4$ mm, sub-basifixed, 2-thecous and 4-locular. Ovary 2.2 × 1.4 mm. Fruit a fleshy yellow berry, $\pm 2.6 \times 2.3$ mm, enveloped by persistent, loosely arranged hypanthium, fragmenting over widest circumference of fruit, the fragmented hypanthium, sepals and androecium being shed. Seed ± 1.4×1.1 mm. Figure 8.

Nomenclatural notes: in his description of P. rigida, Wikström (1818) clearly indicated the specimen of Sparrman, in the Thunberg Herbarium, as the type. This specimen bears the inscriptions Passerina glomerata β and epithet rigida in pencil. Thoday (1924a) identified the handwriting of the pencilled 'rigida' as Wikström's, comparing it to signed letters in the library at Kew. Wikström's handwriting was also confirmed in the present study, using examples published by Burdet (1979). According to Stafleu & Cowan (1986), original specimens of the Thunberg Herbarium (to which Sparrman also contributed) were donated to UPS and the duplicates

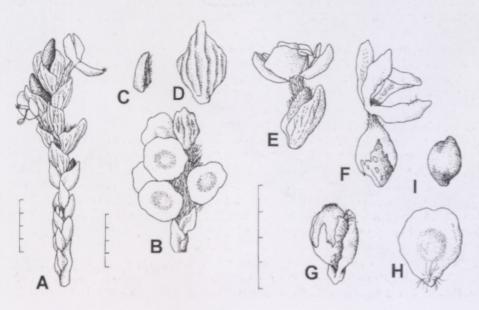


FIGURE 8.—Passerina rigida, Bredenkamp 1013. A, flowering inflorescence; B, fruiting inflorescence; C, leaf; D, bract; E, flower clasped by bract; F, hypanthium fragmenting at circumference of ovary. G, H, fruit: G, clasped in tomentum of bract; H, yellow, fleshy berry. I, seed. Scale bars: 4 mm. Artist: A. Stadler.

were sent to S. Thus three other *Sparrman s.n.* specimens of Herb. Swartzii, Herb. Wikströmii and Herb. Gaströmii, housed at S, and a fourth one from Schreber's herbarium, housed at M, are duplicates. As Wikström clearly indicated the specimen in the Thunberg Herbarium as the type, we regard it as the holotype and the other four Sparrman specimens as isotypes.

Diagnostic characters and relationships: Passerina rigida is easily distinguished as a robust, rigid shrub, usually 1–2 m high. The ascending branches are conical in shape due to many branchlets that are pendulous when fertile. The flowers are yellow and membranous and bright yellow berries are borne subterminally. The leaves are narrowly lanceolate to ovate and the apex is acute, with the main vein visible as a blunt keel. The bracts are widely ovate with the apex acute. This species is easily distinguished from P. paleacea which occurs on secondary dunes and is distributed mainly along the southern coast of Western Cape. Plants of the latter species are less robust, reaching a maximun height of 1.5 m, characterized by an abundance of subcapitulate inflorescences and the fruits are dry (achenes).

Etymology: the epithet *rigida* refers to the rigid, ascending branches, characteristic of the growth form of this plant.

Common names: Smith (1966) recorded the vernacular names gonnabas and seekoppiesgonna, while both Palmer & Pitman (1972) and Coates Palgrave (1977) added the name 'dune gonna'. Palmer & Pitman (1972) also listed the names ishoba and unyenyevu. The names 'dune-string', duinetaaibos and gonnabos were used by Lubke & Van Wijk (1998). Duin-gonna, 'dune gonna', inwele, unyenyevu are names given by Von Breitenbach et al. (2001).

Uses: Passerina rigida is a pioneer of the coastal dunes along large portions of the South African coast. Because these robust plants are excellent sand binders and are completely adapted to maritime winds and salt spray, they can be used in the rehabilitation of coastal dunes in disturbed areas. P. rigida has an extensive root system from which resprouting commonly takes place.

The yellow berries are an important food source for animals inhabiting coastal areas, especially birds.

Distribution and ecology: Passerina rigida is distributed from Witsand River Mouth on the western coast of the Cape Peninsula, along the coastline of South Africa to Lake Sibayi on the northeastern coast of KwaZulu-Natal (Figure 9). It is endemic to the coastlines of KwaZulu-Natal, Eastern Cape and Western Cape. The specimen Taylor 4143, recorded as far North as Lambert's Bay on the West Coast, is regarded as an outlier, as no other specimens have been recorded in the grid 3318. Thoday (1924a) mentioned Bowker s.n. from Somerset, Cooper 2301 from Albany and Ecklon & Zeyher s.n. (SAM 19801) as specimens from inland localities. In recent years more cases of P. rigida growing along sandy banks of rivers adjacent to the coast have been noted.

This species occurs on littoral sand dunes and hammock dunes just above the level of spring tide. It is also found in marshy places and on sandy banks of river mouths and lagoons. A stunted form is present on shallow marine sand over limestone and on rocky hills facing the sea. Lubke & Van Wijk (1998) regard P. rigida on the southern and Eastern Cape coast as a pioneer found in bush clumps or bush pockets on rear dunes. According to them, there are often no pioneer communities on the vast dune sands and the first vegetation encountered as one moves away from the shore is dune thicket, in which P. rigida is one of the dominant shrubs. Passerina species occurring on littoral dunes in Western Cape are found mainly in Coastal Fynbos (Acocks 1988). From the southern Cape coast to Port Alfred, dune fynbos and dune thicket form a mosaic as well as a successional series between the two vegetation types (Lubke & Van Wijk 1998).

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

8. **Passerina nivicola** *Bredenk.* & *A.E.van Wyk* in Bothalia 32: 77 (2002c). Type: Western Cape, 3319 (Worcester): Ceres Dist., Waboomberg, 1 760 m, (–DD),

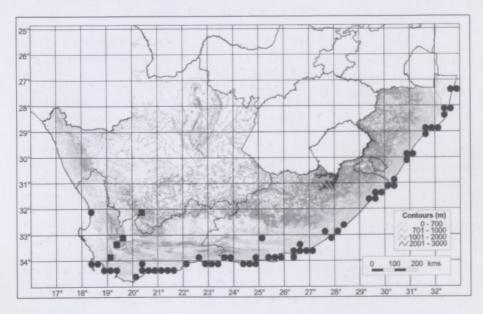


FIGURE 9.—Known distribution of Passerina rigida, ●; P. nivicola, ■.

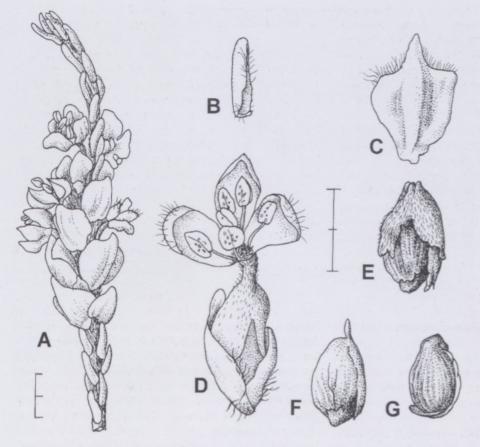


FIGURE 10.—Passerina esterhuyseniae, Esterhuysen 26859.
A, inflorescence with apex
growing out, returning to
vegetative growth (proliferating growth); B, leaf; C,
bract; D, flower clasped by
bract showing fragmentation
of hypanthium at circumference of fruit; E, fruit with
remnants of hypanthium. F,
G, achene: F, enveloped by
membranous pericarp; G, lateral view. Scale bars: 2 mm.
Artist: A. Stadler.

12 November 1989, E.G.H. Oliver 9281 (PRE!, holo.; NBG!, iso.).

Passerina sp. nov. 1 Bredenk. & A.E.van Wyk: 70 (2000); Bredenk. & A.E.van Wyk: 56 (2001a); Bredenk. & A.E.van Wyk: 217 (2001b).

For a complete description see Bredenkamp & Van Wyk in *Bothalia* 32: 76–79 (2002c). Distribution (Figure 9).

Conservation status: Lower Risk [LR-lc] (Victor 2002). This species is rare, but does not qualify for Red List status under IUCN (2000) guidelines.

9. Passerina esterhuyseniae Bredenk. & A.E.van Wyk, sp. nov., P. comosae C.H.Wright affinis. Bracteae galeiformes, circumscriptione obovatae; lamina adaxialiter concava, abaxialiter convexa, intra setosa, extra glabra, tenuiter chartacea, laevis, concolorans, brunneola, in marginem membranaceum vel in alas membranaceas expansa; basis cuneata; costa excurrens apicem subacutum vel acutum faciens; margines ciliati dimidio superiore. Flores membranacei, tempore pollinationis flavidi, postea rubri ad brunnei.

TYPE.—Western Cape, 3218 (Clanwilliam): N Cederberg Mountains, (–BB), Groenberg near Pakhuis, along base of high rugged rock, rocky slopes, ± 1 167 m, 27-12-1956, *Esterhuysen 26859* (BOL, holo.!).

Passerina sp. nov. 2 Bredenk. & A.E.van Wyk: 70 (2000); Bredenk. & A.E.van Wyk: 56 (2001a); Bredenk. & A.E.van Wyk: 217 (2001b).

Shrubs or shrublets 0.3–0.5 m high. *Stems* greyish brown, younger branchlets greyish tomentose; cork finely fissured, grey-brown, displaying whitish sclerenchy-

ma fibres at scars. Leaves imbricate on young branchlets, closely appressed to stem, cymbiform, plane shape linear-lanceolate, length × depth ± 2.0 × 0.5 mm; lamina inversely ericoid, adaxial surface concave, setose, abaxial surface convex, glabrous; base sessile; apex rounded into subacute point; margins sometimes ciliate. Inflorescences polytelic synflorescences; main florescences and co-florescences spicate. Bracts enveloping flowers and fruits, largest after anthesis of flowers, becoming more coriaceous and rounded at fruit set, decussate, imbricate, sessile, helmet-shaped, widely obovate in outline, length × depth ± 3.1 × 2.4 mm; lamina adaxially concave (inside), abaxially convex (outside), setose on inside, glabrous on outside, thinly chartaceous, smooth, concolorous, brownish, extending into a membranous rim or membranous wings; base cuneate; main vein extending to form a subacute to acute apex; margins ciliate in distal half. Floral envelope constituting hypanthium (fused calyx and androecium) and sepals; membranous and yellowish during pollination, dehydrated after shedding of pollen, becoming papyraceous, turning red to brown, ± 4.6 mm long. Hypanthium a membranous cylindric tube, indumentum at ovary and neck tomentose, neck ± 0.7 mm long, abscission tissue and articulation plane absent. Sepals 4, petaloid, imbricate in bud, flexed in flower; outer sepals concave oblong with apex adaxially tomentose, abaxially setose; inner sepals concave, obovate with apex adaxially glabrous, abaxially setose. Androecium: filaments of antipetalous whorl ± 0.4 mm and those of antisepalous whorl ± 1.2 mm long. Ovary ± 1.8×0.5 mm. Fruit enveloped by persistent, loosely arranged hypanthium fragmenting over widest circumference of fruit, the fragmented hypanthium, sepals and androecium being shed; an achene with pericarp membranous and dry, $\pm 2.5 \times 1.2$ mm. Figure 10.

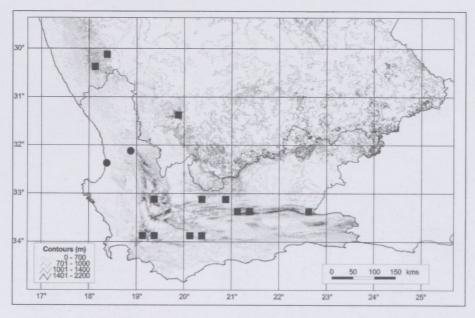


FIGURE 11.—Known distribution of Passerina esterhuyseniae, •;

Diagnostic characters and relationships: Passerina esterhuyseniae is easily distinguished from P. comosa by its helmet-shaped bracts, which are widely obovate in outline. The concolorous, brownish bracts are thinly chartaceous and smooth in texture, the lamina extends into a membranous rim or membranous wings and the main vein elongates forming a subacute to acute apex. The flowers are membranous and yellowish during pollination and red to brown after shedding of the pollen.

Eponymy: this species is dedicated to Elsie Esterhuysen who diligently collected plants especially the high-mountain flora of the Northern, Western and Eastern Cape.

Distribution and ecology: Passerina esterhuyseniae has been collected on the northern Cederberg Mountains at Groenberg near Pakhuis and at Konpoort (Figure 11). It is endemic to the Northwestern Centre within the CFR. The northern Cederberg area is covered by Mountain Fynbos (Rebelo 1998). This species grows at the peaks of mountain tops at altitudes of ± 1 167 m, or against rocky slopes amongst high rugged rocks. Confined mostly to mountainous areas, this species is still undercollected. Pillans 7689 (BOL) collected on slopes near the road SE of Redelinghuis has been classified under P. esterhuyseniae, although these plants seem to be more robust and grow at lower altitudes.

Conservation status: Lower Risk (LR-lc) (Victor 2002), as the population size of this species is probably very small or restricted.

10. **Passerina comosa** (*Meisn.*) *C.H.Wright* in Flora capensis 5,2: 11 (1915); Thoday: 158 (1924a); Bond & Goldblatt: 432 (1984); Goldblatt & Manning: 683 (2000). Type: Little Namaqualand; Khamiesberg Range, between Pedros Kloof and Leliefontein, 3000–4000 ft., *Drège* 2570 [K!, lecto., designated by Thoday: 388 (1924b); PRE!].

P. filiformis L. var. comosa Meisn.: 399 (1840); Meisn.: 562 (1857); C.H.Wright: 11 (1915); Thoday: 158 (1924a).

P. falciformis Drège: 68 (1843) nom. nud.

Small shrubs, (0.3–)0.6–0.9(–1.06) m high, branches often virgate. Stems grey-brown; cork fissured lengthwise, grey-brown, surrounding prominent leaf scars; indumentum of young branches densely white or yellowish tomentose, flaking off with cork on older branchlets, which become glabrous. Leaves imbricate, overlapping ± 20%, diverging at an angle of 20°, plane shape linear to narrowly lanceolate, length \times depth (2.7–)3.0(–4.0) \times 0.6-0.9 mm, adaxial surface concave, villous, abaxial surface convex, tomentose to villous, rugose or warty with bases of fallen hairs; base sessile; apex obtuse to subacute; margins glabrous, basally setose. Inflorescences with spikes usually extended, 6-12-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts appressed, widely ovate, length × depth $(3.0-)5.5 \times 1.5(-1.8)$ mm; lamina adaxially concave (inside), abaxially convex (outside), villous inside, tomentose to villous outside, coriaceous, rugose with bases of fallen hairs, ± 3-ribbed on each side of main vein, margins submembranous or wings extended and abaxially setose to villous; base sessile; apex acute; margins glabrous, basally setose, involute. Floral envelope ± 7.3 mm long, yellow-pink during pollination, dehydrated after shedding of pollen, turning red. Hypanthium glabrous at ovary, neck tomentose, ± 0.9 mm long. Sepals: outer sepals cymbiform, inner sepals elliptic, outer and inner sepals adaxially glabrous, abaxially tomentose. Androecium with filaments of antipetalous whorl \pm 0.7 mm and those of antisepalous whorl \pm 1.5 mm long; anthers 0.7×0.4 mm, sub-basifixed, 2-thecous and 4-locular. Ovary $\pm 2.7 \times 1$ mm. Fruit an achene with pericarp membranous and dry, ± 2.5 × 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in the sepals and androecium being shed. Figure 12.

Diagnostic characters and relationships: Passerina comosa and P. quadrifaria both have abaxially hairy bracts and are easily confused. However, these two species are geographically segregated, with P. comosa considered as a 'north-western endemic' of the Cape flora (Weimarck 1941), whereas P. quadrifaria is endemic to the Karoo Mountain and Southeastern Centres. Mor-

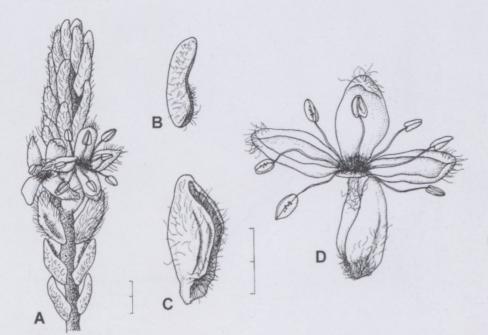


FIGURE 12.—Passerina comosa, Andraea 1288. A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower clasped by bract. Scale bars: 2 mm. Artist: G. Condy.

phologically *P. comosa* is less robust, internodes are longer, leaves adhere closely to the stem and are generally more hairy and the bracts often have extended wings that are abaxially setose to villous.

Etymology: the epithet comosa refers to the hairs on the abaxial surface of the leaves, bracts and sepals, which are characteristic of this species.

Distribution and ecology; Passerina comosa ranges from mountain summits and slopes of the Kamiesberg to Calvinia in the Northern Cape (Figure 11). In Western Cape it is distributed in the area between 33° and 34°S latitude and from 19° to 21°E longitude, with Primos 41 (PRE) as the most easterly outlier. This species is endemic to the Northern Cape, as well as the Northwestern, Southwestern and Karoo Mountain Centres within the CFR. It occurs on the Roggeveld, Witteberg and the Klein Swartberg Mountain Ranges. This species is found in sand among rocks, on rocky ledges, on mountain summits, or on SW-facing slopes at altitudes of 1 000–1 200 m.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

11. Passerina pendula Eckl. & Zeyh. ex Thoday in Bulletin of Miscellaneous Information, Kew 4: 155 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 684 (2000). P rigida Wikstr. var. comosa Meisn.: 402 (1840); Drège: 584 (1847a); Meisn.: 563 (1857); C.H.Wright: 13 (1915) pro parte; Thoday: 155 (1924a). Type: Uitenhage Div., Zwartkops River, Ecklon & Zeyher 7381 [K!, lecto., designated by Thoday: 387 (1924b); BREM!, P!, PRE!, WU!].

P. pendula Eckl. & Zeyh. fide Drège: 210 (1847b) nom. nud.; Meisn.: 563 (1857) nom. nud.

Erect, many-stemmed shrubs, 1.0–1.5 m high, young branchlets often pendulous. *Stems* greyish brown; indumentum of young stems densely white-tomentose, flaking off on older branchlets, which become glabrous, remaining indumentum forming lengthwise patterns with

grey-brown cork; brown leaf scars prominent. Leaves greyish green, imbricate, overlapping ± 25%, appressed, plane shape rhombic, length \times depth 2.5–3.4 \times 1.0–1.2 mm, adaxial surface concave, comose, abaxial surface convex, softly coriaceous and smooth, glabrous; base sessile, abruptly tapered; apex subacute; margins brownish setose. Inflorescences with multiflowered main and co-florescences; spikes usually extended, 6-16-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts greyish green when fresh, grey-brown in dried specimens, appressed, rhombic, length \times depth (3.0–)4.2 \times 1.0(–1.5) mm; lamina adaxially concave (inside), abaxially convex (outside), comose inside, glabrous outside, softly coriaceous and smooth, wings membranous; base cuneate; apex obtusely angled; margins brownish setose. Floral envelope ± 6.5 mm long, membranous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium glabrous at ovary, neck tomentose, ± 0.6 mm long. Sepals: outer sepals cymbiform, adaxially scantily tomentose, abaxially glabrous; inner sepals oblong, adaxially scantily tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl ± 1.5 mm and those of antisepalous whorl ± 2 mm long; anthers $\pm 0.8 \times 0.4$ mm, sub-basifixed, 2-thecous and 4-locular. Ovary ± 2.0 × 0.7 mm. Fruit an achene with pericarp membranous and dry, ± 2.5 × 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in the sepals and androecium being shed. Figure 13.

Nomenclatural notes: Passerina pendula, ascribed to Ecklon & Zeyher, was first published as a nomen nudum by Drège (1847b). This name was placed in synonymy under P. rigida var. comosa by Meisner (1857). Wright (1915) partly followed Meisner's interpretation of P. rigida var. comosa, but in the citation of the specimens he added all those that were later published as P. burchellii by Thoday (1924a). In his revision of Passerina, Thoday (1924a) reinstated the name P. pendula Eckl. & Zeyh. ex Meisn., as the varietal name 'comosa' had already been used at species level by Wright (1915). The present study

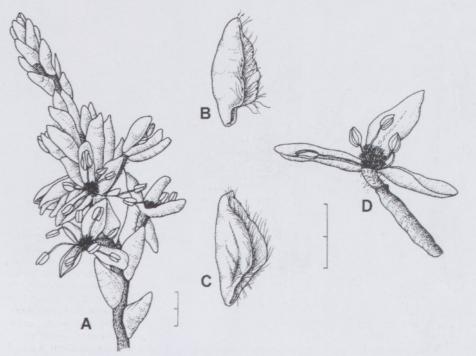


FIGURE 13.—Passerina pendula, Bredenkamp 908. A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower. Scale bars: 2 mm. Artist: G. Condy.

regards Thoday's publication of *P. pendula* as valid, as it is accompanied by a Latin diagnosis, a description of the species and it was published in 1924. As *P. pendula* Eckl. & Zeyh. ex Meisn. was based on a *nomen nudum*, and since Thoday ascribed the name to Ecklon & Zeyher, the correct author citation for this species is *P. pendula* Eckl. & Zeyh. ex Thoday.

Diagnostic characters and relationships: Passerina pendula is distinguished from P. burchellii by being taller (up to 1.5 m), much-branched shrubs with pendulous branchlets, with grey-green, softly coriaceous and smooth leaves and yellow-pink membranous flowers that are abaxially glabrous and adaxially scantily tomentose.

Etymology: the specific epithet pendula refers to the pendulous branchlets of these shrubs as seen in their natural habitat.

Distribution and ecology: Passerina pendula is endemic to the Southeastern Centre within the CFR. It is distributed on hills and slopes from the Kouga Mountains in Western Cape to the Langkloof Mountains and the

Great Winterhoek Mountains in Eastern Cape (Figure 14). The species is also distributed along watercourses as it occurs in the KwaZunga-Catchment Basin and on the banks of the Upper Swartkops River as well as the Bushmans River at Port Elizabeth. *P. pendula* grows at altitudes of (133–)383–600 m. On mountain slopes it is often found in a belt above valley thicket and below mountain fynbos. It grows in sand or shallow, gravelly, sandy loam. The plants are frequent throughout the natural range of the species and a number of populations are conserved in the Groendal Nature Reserve at Uitenhage.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

12. **Passerina galpinii** *C.H.Wright* in Flora capensis 5,2: 10 (1915) as *P. galpini*; Thoday: 161 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 683 (2000). Type: Cape [Western Cape]; Riversdale Dist., Melkhoutfontein, about 600 ft, 7 October 1897, *Galpin 4491* (K, holo.!; PRE!).

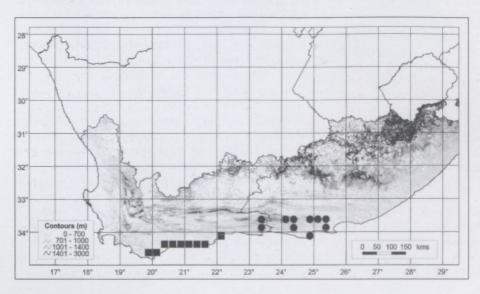


FIGURE 14.—Known distribution of Passerina pendula, ●; P. galpinii, ■.

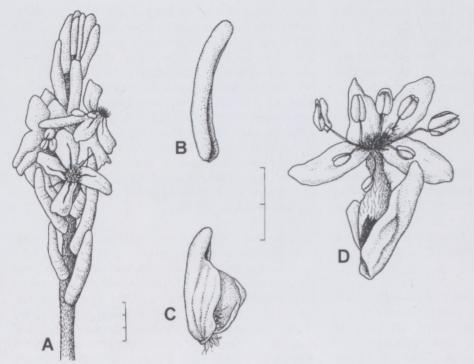


FIGURE 15.— Passerina galpinii, Bredenkamp 946. A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower clasped by bract, hypanthium fragmenting at circumference of ovary. Scale bars: A, 3 mm; B–D, 2 mm. Artist: G. Condy.

Stunted or erect, much-branched shrubs, (0.1–)0.3–1.0 (-1.2) m high. Stems light greyish brown; indumentum of young stems grevish brown tomentose; cork greybrown, fissured lengthwise, with sclerenchyma fibres protruding between fissures; leaf scars rounded. Leaves greyish green, imbricate, overlapping 10%-25%, diverging at an angle of 30°, plane shape linear, incurved towards apex, length × depth 2.9-4.5 × 0.5-0.8 mm, adaxial surface concave, comose, abaxial surface convex, chartaceous, glabrous; base sessile; apex subacute to obtuse; margins glabrous, involute. Inflorescences with spikes subterminal, usually congested, 8-10-flowered, axis whitish tomentose, proliferating growth common. Bracts appressed, oblate, length \times depth (3.6–)4.5 \times 1.1(-2.2) mm; lamina greyish green, adaxially concave (inside), abaxially convex (outside), chartaceous, glabrous outside, midrib extended into leaf-like point, tomentose inside; wings straw-coloured, membranous, broadly rounded, bullate; base cuneate; apex subacute; margins glabrous, involute. Floral envelope ± 5.3 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium pubescent at ovary, neck pubescent, ± 1 mm long. Sepals: outer sepals concave elliptic, glabrous, inner sepals obovate, abaxially glabrous, apex margin adaxially tomentose. Androecium with filaments of antipetalous whorl ± 1.4 mm and those of antisepalous whorl \pm 2.1 mm long; anthers \pm 0.6 \times 0.5 mm, sub-basifixed, 2thecous and 4-locular. Ovary ± 2.0 × 1.1 mm. Fruit an achene with pericarp membranous and dry, ± 2.5 × 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at circumference of ovary, resulting in the sepals and androecium being shed. Figure 15.

Nomenclatural notes: Passerina galpini C.H.Wright (1915), published with a full description, but without a Latin diagnosis, was accepted by Thoday (1924a). In the present study the specific epithet is corrected to 'galpinii' and the name accepted as validly published by Wright, as the starting date for a Latin diagnosis as pre-

requisite for valid publication is 1 January 1935 (Greuter et al. 2000).

Diagnostic characters and relationships: Passerina galpinii is distinguished by its characteristic bracts, which are oblate, with the midrib extended into a leaf-like point; the lamina is cymbiform, greyish green, chartaceous and glabrous, with the midrib adaxially tomentose; the wings are straw-coloured, membranous, broadly rounded and bullate. The distribution of this species is also diagnostic as it is endemic to the Agulhas Plain Centre within the CFR.

Eponymy: Passerina galpinii was named in honour of the botanist E.E. Galpin. The holotype of this name, Galpin 4491, was collected on 7 October 1897. At this time, Galpin organized a collecting trip from Port Elizabeth via the Humansdorp, Knysna, George, Riversdale, Swellendam and Caledon Districts to Cape Town and increased his collecting numbers from 3531 to 4846. All these specimens were probably identified at the Bolus Herbarium in Cape Town, where he also spent a few weeks (Gunn & Codd 1981).

Common name: Rebelo (1998) mentions the vernacular name Elim gonna for this species.

Distribution and ecology: Passerina galpinii is endemic to the Agulhas Plain Centre within the CFR. It is distributed on stony flats, coastal limestone deposits and limestone hills, from Elim to Bredasdorp, Arniston, Stilbaai, Melkhoutfontein, Albertinia and Mossel Bay (Figure 14); it grows at altitudes of 0–290 m. Plants reach a height of ± 1.2 m on stony flats, but become stunted on southeast-facing slopes of limestone hills, overlooking the sea. The plants are frequent in their natural environment. They are conserved in the De Hoop and Potberg Nature Reserves and several private nature reserves. The vegetation of the area is threatened by large stands of Acacia cyclops (rooikrans), an alien invasive tree.

Passerina galpinii is associated with Laterite Fynbos (Rebelo 1998), occurring on the Elim Flats of Western Cape, which is characterized by gravelly, lateritic and seasonally waterlogged soils. The present study also indicates the presence of this species in Limestone Fynbos, where it occurs on coastal limestone deposits.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

13. **Passerina drakensbergensis** *Hilliard* & *B.L.Burtt* in Annals of the Kirstenbosch Botanic Gardens 15: 233 (1987); Hilliard & B.L.Burtt: 88 (1988). Type: Natal [KwaZulu-Natal], Harrismith, Bergville Dist., Cathedral Peak Forest Reserve, Ndedema River, 6000 ft, 5 November 1985, *Hilliard* & *Burtt* 18570 (E, holo.; K!, M!, P!, PRE!, S!).

Erect, many-stemmed shrubs, ± 2 m high, muchbranched. Stems grey-brown; indumentum of young stems closely white-tomentose, flaking off on older branches, forming lengthwise patterns with patches of yellowish brown cork; leaf scars prominent. Leaves greyish green (glaucous), imbricate, overlapping ± 15%, appressed, plane shape linear to lanceolate, longitudinaly folded and somewhat keeled, length \times depth (3.5–) $4.2-5.5(-6.5) \times 0.8-2$ mm, adaxial surface concave, tomentose, abaxial surface slightly convex, rugose, glabrous; base sessile, abruptly narrowed; apex subacute, sometimes incurved; margins setose. Inflorescences with multiflowered main and co-florescences; spikes usually extended, 8–12-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts greyish green (glaucous), appressed, lanceolate, length × depth $4.75-6.0(-7.0) \times 1.5$ mm; lamina folded lengthwise and keeled, adaxially concave (inside), abaxially slightly convex (outside), villous inside, glabrous outside, coriaceous, obscurely ribbed and reticulately veined; base cuneate; apex obtuse to acute; margins sometimes narrowly membranous. Floral envelope ± 5.9 mm long; papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. *Hypanthium* scantily tomentose at ovary, neck tomentose, \pm 1.7 mm long. *Sepals*: outer and inner sepals concave lanceolate, outer sepals adaxially glabrous, abaxially tomentose; inner sepals adaxially and abaxially tomentose. *Androecium* with filaments of antipetalous whorl \pm 1.2 mm and those of antisepalous whorl \pm 2 mm long; anthers oblong, $0.9(-1.25) \times 0.3$ mm, sub-basifixed, 2-thecous and 4-locular. *Ovary* \pm 2.4 \times 0.9 mm. *Fruit* an achene with pericarp membranous and dry, \pm 2.5 \times 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in the sepals and androecium being shed. Figure 16.

Diagnostic characters and relationships: Passerina drakensbergensis is characterized by appressed leaves, up to 6.5 mm long. The bracts are lanceolate, up to 7 mm long, the apex is obtuse to acute without a leaf-like point and membranous wings are absent. It may easily be confused with P. montivaga and P. montana, both occurring in the northern KwaZulu-Natal Drakensberg area. P. montivaga has longer leaves (up to 8 mm) and bracts with the midrib extended, forming a straight or filiform. leaf-like point. The wings are ovate with margins hairy in the distal half, or obtrullate, narrowing abruptly into the midrib. P. montana can be separated by its terminal subcapitulate spikes and short leaves (up to 4 mm long), which are linear to lanceolate, with a dilated base and with a prominent median vein in the upper third of the leaf, incurved at the acute apex. The bracts are ovate to obovate in outline.

Passerina montivaga is a fynbos element which possibly originated in the southern Cape and dispersed eastwards. Both this species and P. montana are distributed from Eastern Cape via the Drakensberg Mountains northwards to Zimbabwe. Although the distribution of P. drakensbergensis, P. montivaga and P. montana overlap in the northern part of the KwaZulu-Natal Drakensberg, significant trends in the geographical and altitudinal ranges of these species have been identified.

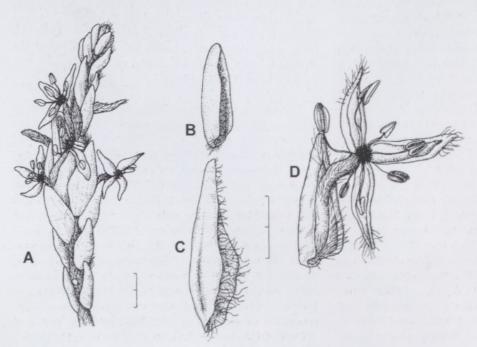


FIGURE 16.— Passerina drakensbergensis, Bredenkamp 1021.
A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower clasped by bract. Scale bars: 2 mm. Artist: G. Condy.

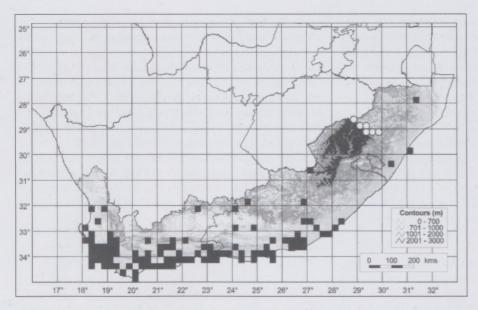


FIGURE 17.—Known distribution of *Passerina drakensbergensis*, O; *P. corymbosa*, ■.

Etymology: the specific epithet refers to the location of this species in the northern KwaZulu-Natal Drakensberg.

Distribution and ecology: Passerina drakensbergensis is endemic to the Bergville District in the northern KwaZulu-Natal Drakensberg (Figure 17). It has been collected from the Royal Natal National Park to Giant's Castle Game Reserve at altitudes between 1 500 and 1 980 m. It is frequent at Cathedral Peak above Ndedema Gorge, where it is found in a marginal belt between forest and grassland. It also grows along streams and riverbanks and on mountain slopes.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

14. **Passerina corymbosa** *Eckl.* ex *C.H.Wright* in Flora capensis 5,2: 14 (1915); Thoday: 162 (1924a). Type: Cape [Western Cape], Bredasdorp, Zwellendam, Pupas Valley, Voormansbosch and Duivelsbosch, Keurboomsrivier, October, *Ecklon & Zeyher 40* (70.10) (B, holo.†; W!, lecto., here designated; MO!, S!).

P. filiformis L. var. vulgaris Meisn.: 399 (1840); Meisn.: 562 (1857): Drège: 87 (1843). P. vulgaris (Meisn.) Thoday: 162 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 684 (2000). Type: locis subhumidis secus rivulum, prope Paarlberg, alt. 1000–1500 ped. [subhumid locality along river, near Paarlberg, alt. 1000–1500 ft], Nov.—Dec. 1839, Drège s.n. [K, lecto., designated by Thoday: 388 (1924b); S!, W!].

P. filiformis sensu C.H. Wright: 11 (1915) pro parte, non L.

Much-branched shrubs, (0.3–)1.0–1.5(–2.5) m high, younger branchlets leafy, older ones leafless, often arcuate, indurate. *Stems* often arising from a woody, underground rootstock, greyish brown; indumentum of young stems closely whitish tomentose, forming lengthwise patterns with cork on older branches, which gradually become glabrous; cork fissured lengthwise, with sclerenchyma fibres protruding between fissures; leaf scars oblate. *Leaves* greyish green, drying greyish brown, ascending, diverging at an angle of ± 40°, imbricate, overlapping up to 50%, laterally compressed, plane shape linear to narrowly lanceolate, length × depth (2.0–)3.2–4.4

 $(-10.0) \times 0.5$ –0.8 mm, adaxial surface concave, tomentose, abaxial surface glabrous; lamina longitudinaly folded, midrib distinct and somewhat keeled, with 2 or more lateral veins visible as faint striae when fresh, base sessile; apex tapered or acute; margins glabrous, involute. Inflorescences with multiflowered main and co-florescences; spikes usually extended, 10-16-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts greyish green, ascending, rhombic to obtrullate, distinctly angled, length × depth (4.4–) $5.1 \times 1.4(-1.7)$ mm; lamina folded lengthwise and keeled, adaxially concave (inside), abaxially slightly convex (outside), shortly villous inside, glabrous outside, coriaceous; wings coriaceous, distinctly 4- or 5-ribbed; base cuneate; midrib shortly extended into an acute apex; margins glabrous, involute. Floral envelope ± 6.2 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium glabrous at ovary, neck scantily tomentose, ± 1.6 mm long. Sepals: outer sepals cymbiform, inner sepals obovate, outer and inner sepals adaxially scantily tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl ± 0.7 mm and those of antisepalous whorl ± 1.4 mm long; anthers ovoid, ± 0.9 × 0.4 mm, sub-basifixed, 2-thecous and 4-locular. Ovary $\pm 2.1 \times 1.1$ mm. Fruit an achene with pericarp membranous and dry, ± 2.6 × 1.5 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in the sepals and androecium being shed.

Diagnostic characters and relationships: Passerina corymbosa is distinguished by its greyish green leaves and grey-brown stems of which the older branchlets are leafless, often arcuate and indurate. The leaves are laterally compressed with the distinct midrib somewhat keeled. The rhombic to obtrullate (diamond-shaped) bracts are always conspicuously angled and distinctly 4- or 5-ribbed. The most diagnostic character in the leaf anatomy of P. corymbosa is the presence of a hypodermal sclerenchymatous sheath, illustrated by Bredenkamp & Van Wyk (2001a). This species has always been confused with P. filiformis sensu lato, which has inconsistently been distinguished by longer, filiform leaves. In Western Cape P. filiformis subsp. filiformis is separated

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by widely obovate bracts, narrowing abruptly into a filiform point. Where the distribution of *P. corymbosa* and *P. montivaga* overlap in the southern Cape, the latter species is distinguished by bracts with ovate wings and margins that are hairy in the distal half. In Eastern Cape *P. montivaga* is distinguished by obtrullate bracts narrowing abruptly into a straight, leaf-like point.

Etymology: of all the species in the genus, Passerina corymbosa is the most common, as it is adapted to a wide range of habitats mostly in Western and Eastern Cape. The specific Latin epithet corymbosa (= with a cluster of flowers or of fruits) indicates the 10–16-flowered, extended spikes usually arranged in multiflowered main and co-florescences.

Common names: according to Van Wyk & Gericke (2000), P. corymbosa (formerly known as P. vulgaris) is also called bakkersbos, a name that commemorates an era when the official bakers in the Cape used this plant to heat up their outdoor ovens.

Uses: from an agricultural point of view, Story (1952) described P. corymbosa at Keiskammahoek as an unpalatable bush, which remained undamaged from grazing, among the few closely cropped specimens of Cliffortia linearifolia and C. paucistaminea. However, the value of P. corymbosa as a pioneer, and also in combatting erosion, cannot be underestimated. This species is commonly found along roadsides and in other disturbed places. It is one of the most successful species for the rehabilitation of embankments along newly built roads in Western and Eastern Cape. The plants are resprouters from woody, underground rootstocks and are excellent sand binders, often found on coastal sand dunes. Considering the human impact and invasion of alien vegetation along the Cape coast, P. corymbosa would be a natural pioneer, combatting erosion in areas where alien vegetation is cleared.

Distribution and ecology: except for a few outliers, P. corymbosa is endemic to Western and Eastern Cape, and all the phytogeographic centres within the CFR. Although this species is distributed from Clanwilliam to Cape Town and eastwards to East London, it most commonly occurs in an area between the coast and the 33°S latitude and from 18° to 29°E longitude (Figure 17). Gerstner 105 (PRE), collected near Compasberg in the Lady Grey District, represents the most northerly distribution of *P. corymbosa* in Eastern Cape. The specimens collected in KwaZulu-Natal are regarded as outliers, representing remnants of a former wider distribution. Hilliard 4081 (PRE), collected at the Ellesmere Farm in Ngome (KwaZulu-Natal), is an anomalous specimen, with a greyish appearance, infected by fungi and recorded from cliff faces. This specimen was classified as P. corymbosa on the basis of the angular bracts and the leaves that are laterally compressed. The other two specimens, Herb. Poeppig s.n., probably collected before 1868, and Rudatis 1204 (PRE), collected in 1910, represent populations that have possibly succumbed to human impact.

Passerina corymbosa is a species with a wide habitat spectrum. It most commonly occurs as a pioneer along

roadsides over the whole range of its distribution. The species is found in stony areas on mountain slopes, peaks and mountain passes. Along the coastal region, it is often found on the rear dunes. It also grows in river valleys and on the banks of river mouths. This species is common in the whole of the Fynbos Biome of the CFR. In Eastern Cape it is found in all the above-mentioned habitats, but also in grassland. Story (1952) reported that *P. corymbosa* is found in sourveld and mixed grassveld but that it showed no sign of advancing into the sweetveld. In open grassland this species is often clustered along streambanks or on rocky areas. *P. corymbosa* occurs at a range of altitudes, from sea level up to 1 300 m.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

15. **Passerina obtusifolia** *Thoday* in Bulletin of Miscellaneous Information, Kew 4:157 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 683 (2000). Type: Cape [Western Cape], Worcester Div., between Osplaats and Tunnel Sidings 2000–3000 ft., *Rogers 16703* [K!, lecto., designated by Thoday: 388 (1924b)].

Erect shrubs with several fairly stout stems, bare at base, branching profusely higher up, (0.3-)0.8-1.8(-2.4)m high. Stems ash-grey; indumentum of young stems whitish tomentose, flaking off, becoming scabrid on older branchlets and forming lengthwise patterns with cork, which fissures lengthwise; sclerenchyma fibres protruding between fissures; leaf scars oblate. Leaves greyish green, imbricate, spreading at an angle of 45°, plane shape linear, straight or incurved, length x depth $4.0-8.0(-12.0) \times 0.9-1.5$ mm, adaxial surface concave, comose, abaxial surface convex, coriaceous, glabrous; base sessile; apex obtuse; margins glabrous, involute. Inflorescences with multiflowered main and co-florescences; spikes usually extended, 8-16-flowered, arrangement terminal, axis whitish tomentose, proliferating growth common. Bracts spreading, narrowly obtrullate, length \times depth (4.0–)5.8 \times 1.5(–1.7) mm; lamina greyish green, adaxially concave (inside), abaxially convex (outside), coriaceous, glabrous outside, tomentose inside. closely 2-ribbed at margins, midrib extending beyond lamina into a leaf-like point, apex obtuse; wings absent; base cuneate; margins glabrous, involute. Floral envelope ± 6.8 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium glabrous at ovary, neck tomentose, ± 1.8 mm long. Sepals: with outer sepals cymbiform, adaxially tomentose, abaxially glabrous; inner sepals narrowly oblong, adaxially tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl ± 1.2 mm and those of antisepalous whorl ± 2.2 mm long; anthers \pm 1.1×0.5 mm, sub-basifixed, 2-thecous and 4-locular. Ovary $\pm 2.5 \times 1.1$ mm. Fruit an achene with pericarp membranous and dry, ± 2.3 × 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at circumference of ovary, resulting in sepals and androecium being shed. Figure 18.

Diagnostic characters and relationships: the growth form of *P. obtusifolia* may easily be confused with that of *P. corymbosa*, but *P. obtusifolia* is distinguished by

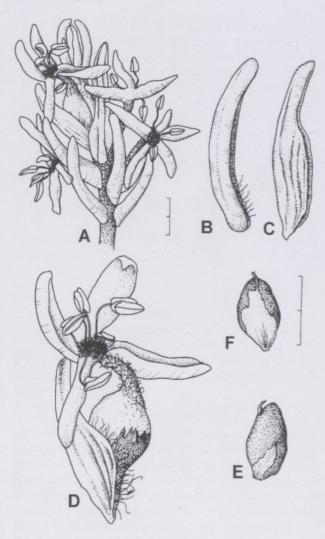


FIGURE 18.—Passerina obtusifolia, Bredenkamp 919. A, flowering inflorescence; B, leaf; C, bract; D, flower clasped by bract, fragmenting at circumference of ovary. E, F, achene: E, lateral view, with basal fragment of membranous pericarp; F, ventral view, with remnant of membranous pericarp. Scale bars: 2 mm. Artist: G. Condy.

the spreading, linear, incurved leaves, with obtuse apices. The bracts are narrowly obtrullate, with the midrib extending beyond the lamina into a leaf-like point and the lamina is closely 2-ribbed at the margins. Intermediate forms between *P. obtusifolia* and *P. trunca*-

ta subsp. truncata are found in the Karoo Desert National Botanical Garden at Worcester. P. obtusifolia usually occurs on the northern side of the southern Cape mountain ranges in drier habitats and P. falcifolia is confined to the summits of mountains and southwards towards the southern Cape coast. Intermediates between P. obtusifolia and P. falcifolia have been found on the boundary between the two species, just north of the Prince Albert Pass.

Etymology: the Latin specific epithet, *obtusifolia*, refers to the obtuse apices of leaves and bracts, which are characteristic of this species.

Common name: the vernacular name karoo gonna is used by the local people at Genadendal.

Uses: according to the curator at the Museum in Genadendal, P. obtusifolia was traditionally used by the local people in the home industry of soap-making. The plants were burnt and the alkaline ashes used to react with the stearic acid in fat at boiling point, thus forming soap. In the Robertson area these plants are used in the wild flower industry. Bayliss 521 (PRE) is a voucher specimen recorded in cancer research, but the results must have been negative; these plants are not currently known for their medicinal value.

Distribution and ecology: Passerina obtusifolia is endemic to Northern, Western and Eastern Cape (Figure 19). It is centred in a belt between 33° and 34°S latitude and from 19° to 27°E longitude, comprising all the centres within the CFR—it is most common in the Karoo Mountain, Langeberg and Southeast Centres. P. obtusifolia is distributed from Worcester in Western Cape, to Grahamstown in Eastern Cape. The most northwesterly distribution are the Vanrhyn's Pass and the Hantamberg. The Karoo National Park and the Nuweveld Mountains in the Beaufort West District and the Lootsberg Pass at Middelburg are the most northerly distribution in Western and Eastern Cape. Hilliard & Burtt 14654 and Acocks 20170 from the Lady Grey District are considered as outliers.

Passerina obtusifolia is well adapted to drier karoo habitats and is often found on the north-facing aspect of mountains in the southern Cape. It frequents the bound-

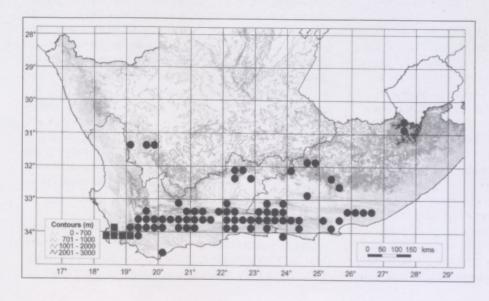


FIGURE 19.—Known distribution of Passerina obtusifolia, ●; P. paludosa, ■.

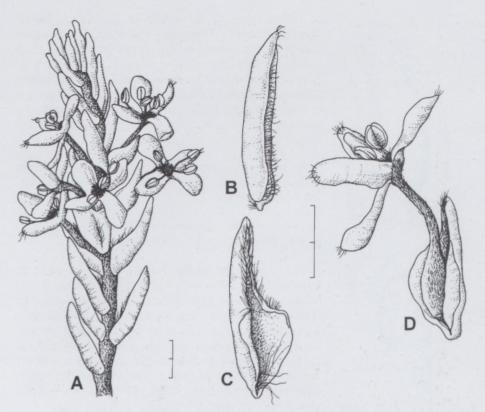


FIGURE 20.—Passerina paludosa, Jangle 156. A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower clasped by bract. Scale bars: 2 mm. Artist: G. Condy.

ary of fynbos and karroid vegetation and is common in the Little Karoo, growing at altitudes of (300–)670–1 400 (-1 700) m. Although this species occurs at high altitudes on the summit of the Swartberg Pass, it grows below the snow line and does not occur on the highest peaks of mountain ranges in its distribution range. It is common in drier mountainous habitats, growing in shallow rocky soil and between rocks on well-drained slopes. It also grows amongst sandstone boulders of upper mountain slopes and on stony ridges of mountain tops. On the Hantamberg it has been recorded in renosterveld on the flat, rocky, dolerite summit. On Jonaskop it grows in a zone below the fynbos and is absent at the summit. This species is also found amongst rocks in river valleys and dry streambanks. The average height of these plants is 0.8-1.8 m, but stunted forms have been recorded from the arid Bergkwagga National Park, which is one of the most northeasterly localities. P. obtusifolia is a very common species and amongst the dominant species within its distribution range.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

16. Passerina paludosa *Thoday* in Bulletin of Miscellaneous Information, Kew 4: 161 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 683 (2000). Types: Cape [Western Cape], Simonstown, Cape Flats, Riet Valley, in and near shallow vleis, November 1922, *Thoday 100* [K!, lecto., designated by Thoday: 388 (1924b); C!, NBG!, PRE!].

Much-branched shrubs or small tree, up to 2 m high. *Stems* fawn-coloured; indumentum of young stems closely white-tomentose, forming lengthwise patterns with cork on older branches, which gradually become glabrous; leaf scars oblate, comose at upper rim. *Leaves*

erect, nearly straight, greyish green, imbricate, overlapping ± 25%, appressed, plane shape narrowly lanceolate to lanceolate, longitudinaly folded and somewhat keeled, length \times depth (3.0–)5.5–6.8(–10) \times 0.8–1.4 mm, adaxial surface concave, tomentose, abaxial surface glabrous; base sessile; apex acute, sometimes incurved, bearing a persistent tuft of white, erect hairs; margins setose. Inflorescences with multiflowered main and co-florescences; spikes usually extended, 10-12-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts greyish green, appressed, narrowly obtrullate with leaf-like point, length x depth $(5.2-)7.0 \times 1.8(-2.0)$ mm; lamina folded lengthwise and keeled, adaxially concave (inside), abaxially slightly convex (outside), basally setose inside, glabrous outside, coriaceous, ± 2-ribbed; wings membranous with obscure venation, glabrous; base cuneate; apex acute; margins white setose. Floral envelope ± 7.2 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium shortly tomentose at ovary, neck strigose, ± 2.6 mm long. Sepals: outer sepals cymbiform, ad- and abaxially glabrous, apex setose; inner sepals oblong, ad- and abaxially glabrous, apex setose with margins tomentose. Androecium with filaments of antipetalous whorl \pm 1.2 mm and those of antisepalous whorl \pm 2.4 mm long; anthers ovoid, $\pm 0.7 \times 0.5$ mm, sub-basifixed, 2the cous and 4-locular. Ovary $\pm 2.3 \times 1$ mm. Fruit an achene with pericarp membranous and dry, $\pm 2.4 \times 1.2$ mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in sepals and androecium being shed. Figure 20.

Diagnostic characters and relationships: Passerina paludosa is a stout shrub up to 2 m high, occurring mostly in marshy ground on lowland flats. It is characterized by erect, nearly straight, greyish green, imbricate, ap-

pressed leaves, which are \pm lanceolate. The bracts are narrowly obtrullate, with the midrib and leaf-like point stout and the apex acute. This species is distinguished from P. filiformis subsp. filiformis which has filiform leaves and widely obovate bracts, which narrow abruptly into a filiform point.

Etymology: the specific epithet paludosa refers to the habitat of this species, namely marshy lowland flats; from the Latin paludosus (= marshy, swampy or boggy).

Distribution and ecology: Passerina paludosa is endemic to the Southwestern Centre within the CFR (Figure 19). Herbarium specimens dated from 1921 to 1995 show that this species used to be distributed from sandy places along the Malmesbury Road (Acocks 2482), along marshy areas of the Cape Flats and the Stellenbosch District to the Palmiet River at Elgin, the most easterly locality. As P. paludosa was severely affected by urbanization and invasion by alien vegetation in the Cape Peninsula, it is currently confined to small marshy areas east of Muizenberg.

According to Smuts (1996) the only three extant populations known, are at the Rondevlei Nature Reserve, Zeekoevlei and along the Strandfontein Road. Label information on Peterson 1263, collected in 1982, states that the population at a housing estate site SE of Zeekoevlei consisted of ± 400 plants, but Smuts (1966) reported only 60 living plants. At the same time the population at Rondevlei consisted of 35 plants and the one along the Strandfontein Road of possibly a few hundred. Currently both the Zeekoevlei and Strandfontein sites are in danger of urban development and are being threatened by invasive alien vegetation, primarily Port Jackson (Acacia saligna) and rooikrans (A. cyclops). Conservation measures proposed by Smuts (1996) include an environmental impact study at the Zeekoevlei site prior to any development and a plea for urgent attention by conservation authorities to ensure the conservation of the Strandfontein population.

The Rondevlei Nature Reserve boasts more than 250 plant species of which many are rare and endangered. Species associated with *P. paludosa* include *Chondropetalum nudum, Juncus krausii* and *Leucadendron levisanus*. In recent years the management at the reserve concentrated on restoring and managing its biodiversity. Alien vegetation has been cleared, plant species that occurred there historically have been re-introduced and *P. paludosa* has been successfully propagated by cuttings to expand the population. As aridification is an important effect of urbanization and as alien vegetation impacts on the natural drainage system of an area, the whole wetland east of Muizenberg can be conserved only if it is included in the Rondevlei Nature Reserve (Smuts 1996).

Recently two new populations of plants, that appear to be *P. paludosa*, were collected at Springfontein Farm near Stanford [3419AD, *Louw 7083* (NBG, PRE)], and in seasonally wet clays at Heidehof, 5 km NW of Pearly Beach [3419CB, *Helme 2376* (NBG, PRE)]. These specimens were not included in the distribution of *P. paludosa* as further population studies need to be done. Taking

urbanization and invasion by alien vegetation into account, the Red List status of *P. paludosa* was also not changed.

Conservation status: Vulnerable, [VU B1B2abcd] (Victor 2002).

17. **Passerina montivaga** *Bredenk.* & A.E.van Wyk in Bothalia 32: 34 (2002d). Type: Natal [KwaZulu-Natal], 2930 (Pietermaritzburg): hills above Pinetown, 2400 ft, (–DD), 3 December 1891, *J.M. Wood in PRE 49409* (PRE!, holo.; MO!).

P. montivagus Bredenk. & A.E.van Wyk: 34 (2002d), sphalm.

P. filiformis L.: 559 (1753) pro parte, excluding type; Thunb.: 75 (1794): Wikstr.: 324 (1818); Thunb.: 374 (1825a); Meisn.: 562 (1857); C.H.Wright: 10 (1915); Thoday: 159 (1924a); Bond & Goldblatt: 432 (1984); Hilliard & B.L.Burtt: 182 (1987); Goldblatt & Manning: 683 (2000).

Passerina sp. nov. 4 Bredenk. & A.E.van Wyk: 70 (2000); Bredenk. & A.E.van Wyk: 56 (2001a); Bredenk. & A.E.van Wyk: 217 (2001b).

For a complete description see Bredenkamp & Van Wyk in *Bothalia* 32: 34 (2002d).

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

18. Passerina filiformis L. in Species plantarum: 559 (1753); Thunb.: 75 (1794): J.C.Wendl.: 18 (1798); Wikstr.: 324 (1818); Thunb.: 374 (1825a); Meisn.: 562 (1857); C.H.Wright: 10 (1915); Thoday: 159 (1924a); Palmer & Pitman: 1587 (1972); Coates Palgrave: 648 (1977); Bond & Goldblatt: 432 (1984); Hilliard & B.L.Burtt: 182 (1987); A.E.van Wyk & P.van Wyk: 68 (1997); Goldblatt & Manning: 683 (2000). Bredenk. & A.E.van Wyk 32: 29 (2002d). Type: Passerina filiformis, Linnean Herbarium 504.1 [LINN, lecto.!, designated by Thoday: 388 (1924b)].

P. cupressina J.C.Wendl. nom. nud. Meisn.: 404 (1840); Meisn.: 563 (1857); Thoday: 159 (1924a). P. cupressoides Steud.: 273 (1841).

P pectinata Lodd.: 18 (1816) nom. nud. Wikstr.: 347 (1818); Meisn.: 404 (1840); Meisn.: 562 (1857); Thoday: 159 (1924a).

For a complete description see Bredenkamp & Van Wyk in *Bothalia* 32: 29–34, figs 1–4 (2002d).

18a. subsp. filiformis

For a complete description see Bredenkamp & Van Wyk in *Bothalia* 32: 31–33, figs 1–3 (2002d).

18b. subsp. **glutinosa** (Thoday) Bredenk. & A.E.van Wvk

For a complete description see Bredenkamp & Van Wyk in Bothalia 32: 33–34, fig. 4 (2002d).

Conservation status: Lower Risk, [LR-nt] (Victor 2002).

19. **Passerina falcifolia** (*Meisn.*) *C.H.Wright* in Flora capensis 5,2: 10 (1915); Thoday: 158 (1924a); Palmer & Pitman: 1585 (1972); Coates Palgrave: 648

(1977); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 683 (2000). *P. filiformis* L. var. *falcifolia* Meisn.: 399 (1840); Drège: 118, 124 (1843); Meisn.: 562 (1857); C.H.Wright: 10 (1915); Thoday: 158 (1924a). Type: Eastern Cape, Uitenhage, Vanstaadesberg [now Van Staden's Mountains], 1000–2000 ft, *Drège s.n.* (K!, lecto., designated by Thoday: 388 (1924b)].

P. filiformis L. var. *divaricata* Wikstr.: 325 (1818); Thoday: 158 (1924a). Type: Caput bonae Spei [Cape Peninsula], *Sparrman s.n.*, *Herb. Thunberg 9573* (UPS, holo.!), *Herb. Swartzii* (S, iso.!).

Much-branched, tall shrubs or small ornamental trees, (1.0-)1.2-2.0(-3.04) m high, branchlets often arcuate, pendulous. Stems grey-brown; indumentum of young stems closely whitish tomentose, forming lengthwise patterns, with cork on older branches, which gradually become glabrous, fissuring lengthwise, yellowish brown lenticulae present; leaf scars oblate. *Leaves* grevish green. drying greyish brown, falcate, linear to narrowly lanceolate, longitudinaly folded, triangular in section, length × depth 3.9-5.6 × 0.6 mm, adaxial surface concave, tomentose, abaxial surface glabrous; base sessile; apex obtuse or tapered; margins glabrous, involute; younger leaves inclined, diverging at an angle of ± 60°, imbricate, overlapping up to 50%, older leaves horizontally spreading, not imbricate. Inflorescences with multiflowered main and co-florescences; spikes usually extended, lax, often arcuate, up to 16-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts mottled grey-green, rose-tinted during flowering time, ascending, imbricate, widely ovate, narrowing into a leaflike, falcate point, length \times depth (5.3–)5.8 \times 1.7(–2.0) mm; older bracts folded lengthwise along midrib and keeled, younger bracts adaxially concave (inside), abaxially slightly convex (outside), villous inside, glabrous outside, coriaceous; wings chartaceous, distinctly ± 4ribbed and reticulately veined; base cuneate; apex obtuse or tapered; margins ciliate in distal half. Floral envelope ± 8.4 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium scantily tomentose at ovary, neck

exserted, often arcuate, tomentose, \pm 3 mm long. *Sepals*: outer sepals concave, obovate, adaxially scantily tomentose, abaxially glabrous; inner sepals obovate, adaxially tomentose, abaxially glabrous. *Androecium* with filaments of antipetalous whorl \pm 1.4 mm and those of antisepalous whorl \pm 2.1 mm long; anthers ovoid, \pm 1 × 0.4 mm, sub-basifixed, 2-thecous and 4-locular. *Ovary* \pm 2.5 × 0.7 mm. *Fruit* an achene with pericarp membranous and dry, \pm 2.5 × 1.4 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in sepals and androecium being shed. Figure 21.

Nomenclatural notes: Wright (1915) overlooked the combination *P. filiformis* L. var. *divaricata* Wikstr. (1818), also indicated by Thoday (1924a), which is the earliest name for the taxon. However, this name based on *Sparrman s.n.* (Herb. Thunberg 9573) falls into synonymy under *P. falcifolia*, as the name of a taxon does not have priority outside the rank in which it was published (Greuter *et al.* 2000).

Diagnostic characters and relationships: Passerina falcifolia is distinguished by the mottled grey-green falcate leaves, which are inclined or horizontally spreading. The widely ovate bracts are villous inside and narrow into a leaf-like, falcate point, with chartaceous wings that are distinctly ± 4-ribbed and reticulately veined. The most conspicuous floral character is the slender, often arcuate, tomentose hypanthium neck, exserted from the clasping bract. This species may be confused with P. filiformis subsp. filiformis which has widely obovate bracts, narrowing abruptly into a filiform point. The bracts are basally to centrally setose on the inside and the wings are glabrous. P. montivaga is another close species, but is distinguished by bracts that are basally setose on the inside, with glabrous wings.

Etymology: the specific epithet falcifolia is derived from the Latin falcatus (= curved like a sickle), referring to the falcate or sickle-shaped leaves of these plants.

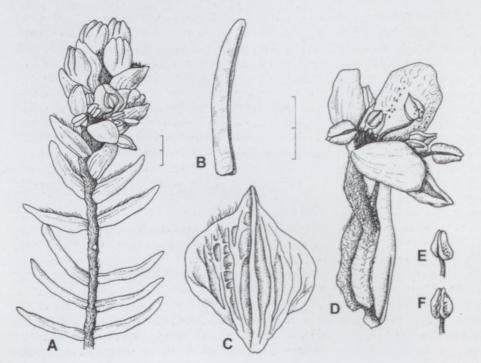


FIGURE 21.—Passerina falcifolia, Bredenkamp 917. A, flowering inflorescence; B, leaf; C, bract; D, flower clasped by bract. E, F, anther: E, dorsal view; F, ventral view. Scale bars: 2 mm. Artist: G. Condy.

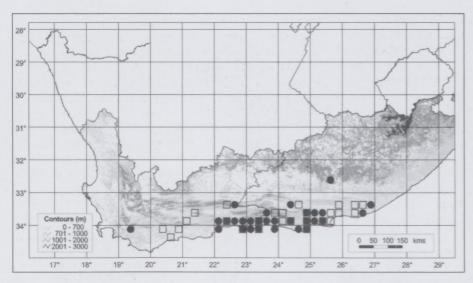


FIGURE 22.—Known distribution of *Passerina falcifolia*, ●; *P. rubra*, □.

Common names: Passerina falcifolia is also known as the 'Outeniqua gonna' or the Outeniekwagonna, referring to the Outeniqua Mountains where it occurs (Coates Palgrave 1977). Palmer & Pitman (1972) use the vernacular name forest gonna, as these plants are commonly seen along roadsides on mountain passes of the southern Cape forests. Von Breitenbach et al. (2001) use the names Outeniekwa-gonna, gonnabas, gonnabos, kannabas and Outeniqua gonna, gonna bush.

Uses: Passerina falcifolia is used for fuel or for making cord (Palmer & Pitman 1972). According to label information on Dahlstrand 1905 (PRE), the species is cultivated by florists. Plants grow into small ornamental trees and could be used more widely in horticulture. According to Grobbelaar 63 (PRE), P. falcifolia is a host to members of the insect genus Eremnus.

Distribution and ecology: Passerina falcifolia is associated with forests and Mountain Fynbos (Rebelo 1998) in the southern Cape and the southern parts of Eastern Cape. It is a near-endemic to the CFR and occurs in the Karoo Mountain, Southwestern and Southeastern Centres, as well as the Zuurberg, Blaauwkrantz and Alexandria Forests of Eastern Cape. It most commonly occurs in a belt between the coast and the 33°S latitude and from 22° to 26°E longitude (Figure 22). The two specimens, Brown 25975 and Rogers 28858, collected near Caledon in October 1924, are regarded as outliers, possibly indicating a wider previous distribution of the species into areas with woody vegetation in Western Cape. P. falcifolia occurs from Meiringspoort, in the Oudtshoorn area, to Ruytersbosch in the Mossel Bay area, and along the Outeniqua, Tsitsikamma and Great Winterhoek Mountains to the Grahamstown area.

Passerina falcifolia is found on mountain plateaus and southeast-facing slopes on Table Mountain Sandstone in shallow, sandy loam soil. Plants commonly occur along forest margins, in open patches, or disturbed areas along roadsides. This species is also found in coastal regions and riverine fynbos. P. falcifolia grows at a range of altitudes, from sea level up to 1 100 m.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

20. Passerina rubra *C.H.Wright* in Flora capensis 5,2: 12 (1924a); Thoday: 156 (1924a); Bond & Goldblatt: 433 (1984); Goldblatt & Manning: 684 (2000). Type: Cape [Western Cape], Ladismith, Muiskraal, near Garcia Pass, 1200 ft, 3 October 1897, *Galpin 4492* [K!, lecto., designated by Thoday: 388 (1924b); GRA!, PRE!].

P. filiformis L. var. *squarrosa* Meisn.: 562 (1857); Thoday: 156 (1924a). Type: Eastern Cape, prope Cradockstad et Port-Elisabeth [near Cradock Town and Port Elizabeth], October, *Zeyher* 3779 (S!, lecto., here designated; BM!, MEL!, NBG!, W!).

Much-branched, erect shrubs, with rigid branchlets and inflorescences or smaller, extensively branched, rounded shrublets under arid, calcareous habitat conditions, (0.2-)0.3-0.75(-1.1) m high. Stems: older ones grey-brown, indurate, and sclerenchyma fibres exposed; young stems reddish brown, indumentum whitish tomentose, forming lengthwise patterns with cork on older branches, which gradually become glabrous; cork fissuring lengthwise; internodes longer than leaves during prolific lengthening of branchlets or shorter under arid conditions. Leaves greyish green, ascending, appressed, decussate and rigid, or under arid conditions, imbricate (overlapping 5–30%), appressed or ascending, diverging at an angle of up to 30°; lamina narrowly lanceolate or oblong, longitudinaly folded, triangular in section, length \times depth 2.4–4.3 \times 0.7 mm, adaxial surface concave, tomentose, abaxial surface glabrous; base sessile; apex obtuse; margins glabrous, involute. Inflorescences with conspicuous, multiflowered main and co-florescences; spikes robust, rigid, extended, narrowly ellipsoid, with rows of enlarged, decussate, pointed bracts, 20-30-flowered, arrangement subterminal, axis white-tomentose, proliferating growth common. Bracts grey-green, rose-tinted during flowering time, ascending, imbricate, widely ovate, midrib shortly extended into a point, length \times depth (4.3–)5.1 \times 1.8(-2.0) mm; older bracts folded lengthwise along midrib, younger bracts adaxially concave (inside), abaxially slightly convex (outside), villous inside, glabrous outside, coriaceous; wings widely ovate, chartaceous, ± 5-ribbed, reticulately veined; base cuneate; apex acute; margins ciliate in distal half. Floral envelope ± 8.4 mm long, papyraceous and yellow-pink during pollination, dehydrated after shedding of pollen, turning red to brown. Hypanthium glabrous at ovary, neck exserted, sparsely pubescent, ± 2 mm long. Sepals: outer sepals cymbiform,

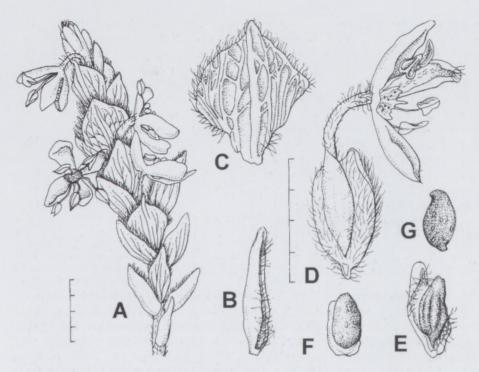


FIGURE 23.—Passerina rubra, Bredenkamp 914. A, inflorescence with apex growing out, returning to vegetative growth (proliferating growth); B, leaf; C, bract; D, flower clasped by bract; E, fruit clasped in tomentum of bract, enveloped by hypanthium, fragmented at neck base. F, G, achene: F, enveloped by membranous pericarp; G, side view. Scale bars: 4 mm. Artist: A. Stadler.

ad- and abaxially glabrous, inner sepals obovate, adaxially scantilly tomentose, abaxially glabrous. Androecium with filaments of antipetalous whorl \pm 0.7 mm and those of antisepalous whorl \pm 1.7 mm long; anthers ovoid, \pm 0.9 \times 0.3 mm, sub-basifixed, 2-thecous and 4-locular. Ovary \pm 2.7 \times 1.1 mm. Fruit an achene with pericarp membranous and dry, \pm 2.1 \times 1.2 mm, enveloped by persistent, loosely arranged hypanthium, breaking up at neck base due to dehydration and torsification of tissue, resulting in the sepals and androecium being shed. Figure 23.

Nomenclatural notes: as the starting date for a Latin diagnosis is 1 January 1935 (Greuter et al. 2000), P. rubra is a valid name, although it was published with a full description, but without a Latin diagnosis, by Wright (1915). The combination P. filiformis L. var. squarrosa (Meisner 1857), was overlooked by Wright (1915), but mentioned in synonymy by Thoday (1924a). In the present revision all the type material cited by both Wright (1915) and Meisner (1857) was studied. Galpin 4492 (K) was selected as the lectotype of P. rubra by Thoday (1924b) and Zeyher 3779 in S was selected as lectotype for P. filiformis var. squarrosa as it is internationally available in many herbaria.

Diagnostic characters and relationships: the distribution of P. rubra partly coincides with that of P. corymbosa, P. montivaga and P. falcifolia. P. rubra is a smaller shrub (average height 0.3-0.75 m), often occurring in calcareous soil. It is distinguished from the other three species which are taller (average heights 1-2 m), and especially from P. falcifolia, which is a tall shrub or a small tree (up to 3.04 m), often associated with indigenous forests. P. rubra may also be separated by the inflorescences which have extended, robust spikes, with up to 30 fertile, enlarged bracts. The bracts are typified by the midrib which is shortly extended into a point and by the wings which are adaxially tomentose, widely ovate, chartaceous, ± 5-ribbed and reticulately veined. Flowers are distinguished by the exserted hypanthium neck, which is ± 2 mm long and glabrous to sparsely pubescent.

Etymology: the specific epithet rubra was derived from the Latin ruber (= red), referring to the conspicuous, multiflowered inflorescences of these plants, which have 20–30 flowers arranged in four rows and turning red after wind pollination.

Uses: Passerina rubra is a pioneer which often occurs along roadsides or in disturbed places, e.g. close to the salt works in the vicinity of Port Elizabeth. It is also found on calcareous soils between Port Elizabeth and Cradock. In the Coega area, earmarked for industrial development, P. rubra might be a useful plant for combatting erosion.

Distribution and ecology: Passerina rubra is nearendemic to the CFR, occurring in the Langeberg, Karoo Mountain and Southeastern Centres, as well as southern parts of the Eastern Cape. It most commonly occurs in a belt between the coast and the 33°S latitude and from 20° to 26°E longitude. P. rubra is distributed from the Bontebok National Park in the Swellendam District, eastwards to Gowie's Kloof near Grahamstown (Figure 22). This species is somewhat variable. It was initially thought that plants in Western Cape were more rigid, with longer internodes and appressed leaves, which did not overlap, whereas those in Eastern Cape tended to be rounded shrublets, with imbricate, ascending leaves. After many specimens, from all parts of the range had been studied, no geographical or morphological discontinuity between the two forms could be shown, and it was decided that the morphological differences were probably due to plasticity. Plants growing in more arid conditions and calcareous soil, typical of the Port Elizabeth and Cradock areas, tend to be rounded much-branched shrublets, with short internodes and imbricate, ascending leaves. Under more favourable conditions in sandy loam, the plants are taller, less branched, internodes are longer and the appressed leaves do not overlap.

Passerina rubra is common in the Steytlerville, Humansdorp, Port Elizabeth and Grahamstown areas of Eastern Cape and less frequent in Western Cape. The area between Cradock and Port Elizabeth is renowned for the ancient dunes and flats, abounding in limestone. Acocks (1988) described the vegetation occurring on the limestone as False Fynbos (A70), also known as Mountain Fynbos or Grassy Fynbos (Rebelo 1998). *P. rubra* seems to be well adapted to the calcareous soils on which it occurs. These plants are often pioneers in disturbed areas and along roadsides, as in the Colchester, Coega and the Markman industrial areas of Port Elizabeth. At the Groendal Catchment Basin, this species occurs in grassland on sandstone and it is also found on semi-karroid, dry, rocky hillsides in the Baviaanskloof area. At the Bontebok National Park it is found in flat areas between fynbos species. *P. rubra* grows at altitudes of 70–700 m.

Conservation status: Least Concern (LC) (IUCN Species Survival Commission 2000).

DOUBTFUL NAME

Passerina hamulata Gand.: 418 (1913). Hab.: Cap, in dunis arenosis prope Wynberg [Western Cape, Simonstown, sand dunes near Wineberg], Bolus s.n. Note: P. paleacea, Bolus 2926, complies with this distribution, but the name P. hamulata is not written on this specimen, therefore P. hamulata was not placed in synonymy under P. paleacea.

EXCLUDED NAMES

Sources of information: 1, *Passerina* databases at National Botanical Institute; 2, literature cited in the list; 3, Royal Botanic Gardens, Kew. Index Kewensis on Compact Disc (1997).

Accepted names are in bold roman type and synonyms are in italics.

Lachnaea conglomerata L.: 560 (1753); 374 (1784); Willd.: 434 (1799); Wikstr.: 323 (1818); Thunb: 374 (1825a); Meisn.: 562 (1857); Cafferty & Beyers: 171, 172 (1999); Brummitt: 805 (2000) nom. rejic. = **Phylica stipularis** L. (Rhamnaceae) in Cafferty & Beyers: 171 (1999).

P. ammodendron Kar. & Kir.: 444 (1842) = Stellera lessertii C.A.Mey. in Meisn.: 550 (1857). Iran.

P. annua Auch. ex Meisn.: 552 (1857) = **Thymelaea aucheri** Meisn.: 552 (1857); K.Tan: 235 (1980). Syria, Lebanon, W Iran.

P. annua (Salisb.) Wikstr.: 320 (1818) = **Thymelaea passerina** (L.) Coss. & Germ. in K.Tan: 236 (1980) [= *Thymelaea arvensis* Lam. in Meisn.: 551 (1857) (= *Stellera passerina* L.)]. Widespread: C and S Europe, SW Asia, C Russia.

P. anthylloides L.f.: 226 (1782) = **Gnidia virescens** Wikstr. in Meisn.: 592 (1857). South Africa: Cape Peninsula.

P. anthylloides Thunb.: 75 (1794) = **Gnidia virescens** Wikstr. in Meisn.: 592 (1857). South Africa: Cape Peninsula.

P. aragonensis Rouy: 123 (1910). Pyrenees.

P. argentata Pau: 67 (1922) = **Thymelaea argentata** (Lam.) Pau in K.Tan: 212 (1980). Spain and N Africa.

P. arvensis Ball: 653 (1878) = **Thymelaea passerina** (L.) Coss. & Germ. in K.Tan: 236 (1980) [= Thymelaea arvensis Lam. in Meisn.: 551 (1857) (= Stellera passerina L.)]. Widespread: C and S Europe, SW Asia, C Russia.

P. axillaris Thunb.: 106 (1792) = Pimelea virgata Vahl in Meisn.: 516 (1857). New Zealand.

Passerina baccata, Pappe s.n. (NBG!, SAM!) err. typogr. = P. ericoides L.

P. baccifera Mihi? nom. nud., Eckl. & Zeyh. s.n. (BOL!, P!, W!) = P. ericoides L.

P. bartlingiana Meisn. True identity unknown.

P. broteriana (Cout.) Sampaio & Da Silva: 104 (1913) = **Thymelaea broteriana** Cout.: 145 (1909); K.Tan: 227 (1980). Endemic to N and C Portugal.

P. bruniades Eckl. & Zeyh. ex Meisn.: 579 (1857) = **Lachnaea penicillata** Meisn.: 579 (1857); J.B.P.Beyers: 96 (2001). South Africa: Western Cape.

P. brunioides Eckl. & Zeyh. in Meisn.: 563, 579 (1857) = P. bruniades Eckl. & Zeyh. ex Meisn.

P. calocephala Meisn.: 393 (1840) = **Gnidia calocephala** (C.A.Mey.) Gilg: 226–228 (1894a). South Africa: KwaZulu-Natal and Eastern Cape.

P. calycina Lam. & DC.: 360 (1805) = **Thymelaea calycina** (Lapeyr.) Meisn.: 555 (1857); K.Tan: 226 (1980). C Pyrenees, rare.

P. campanulata E.Mey. ex Meisn.: 407 (1840) = **Lachnaea** grandiflora (L.f.) Baill. in J.B.P.Beyers: 45 (2001). South Africa: Western Cape.

P. canescens Schousb.: 190 (1800) = **Thymelaea lanuginosa** (Lam.) Ceballos & Vicioso in K.Tan: 211 (1980) [= *Thymelaea canescens* (Schousb.) Endl. in Meisn.: 556 (1857)]. S Spain, Morocco and Gibraltar.

P. cantabrica Pourr. ex Willk. & Lange: 299 (1862) = **Thymelaea coridifolia** (Lam.) Endl. in K.Tan: 226 (1980). Endemic to N Spain.

P. capitata L.: 88 (1760) = **Lachnaea capitata** (L.) Crantz in J.B.P.Beyers: 66 (2001). South Africa: Western Cape.

P. cephalophora Thunb.: 75 (1794) = **Lachnaea eriocephala** L. in J.B.P.Beyers: 98 (2001). South Africa: Western Cape.

P. chamaedaphne Bunge: 58 (1833) = **Wikstroemia canescens** Meisn.: 547 (1857). Nepal.

P. chamaejasme Fisch. ex Meisn.: 549 (1857) = **Stellera** chamaejasme L. in Meisn.: 549 (1857). Siberia.

P. chamaejasme Schangin: in Meisn.: 549 (1857) = Stellera altaica Thieb. in Meisn.: 549 (1857). Siberia.

P. ciliata L.: 559 (1753) = **Gnidia penicillata** Licht. ex Meisn. in B.Peterson: 476 (1959). South Africa: Western Cape.

P. ciliata Thunb.: 75 (1794) = *Lachnaea ciliata* (L.) Crantz in J.B.P.Beyers: 109 (2001), excluded species.

P. conglomerata (L.) Thunb.: 75 (1794); Wikstr.: 322 (1818) = **Phylica stipularis** L. (Rhamnaceae) in Cafferty & Beyers: 171 (1999).

Passerina coridifolia Wikstr.: 334 (1818) = Thymelaea coridifolia (Lam.) Endl. in K.Tan: 226 (1980). Endemic to N Spain.

P. cornucopiae = Pimelea cornucopiae Vahl in B.Hansen & P.Wagner: 352 (1998). Australia.

Passerina corsica J.Gay ex Litard. in Briq.: 6 (1938) = **Thymelaea tartonraira** All. subsp. **thomasii** (Duby) Briq. in K.Tan: 220 (1980) [= *Thymelaea tartonraira* All. var. calvescens Gren. & Godr. in Meisn.: 556 (1857)]. Circum-Mediterranean.

P. costata Griff.: 367 (1854) = **Diarthron vesiculosum** C.A.Mey. in Meisn.: 558 (1857). Iran.

P. diarthronoides Griff.: 365 (1854) = **Thymelaea passerina** (L.) Coss. & Germ. in K.Tan: 236 (1980) [= Thymelaea

- arvensis Lam. in Meisn.: 551 (1857) (= Stellera passerina L.)]. Widespread: C and S Europe, SW Asia, C Russia.
- *P. dichotoma* Steud.: 273 (1841) = **Stellera chamaejasme** L. in Meisn.: 549 (1857). Siberia.
- *P. dioica* Ramond: 139 (1800) = **Thymelaea dioica** (Gouan) All. in K.Tan: 222 (1980). W Pyrenees to SW Alps.
- *P. dodecandra* L.: 10 (1755) = **Struthiola dodecandra** (L.) Druce in Levyns: 599 (1950). South Africa: Eastern Cape and Western Cape.
- *P. elliptica* Boiss.: 556, t. 158 (1842) = **Thymelaea pubescens** (L.) Meisn. subsp. **elliptica** (Boiss.) K.Tan: 231 (1980) [= *T. elliptica* (Boiss.) Endl.: 66 (1847)]. Endemic to S Spain.
- *P. empetrifolia* Lapeyr.: 212 (1813) = **Thymelaea dioica** (Gouan) All. subsp. **dioica** in K.Tan: 222 (1980). W Pyrenees to SW Alps.
- *P. eriocephala* Thunb.: 75 (1794) = **Lachnaea globulifera** Meisn.: 576 (1857); J.B.P.Beyers: 107 (2001). South Africa: Western Cape.
 - P. filiformis Mill.: no. 1 (1768) = Struthiola sp.
- *P. filiformis* var. *ereifoifolia* Eckl. & Zeyh. *herb. no. 39* (G!), err. typogr. = **P. ericoides** L.
- *P. ganpi* Sieb. ex Miq. in Meisn.: 564 (1857) = **Stellera**? **ganpi** Meisn.: 550 (1857). Japan.
- *P. geminiflora* Ram. in K.Tan: 222 (1980) = **Thymelaea dioica** (Gouan) All. subsp. **dioica** in K.Tan: 222 (1980). W Pyrenees to SW Alps.
- *P. globosa* Lam.: 431 (1792) = **Lachnaea alpina** Meisn.: 578 (1857); J.B.P.Beyers: 59 (2001). South Africa: Western Cape.
- *P. gnidia* Forst.: 28 no. 170 (1786); Meisn.: 516 (1857) = **Pimelea gnidia** Banks et Sol. in Meisn.: 516 (1857). New Zealand.
- Passerina gnidia L.f.: 226 (1782) = Pimelea gnidia Banks et Sol. New Zealand.
- Passerina granatensis Pau: 7 (1904) = **Thymelaea dioica** (Gouan) All. subsp. **granatensis** (Pau) Malag in K.Tan: 222 (1980). SE Spain, C and W Pyrenees.
- P. grandiflora L.f.: 226 (1782) = Lachnaea grandiflora (L.f.) Baill. in J.B.P.Beyers: 45 (2001). South Africa: Western Cape.
- *P. gymnostachya* Meisn.: 397 (1840). = **Gnidia gymnostachya** (C.A.Mey.) Gilg in E.Phillips: 63 (1944). South Africa: North-West, Gauteng, Mpumalanga, Free State, KwaZulu-Natal and Eastern Cape, also in Lesotho.
- *P. hirsuta* Asso in Meisn. 555 (1857) = **Thymelaea tinctoria** (Pourr.) Endl. subsp. **tinctoria** in K.Tan: 223 (1980). Portugal (?), NE and E Spain and S France.
- *P. hirsuta* Brot. in K.Tan: 227 (1980) = **Thymelaea broteriana** Cout.: 145 (1909); K.Tan: 227 (1980). Endemic to N and C Portugal.
- *P. hirsuta* L.: 559 (1753) = **Thymelaea hirsuta** Endl. in K.Tan: 209 (1980). Circum-Mediterranean, Europe, SW Asia and N Africa.
 - P. imbricata Burm.f.: 12 (1768). True identity unknown.
 - P. imbricata Sennen: 69 (1924). True identity unknown. Spain.
- *P. incana* Pourr. ex Willk. & Lange: 300 (1862) = **Thymelaea velutina** (Pourr. ex Camb.) Endl. in K.Tan: 221(1980). Endemic to the Balearics.
- *P. inconspicua* Meisn.: 397 (1840) = **Gnidia inconspicua** Meisn. in Arnold & De Wet: 516 (1993) [= *Arthrosolen inconspicuus* Meisn. 560 (1857)]. South Africa: Western Cape.
- P. incurva Wendl. ex Bartl.: 404 (1840). True identity unknown.

- *P. involucrata* Spreng. ex Meisn.: 595 (1857) = **Gnidia capitata** L.f. in B.Peterson: 627 (1958) [= *Lasiosiphon linifolius* Decne. var. *glabrata* Meisn.: 595 (1857)]. South Africa: Limpopo [Northern Province], North-West, Gauteng, Mpumalanga, Free State, KwaZulu-Natal and Eastern Cape, also Botswana, Swaziland and Lesotho.
- *P. involucrata* Thunb.: 106 (1792) = **Pimelia linifolia** Sm. in B.Hansen & P.Wagner: 352 (1998). Australia.
- Passerina japonica Sieb. & Zucc.: 76 (1846) = Wikstroemia japonica Miq.: 184 (1863). Japan.
 - P. javanica Thunb.: 19 (1825b). True identity unknown. Java.
- *P. juniperifolia* Lapeyr.: 213 (1813) = **Thymelaea tinctoria** (Pourret) Endl. subsp. **nivalis** (Ramond) K.Tan [= *Thymelaea nivalis* (Ram.) Meisn.: 555 (1857)] K.Tan: 224 (1980). Endemic to C and W Pyrenees.
- P. kalifolia Pourr. ex Willk. & Lange: 301 (1862) = Thymelaea lanuginosa (Lam.) Ceballos & Vicioso in K.Tan: 211 (1980) [= Thymelaea canescens (Schousb.) Endl. in Meisn.: 556 (1857)]. S Spain, Morocco and Gibraltar.
- *P. laevigata* L.: 15 (1756) = **Gnidia oppositifolia** L. in Meisn.: 586 (1857). South Africa: Eastern Cape and Western Cape.
- *P. laniflora* C.H.Wright: 11 (1915) = **Lachnea laniflora** (C.H.Wright) Bond in J.B.P.Beyers: 81 (2001). South Africa: Western Cape.
- *P. lanuginosa* Pau: 67 (1922) = **Thymelaea lanuginosa** (Lam.) Ceballos & Vicioso in K.Tan: 211 (1980). Morocco, S Spain, Gibraltar.
- *P. lateriflora* Hort. ex Wikstr.: 347 (1818) = **Gnidia spicata** (L.f.) Gilg in E.Phillips: 63 (1944). South Africa: Western Cape.
- *P. laxa* L.f.: 226 (1782) = **Gnidia laxa** (L.f.) Gilg: 226 (1894a). South Africa: Eastern Cape and Western Cape.
- *P. lessertii* Wikstr.: 341 (1818) = **Stellera Lessertii** C.A.Mey. in Meisn. 14: 550 (1857). Iran.
- *P. linariaefolia* Pourr. ex Wikstr.: 333 (1818) = **Thymelaea pubescens** (L.) Meisn. subsp. **thesioides** (Lam.) K.Tan: 231 (1980). Spain.
- *P. linearifolia* Wikstr.: 343 (1818) = **Gnidia linoides** Wikstr. var. **major** Meisn.: 583 (1857).
- P. linearis Wendl. ex Bartl.: 404 (1840). Species non descriptae, Meisn.: 563 (1857).
- *P. linoides* Thunb.: 75 (1794) = **Gnidia linoides** Wikstr. in Meisn.: 582 (1857). South Africa: Western Cape.
- *P. longiflora* Thunb.: 189 (1800) = **Pimelea longifolia** Banks et Sol. in Meisn.: 516 (1857). New Zealand.
- Passerina longifolia Thunb.: 32 (1797) = Pimelia longifolia Banks et Sol. in Meisn.: 516 (1857). New Zealand.
- *Passerina matnak* J.F.Gmel.: 1597 (1792), sphalm. = *P. metkan* J.F.Gmel.: 634 (1791), sphalm.
- P. metnam Forssk.: 81 (1775) = **Thymelaea hirsuta** Endl. in K.Tan: 209 (1980). Circum-Mediterranean, Europe, SW Asia and N Africa.
- P. metnan Forssk, in Meisn.: 564 (1857) = **Thymelaea hirsuta** Endl. in K.Tan: 209 (1980). Circum-Mediterranean, Europe, SW Asia and N Africa.
- *P. microphylla* Coss. & Durand: 744 (1856) 744 = **Thymelaea microphylla** Meisn.: 556 (1857); K.Tan: 214 (1980). N Africa.
- *P. nervosa* Thunb.: 75 (1794) = **Lachnaea nervosa** (Thunb.) Meisn. in J.B.P.Beyers: 69 (2001). South Africa: Western Cape.

P. nervosa Wikstr.: 328 (1818) = **Lachnaea striata** (Poir.) Meisn.: 577 (1857); J.B.P.Beyers: 77 (2001). South Africa: Western Cape.

P. nitida (Vahl) Desf.: 331, t. 94 (1798) = **Thymelaea argentata** (Lam.) Pau in K.Tan: 212 (1980) [= *Thymelaea nitida* Endl. in Meisn.: 554 (1857)]. Spain and N Africa.

P. nivalis Ram.: 131, t. 9 (1800) = **Thymelaea tinctoria** (Pourret) Endl. subsp. **nivalis** (Ramond) K.Tan: 224 (1980). Endemic to C and W Pyrenees.

P. orientalis Willd.: 431 (1799) = **Thymelaea tinctoria** (Gouan) All. subsp. **dioica** [= *Thymelaea orientalis* Meisn.: 557 (1857)] in K.Tan: 222 (1980). W Pyrenees to SW Alps.

P. passerina Huth.: 135 (1898) = **Thymelaea passerina** (L.) Coss. & Germ. in K.Tan: 236 (1980) [= *Thymelaea arvensis* Lam. in Meisn.: 551 (1857) (= *Stellera passerina* L.)]. Widespread: C and S Europe, SW Asia, C Russia.

P. pentandra Thunb.: 76 (1794) = **Lonchostoma obtusiflorum** Wikstr. in Meisn.: 564 (1857).

P. persica Boiss.: 85 (1846) = **Stellera lessertii** C.A.Mey. in Meisn.: 550 (1857). Iran.

P. pilosa G.Forst. in Meisn.: 564 (1857); B.Hansen & P.Wagner: 352 (1988) = **Pimelea tomentosa** (J.R.Forst. & G.Forst.) Druce in B.Hansen & P.Wagner: 352 (1998). New Zealand.

Passerina pilosa L.f.: 226 (1782) = **Pimelea tomentosa** (J.R. & G.Forst.) Druce in B.Hansen & P.Wagner: 352 (1998) (= Pimelea virgata Vahl). New Zealand.

Passerina planifolia Burm.f.: 12 (1768) = Lachnaea alpina Meisn.: 578 (1857); J.B.P.Beyers: 59 (2001). South Africa: Western Cape.

P. polycephala E.Mey ex Meisn.: 390 (1840) = Gnidia polycephala (C.A.Mey.) Gilg: 227 (1894a). Namibia, Botswana and South Africa: North-West, Gauteng, Mpumalanga, Free State, Eastern Cape and Northern Cape.

P. polygalaefolia Lapeyr.: 214 (1813) = **Thymelaea hirsuta** Endl. in K.Tan: 209 (1980). Circum-Mediterranean, Europe, SW Asia and N Africa.

P. prostrata G.Forst.; 28 (1786) = **Pimelea prostrata** (J.R.Forst. & G.Forst.) Willd. in B.Hansen & P.Wagner: 352 (1998). New Zealand.

Passerina prostrata L.f.: 227 (1782) = Pimelea prostrata (J.R.Forst. & G.Forst.) Willd. in B.Hansen & P.Wagner: 352 (1998). New Zealand.

Passerina pubescens Guss.: 451 (1843) = **Thymelaea** mesopotamica (Jeffrey) Peterson in K.Tan: 239 (1980) [= *Thymelaea arvensis* Lam. var. pubescens (Guss.) Meisn.: 552 (1857)]. SE Anatolia, Iraq, W Iran, Kuwait.

P. pubescens (L.) Loscos var. *virgata* Pau: 33 (1925) = **Thymelaea pubescens** (L.) Meisn. subsp. **thesioides** (Lam.) K.Tan: 231 (1980). Spain.

P. pubescens (L.) Wikstr.: 332 (1818) = **Thymelaea pubescens** (L.) Meisn. subsp. **pubescens** in K.Tan: 231 (1980). Pyrenees, S and E Spain.

P. purpurea Wikstr.: 323 (1818) = **Lachnaea eriocephala** L. in J.B.P.Beyers: 99 (2001). South Africa: Western Cape.

P. racemosa Wikstr.: 320 (1818) = **Stellera altaica** Thieb. in Meisn.: 549 (1857). Subalpine region, the Caucasus.

P. rosmarinifoliae fide Meisn.: 400 (1840) = Phylica sp., Herb. Lamarck (P, microfiche 574).

Passerina ruizii (Loscos.) Font-Quer in K.Tan: 225 (1980) = **Thymelaea ruizii** [Loscos] Casav. in K.Tan: 225 (1980). High ranges, Sierra de Cabrera in NE Portugal to C Pyrenees.

P. salina Munby = **Thymelaea passerina** (L.) Coss. & Germ. in K.Tan: 236 (1980); seen from (C), specimen without collector and number.

P. salsa Hunley, Debeaux s.n. (PR), cf. P. annua (Salisb.) Wikstr. var. salsa Munby in K.Tan: 238 (1980) = **Thymelaea salsa** Murb. in K.Tan: 238 (1980). S Spain, Morocco and Algeria.

P. salsolaefolia Poir.: 41 (1804) = **Phylica sp.** in Meisn.: 400 (1840); Herb. Lamarck (P. microfiche 574).

Passerina sanamunda (All.) Bub.: 135 (1897) = **Thyme-laea sanamunda** All. in Meisn.: 553 (1857); K.Tan: 233 (1980). Spain, S France and Italian Riviera.

P. segobriensis Pau: 25 (1887) = **Thymelaea argentata** (Lam.) Pau in K.Tan: 212 (1980) [= *Thymelaea nitida* Endl. in Meisn.: 554 (1857)]. Spain and N Africa.

P. sericea L.: 15 (1755) = **Gnidia sericea** L. in Meisn.: 583 (1857). South Africa: Eastern Cape and Western Cape.

P. setosa Thunb.: 75 (1794) = **Gnidia setosa** Wikstr. in Meisn.: 590 (1857). Eastern Cape.

P. spicata L.f.: 226 (1782) = **Gnidia spicata** (L.f.) Gilg in E.Phillips: 63 (1944). South Africa: Western Cape.

P. stachyoides Schrenk in Meisn.: 550 (1857) = Stellera stachyoides Schrenk in Meisn.: 550 (1857), Siberia.

P. stellera Ram. ex Lam. & DC.: 361 (1805) = Thymelaea passerina (L.) Coss. & Germ. in K.Tan: 236 (1980) [= Thymelaea arvensis Lam. in Meisn.: 551 (1857)]. Widespread: C and S Europe, SW Asia, C Russia.

P. stelleri Wikstr.: 321 (1818) = **Stellera chamaejasme** L. in Meisn.: 549 (1857). Siberia.

P. striata Poir.: 44, t. 291, f. 2 (1804) = **Lachnaea striata** (Poir) Meisn.: 415–416 (1840); J.B.P.Beyers: 77 (2001). South Africa: Western Cape.

P. stricta Thunb.: 75 (1794) = **Gnidia wikstroemiana** Meisn.: 582 (1857). South Africa: Northern Cape and Free State.

P. subspicata Meisn.: 395 (1840) = **Wikstroemia subspicata** Meisn. 14: 547 (1857).

P. tartonraira Schrad.: 89 (1810) = **Thymelaea Tartonraira** All. in Meisn.: 556 (1857); K.Tan: 215 (1980). Circum-Mediterranean.

P. tenuiflora Willd.: 426 (1809) = **Gnidia laxa** (L.f.) Gilg: 226 (1894a). South Africa: Eastern Cape and Western Cape.

P. tetragona Steud.: 274 (1841) = **Struthiola dodecandra** (L.) Druce in Levyns: 599 (1950). South Africa: Eastern Cape and Western Cape.

P. thesioides Wikstr.: 333 (1818) = **Thymelaea pubescens** (L.) Meisn. subsp. **thesioides** (Lam.) K.Tan [= *Thymelaea thesioides* (Lam.) Endl. in Meisn.: 553 (1857)] in K.Tan : 231 (1980). Spain.

P. thomasii Duby: 406 (1828) = **Thymelaea tartonraira** All. subsp. **thomasii** (Duby) Briq. [= *Thymelaea tartonraira* All. var. calvescens Gren. & Godr. in Meisn.: 556 (1857)] in K.Tan: 220 (1980). Circum-Mediterranean.

P. thumbergii Wikstr.: 343 (1818) = **Gnidia sericea** L. in Meisn.: 583 (1857). South Africa: Eastern Cape and Western Cape.

P. thymelaea (Lam.) DC. in Lam. & DC.: 366 (1815) = **Thymelaea sanamunda** All. in Meisn.: 553 (1857); K.Tan: 233 (1980). Spain, S France and Italian Riviera.

P. tinctoria Pourr. in Meisn.: 565 (1857) = **Thymelaea tinctoria** (Pourr.) Endl. subsp. **tinctoria** in K.Tan: 223 (1980). Portugal (?), NE and E Spain and S France.

P. tinctoria var. angustifolia Boiss.: 556 (1842) = **Thymelaea** calycina Meisn.: 555 (1857); K.Tan: 226 (1980). C Pyrenees, rare.

P. tingitana Salzm. ex Meisn.: 554 (1857) = **Thymelaea villosa** (L.) Endl. in K.Tan: 229 (1980). S Portugal, Spain, Gibraltar, Morocco.

P. tomentosa Wikstr.: 332 (1818) = **Thymelaea tartonraira** All. in Meisn.: 556 (1857); K.Tan: 215 (1980). Circum-Mediterranean.

P. uniflora Drège ex Meisn.: 574 (1857) = **Gnidia penicillata** Licht. ex Meisn. in B.Peterson: 476 (1959), [=*Cryptadenia ciliata* (Thunb.) Meisn. in J.B.P.Beyers: 45 (2001), excluded species].

P. uniflora L.: 560 (1753) = **Lachnaea uniflora** (L.) Crantz in J.B.P.Beyers: 43 (2001). South Africa: Western Cape.

P. uniflora L. var. *alba* P.J.Bergius: 129 (1767) = **Lachnaea grandiflora** (L.f.) Baill. in J.B.P.Beyers: 45 (2001). South Africa: Western Cape.

P. uniflora L. var. *angustifolia* Burm.f.: 12 (1768). = **Lachnaea uniflora** (L.) Crantz in J.B.P.Beyers: 43 (2001). South Africa: Western Cape.

P. uniflora L. var. latifolia Burm.f.: 12 (1768). = Lachnaea grandiflora (L.f.) Baill. in J.B.P.Beyers: 45 (2001). South Africa: Western Cape.

P. uniflora L. var. *purpurea* P.J.Bergius: 128 (1767). = **Lachnaea uniflora** (L.) Crantz in J.B.P.Beyers: 43 (2001). South Africa: Western Cape.

P. velutina Boiss.: 81 (1838) = **Thymelaea pubescens** (L.) Meisn. subsp. **elliptica** (Boiss.) K.Tan: 231 (1980). Endemic to S Spain.

P. velutina [Pourr.] Cambess.: 183 (1827) = **Thymelaea velutina** (Pourr. ex Camb.) Endl. in K.Tan: 221 (1980). Endemic to the Balearics.

P. vesiculosa Fisch. & C.A.Mey.: 170 (1839) = **Diarthron** vesciculosum C.A. Mey. in Meisn.: 558 (1857). Iran.

P. villosa Thunb.: 106 (1792) = **Pimelea arenaria** Cunn. in Meisn.: 517 (1857). New Zealand.

Passerina villosa Wikstr.: 332 (1318) = **Thymelaea villosa** (L.) Endl. in Meisn.: 554 (1857); K.Tan: 229 (1980). S Portugal and Spain, Gibraltar, Morocco.

P. virescens Coss. & Dur. ex Meisn.: 554 (1857) = **Thymelaea virescens** Meisn.: 554 (1857); K.Tan: 212 (1980). NW Africa.

P. virgata Desf.: 331, t. 95 (1798) = **Thymelaea virgata** (Desf.) Endl. in K.Tan: 228 (1980); subsp. **virgata** from NW Africa and S Spain & subsp. **broussonetii** from NW Africa.

SPECIMENS EXAMINED

Abel s.n. (19) NBG. Abrahams s.n. & A7759 (7) GRA, MO, PRE. Acocks 184, 583, 1001, 5756 (1) S; 690 (6) S; 1005 (4) S; 1006 (7) S; 1007, 1549 (15) S; 1067, 1519 (14) S; 2482 (16) S; 10660, 12125, 12175 (4) PRE; 10967 (7) PRE; 16000, 18634, 19000, 21252, 22862, 24427 (15) K, PRE; 19066 (6) K, PRE; 19810 (14) K, NBG, PRE; 20170 (15) M, PRE; 20714, 21309, 23379, 23848, 24257 (14) PRE; 21424 (20) K, M, PRE; 21455 (7) M, PRE; 22365 (20) PRE; 22528, 22606, 22615 (1) PRE; 22608 (12) K, PRE; 22784 (6) PRE; 24212 (1) K, NBG PRE; 24213 (12) K, NBG, PRE. Adamson 1515 (10) PRE; 5124 (15) PRE. Alexander-Prior s.n. (1) PRE; s.n. (14) PRE. Allardice 1726 (15) NBG, Anderson 76 (7) PRE. Andersson s.n. (14) S. Andraea 385, 385b (6) NBG, PRE; 581 (1) NBG, PRE; 597 (14) PRE; 979, 982 (15) NBG, PRE; 1018, 1030 (15) PRE; 1222, 1288, 1288A (10) PRE; 1227 (14) NBG, Archibald 677 (4) GRA; 3833 (20) GRA; 4552/52 (7) GRA, PRE; 4558, 5259 (20) K, PRE; 5583, 5727, 6053 (14) PRE.

Balkwill 456 (15) K PRE. Balsinhas & Kersberg 2114 (4) PRE. Barber 745 (4) GRA. Barbosa 9447 (4) K. Barker 349 (14) PRE; 2726 (6) NBG; 5565 (20) NBG; 5925 (15) C. NBG; 6036 (19) NBG, S: 10580 (15) NBG. Barnard 526, 699 (20) PRE; 660 (14) PRE; s.n. (10) NBG; s.n. (14) NBG. Barnes 88 (14) GRA. Bayer 786 (4) PRE; 1307 (7) MO. Bayliss 521 (15) K. MO, PRE; 546 (20) K. M. MO, NBG; 650 (7) K. MO, M, PRE; 1349, 1362, 1432 (4) PRE; 1684 (15) PRE; 2468 (14) B. MO, NBG; 5023 (14) M, MO; 5035 (14) B. MO; 5164 (20) MO; 5238, 5289 (14) MO; 6017 (7) PRE; 6060, 6093 (15) PRE; 6117 (14) PRE;

6856, 8850 (20) M, MO; 6861 (20) K, MO, S; 7731, 8908 (15) M, MO; BRI B32 (14) GRA, MO, PRE; BRI B295 (19) PRE; BRI B1088 (20) PRE; BRI B1105 (14) PRE. Bean 1385 (15) BOL. MO, NBG. Bean & Viviers 1508 (14) BOL. Behemiae s.n. (14) PR. Bengis 344 (15) PRE. Beverly 72 (4) PRE. Blom 275 (4) PRE. Boemert, Herb. Reg. Monacense s.n. (14) M. Bohnen 94.04 (7) PRE; 4012 (12) C, PRE; 4065 (12) NBG, PRE; 4911, 8147 (7) NBG, PRE. Bolton s.n. (20) DUB. Bolus 170 (14) BOL, K; 687 (5) BM, BOL, HAL, K, NBG, P, PRE, UPS, W; 1905 (20) BOL; 2924 (14) K; 2926 (1) K; 4498 (6) BM, BOL, K; 11630 (15) BM, BOL; 17197 (1) BOL; s.n. (1) BOL, PRE; s.n. (14) BOL. Bond 1820 (15) NBG. Borges 112 (4) M, PRE. Boshoff 156 (15) NBG. Bot. Mus. Univ. Wien s.n. (14) WU. Botha 372 (7) PRE; 2610 (4) PRE; 2618 (14) PRE; 5656 (20) GRA. Botha & Coetzee 1605 (7) PRE. Boucher 468, 469 (1) NBG, PRE: 470 (7) K: 761, 1955, 2012. 2014, 2449, 3243, 3301 (14) PRE; 856 (6) K, NBG, PRE; 1559a (15) NBG, PRE; 1603 (1) PRE; 1677 (7) PRE; 1690 (7) NBG, PRE; 1691 (6) NBG; 1862a, 1862b (4) NBG; 2008 (15) PRE; 2219 (1) NBG, PRE; 2224 (12) NBG; 2244, 2439 (15) PRE; 2951 (1) NBG, PRE; 3363 (7) NBG; 3973 (6) PRE. Boucher & Shepherd 4828 (14) PRE. Bower 602 (7) PRE. Bowie 1 (6) BM; 3 (20) BM; 5 (19) BM. Bowker s.n. (7) K. Brain 6950 (4) MO, PRE. Bredenkamp 889-891, 893-895 (4) PRE; 896 (14) PRE; 897-899 (7) PRE; 900 (20) PRE; 901-903 (14) PRE; 904 (15) PRE; 905, 906 (20) PRE; 907 (14) PRE; 908, 909 (11) PRE; 911, 913 (7) PRE; 912 (14) PRE; 914 (20) PRE; 915 (19) PRE; 916 (14) PRE; 917 (19) PRE; 918 (14) PRE; 919 (15) PRE; 920, 921 (19) PRE; 922 (15) PRE; 923 (12) PRE; 924-927 (14) PRE; 928-930 (15) PRE; 931 (14) PRE; 932 (12) PRE; 933, 934 (12) PRE; 935 (7) PRE; 936 (14) PRE; 937, 938 (1) PRE; 939 (12) PRE; 940 (1) PRE; 943-945 (14) PRE; 946, 947 (12) PRE; 948 (7) PRE; 949, 950 (1) PRE; 951, 953 (14) PRE: 952, 954 (1) PRE: 955 (14) PRE: 956 (6) PRE: 957-959 (14) PRE; 960, 961 (1) PRE; 962 (6) PRE; 963-965 (14) PRE; 966-968, 970, 971 (15) PRE; 974 (14) PRE; 979, 980 (14) PRE; 1013, 1014 (7) PRE; 1018-1021 (13) PRE; 1022-1026 (4) PRE; 1035 (16) PRE; 1545, 1546 (5) PRE; 1549, 1554 (15) PRE. Brehm.: Herb. Reg. Monacense s.n. (14) M. Breijer TRV 16577 (14) PRE; TRV 16898 (14) PRE. Bremekamp & Schweickerdt 417 (4) PRE. Bremer 299 (14) BOL. Brink 232 (20) GRA, K, PRE; 924 (15) GRA. Britten 130 (19) PRE; 1245 (15) GRA; 1633 (15) GRA, PRE; 2239 (14) PRE; 2562 (14) GRA; 2828 (14) PRE; 5008 (14) GRA, PRE; 5014 (7) GRA; 5489, 5522 (14) PRE; 5523 (15) PRE; 5812 (19) GRA, PRE. Britton 12 (1) NBG. Brooker-Leslie s.n. (15) GRA. Brown 490 (14) PRE; 28858 (19) PRE. Bryce s.n. (4) K. Buchenau s.n. Buff 760610-2/1 (7) WU. Buitendag 104 (4) NBG, PRE. Burchell 473, 3835, 6163, 6721 (14) K; 4049, 7463 (7) K; 6109 (19) K, PRE; 7129 (10) K; 7761 (5) K, M; 8389 (1) K. Burgers 1015, 2186 (1) PRE; 1188, 2270 (14) PRE; 1464 (20) PRE; 2259 (12) PRE; 2268 (1) NBG, PRE; 2924 (7) NBG. Burke 45 (14) K, PRE. Burrows 2464 (19) PRE; 3066, 3956 (14) GRA; 3395 (7) GRA; 4118, 4682 (15) GRA. Burtt-Davy 2410 (7) K.

Campbell 13541 (19) NBG. Capener CF/2 (19) PRE. Cattell & Cattell 167 (15) NBG. Catterrell 41 (4) PRE. Chase 592 (4) BM, PRE. Codd 3602 (7) PRE, K; 8772 (4) PRE; Codd & De Winter 3237 (4) PRE; Codd & Dyer 4473 (4) PRE. Cole s.n. (14) TCD. Collector unknown s.n. (14) S; (6) BOL, P, W; (6) S; 170 (14) S; (19) S. Comins 1930 (7) PRE. Compton 2241, 21048, 21236, s.n. (4) NBG; 10633 (6) NBG; 11594 (1) NBG, S; 9172, 13486, 13600, 15996, 22941, 23625, 24363 (1) NBG; 18159 (1) MO, NBG; 18278 (15) M, NBG; 18380, 18449 (15) NBG; 19930 (1) C, NBG; 20274 (19) NBG; 21673 (14) C, MO; 22122 (1) BOL, NBG; 22218 (10) NBG, S; 22289 (6) NBG, S; 22870 (10) C, NBG; 2884 (15) BOL, NBG; 3005, 3166, 5915 (14) BOL, K; 3835 (15) BOL, NBG; 5725, 7427, 7438, 7498 (15) NBG; 7422 (19) NBG; 7584 (19) C; 9066, 14743 (12) NBG; s.n. (7) S. Cooper 15 (7) GRA; 625 (4) BM, BOL, K, TCD; 702 (4) BM, BOL, K, M, PRE, TCD; 842 (4) BOL, K; 2301 (7) K; 2302 (4) K. Corneliussen 1875 (6) C. Cowling 51 (14) GRA; 796 (11) GRA; 926 (15) GRA; 1223 (20) GRA; 1419 (19) GRA; 3436 (1) NBG. Crawford 384 (7) PRE. Cross 58 (15) MO, NBG, Cruden 37 (15) NBG, Cummings s.n. (14) GRA. Curator Bloemfontein Museum 4 (4) PRE. Curator Pretoria Botanical Garden P42 (10) PRE; s.n. (4) PRE, Curson & Irvine 91 (4) PRE

Dacombe s.n. (14) GRA. Dahlgren & Peterson 168 (14) M; 1661 (4) B, K. Dahlstrand 148 (7) MO; 1285 (19) C; 1490 (15) C, MO, NBG, PRE: 1905 (19) MO, PRE: 2836, 2849, 2850, (20) C, GRA, MO, NBG, PRE: 2949 (7) NBG, PRE: 3251 (20) C, GRA, MO, NBG, PRE. Davidson 24904 (14) PRE. Davidson & Mogg 32859, 32881 (4) UPS; 32909 (4) PRE. Davies 33 (14) PRE, 48954 (4) PRE. De Beer TRV 16536 (14) PRE. De Kock 134 (14) PRE. De Kruif 1158 (4) PRE. Deall 34 (4) PRE. Dela Bat s.n. (1) NBG, Devenish 712 (4) BM, K, M, NBG, PRE. Dieterlen 49 (4) K, MO, NBG, PRE. S; 1247 (4) NBG, PRE. District Forest Officer 97 (7) GRA. Dobay 45 (15) NBG. Dold 840 (4)

GRA; 1063 (20) GRA; 1099 (14) GRA; 1617, 2053 (4) GRA; 2299 (15) GRA. Doubell 27 (15) GRA. Downing 401 (6) NBG, PRE. Drège 85 (14) UPS; 285 (14) GRA; 2570 (10) K, PRE; 3006 (20) GRA, PRE; s.n. (1) S; s.n. (7) K, MO; s.n. (10) BM, NBG, S; s.n. (14) MO, P; s.n. (14) S, W; s.n. (19) K; TRV10800 (14) PRE. Du Toit 155 (15) PRE. Dunne s.n. (4) BM. Duthie 533 (7) NBG; 1517 (16) BOL. Dux de Wirte s.n. (14) M. Dyer 179, 180 (14) PRE; 596 (15) K, PRE; 597 (14) GRA, K, PRE; 752–754 (4) GRA, K, PRE; 960, 961 (15) K, PRE; 962, 967, 968 (14) GRA, K, PRE; 963–966 (14) GRA, PRE; 970 (14) PRE.

Ebersohn 136 (14) NBG. Ecklon 1.11 (14) B, S; 508 (14) BOL, HAL. M, PR, PRE, S, W; 590 (14) S; 598 (20) S; s.n. (6) C; s.n. (12) S; s.n. (14) C. Ecklon & Zeyher 37 (14) MO, S; 100/12 (14) S; 38 (19) MO; 39 (6) BOL, G, MO, P, W; 40 (70.10) (14) B, BREM, C, MO, S, W; 41 (14) MO; 98 (15) BOL; 3781 (11) G, HAL, P, PRE, S, W, WU; 3782 (14) NBG, W; 7381 (11) BREM, K, P, PRE, WU; s.n. (1) PRE; s.n. (7) BREM, NBG, S; s.n. (10) C, UPS; s.n. (11) C, S, UPS; s.n. (14) BREM, GRA, MO, WU; s.n. (15) WU. Edwards 973, 974 (13) BOL. PRE: 1991 (4) PRE; 2239, 2276, 4058, 4196 (4) K, PRE; 4187 (4) K, MO, PRE; 17260 (20) BM. Eicker 1 (7) PRE. Elan-Puttick 146 (4) PRE. Erlangh 1017 (14) M. Esterhuysen 781 (14) MO; 961, 23319, 27139a (14) BOL; 1855 (1) BOL; 1998, 8978, 23446, s.n. (15) BOL; 3016 (12) BOL; 6634 (19) BOL; 6678 (11) K, NBG; 6962, 10733 (11) BOL; 11148 (10) BOL; 12189 (9) BOL; 16295 (14) PRE; 23322 (12) MO; 26714, 35616 (5) BOL, K; 29153 (4) BOL; 29962 (16) BOL, MO; 33533a (5) BOL M; 34537, 36404 (5) BOL, K, S; 35415 (5) BOL; 35504 (1) BOL, C, M, MO; 35507 (1) S. Euckermann 7889 (7) PRE. Euston-Brown 41 (11) BOL. Eyles 8527 (4) K, S; Eyles Herbarium 7945 (4) BM.

Fellingham 169 (15) NBG, PRE; 202 (14) PRE; 773 (1) C, PRE. Ferreira 5 (4) PRE. Flanagan 418 (7) NBG, PRE; 1478 (14) BOL; 1635, 1892 (4) K, NBG, PRE, S. Fokkens 5 (7) PRE. Forrester & Gooyer 203 (4) PRE. Fouche s.n. (7) PRE. Fourcade 74 (19) BOL, K; 741 (15) BOL, K, GRA; 952 (14) BOL, GRA; 1478 (14) BOL, GRA, K, PRE; 1484 (14) BOL, K; 1646 (20) BOL; 1708 (11) BOL, K, NBG, PRE; 1806 (7) K, PRE; 1806 (7) NBG; 3043 (11) MO, PRE; 4417 (20) BOL; 4624, 4625 (15) BOL; 5747 (20) BOL; 5747 (20) NBG; 5940 (20) BOL, PRE. Frankish 253 (7) MO. Franks s.n. (4) PRE. Fries 2248, 2496 (4) K, MO. Fries, Nordlindh & Weimarck 3077 (4) M.

Gafney 11 (7) MO, PRE. Galpin 178 (15) PRE; 251 (14) PRE; 2028 (4) K, PRE; 3363 (7) PRE; 4491 (12) K, PRE; 4492 (20) GRA, K, PRE; 6825 (4) K; 6825 (4) PRE; 8279 (14) PRE; 9353 (7) K, PRE; 10146 (4) PRE; 14018, 14573, 14578, s.n. (4) BOL, PRE, S. Garside 491 (1) K; 502 (6) K; 1577 (1) K. Gentry 18995 (14) PRE. Germishuizen 4024 (15) PRE; 4077, 4097 (14) PRE. Gerrard 95 (7) K. Gerstner 105 (14) PRE; Gerstner 119, s.n. (4) PRE. Gibbs Russell 4094 (7) PRE. Giess 1293 (6) M; 9014, 9429, 13136 (4) M, PRE. Giffen s.n. (4) GRA. Gilfillan 7 (7) GRA. Gill 240 (4) BOL. Gillett 814, 1012 (1) NBG; 1207 (7) PRE; 1986 (15) NBG; 3457 (6) NBG; 4536 (19) BOL, PRE; 4536 (19) K. Gilliland 881 (4) BM; 904, 905 (4) BM, K; 1183, 2025 (4) BM. Glen 1089 (6) PRE; 1568 (15) C, PRE. Glen & Glen 3911 (4) PRE. Goldblatt 2599 (1) MO, PRE, S; 4144 (12) MO, NBG, PRE; 4380 (10) MO, NBG. Goldblatt & Manning 9589 (15) NBG. Goldsmith 15/73 (4) K. MO. PRE. Goossens 375 (4) PRE). Gormley & Barber 23 (4) PRE. Greuter 21500 (1) PRE; 21517 (14) PRE; 21845 (14) B. PRE; 22180 (15) PRE. Grey 4 (1) C. Grobbelaar 63 (19) PRE. Grobler 454 (14) NBG, PRE. Grondahl s.n. (14) S. Guthrie 17413 (7) BOL; 17414 (6) BOL

Hafström s.n. (14) S. Hardy 6894 (4) PRE. Harvey 691 (6) BM. Hebblethwaite s.n. (4) GRA. Hedberg & Hedberg 82060 (4) UPS. Hemm 1 (4) PRE. Hendricks 13 (7) GRA. Henkel s.n. (4) K. Henrici 3701 (12) BOL; Henrici s.n. (12) NBG. Hepburn 85 (4) GRA. Herb. Banks & Swartz s.n. (1) S. Herb. Banks & Wikstr. (1) C. Herb. Bot. Hauniense s.n. (14) C. Herb. J. Peterstein acc. no. 16/1946 (14) PR. Herb. Poeppig s.n. (14) M. PRE. Herb. Praga Karlin, Herb. scholae med. (14) PR. Herb. Reg. Monacense s.n. (14) M. Herb. Schmidel s.n. (14) M. Herb. Scholae Lincopensis s.n. (14) S. Herb. Schreberianum s.n. (14) M. Herb. Sieber 89 (14) S. Herb. Swartzii s.n. (14) S. Herb. Thunberg 9578 (7) UPS; 9579, 9596D (1) UPS; s.n. (14) UPS; s.n. (19) UPS. Herbst 5269 (4) PRE. Heydoorn 10 (4) PRE. Hiendlmayr s.n. (6) M. Hilger 22 (4) M. PRE; 83/60 (14) M. Hilliard & Burtt 7139 (4) MO, S; 7220 (4) PRE; 11726, 13511 (4) K, S; 12213 (4) PRE, S; 13970, 18071 (4) PRE; 14654 (15) PRE; 16921 (4) M, PRE, S; 17703, 18443 (4) PRE, S; 18570 (13) K, M, P, PRE, S. Hilliard 4081 (14) K, PRE; 5191 (4) MO, PRE. Hilner 184 (7) PRE. Hoekstra 73 (15) NBG. Hoener 1635, 1846 (4) MO, PRE, S. Hoffenthal 3464 (4) K. Holland 3699 (7) BOL. Homan s.n. (4) M. Hoole 10 (20) GRA, PRE; J1 (7) GRA. Hopkins B1580, 17165 (4) K. PRE; s.n. (4) K. Hubbard 224 (1) NBG. Hugo 1241 (14) PRE: 1405 (19) K, NBG, PRE; 1453 (15) K, PRE; 1912 (7) C, M, PRE; 1992 (7) NBG.

PRE; 2079 (14) NBG, PRE. Humbert 11018, 15801 (4) NBG; 9908 (19) PRE. Huntley 125 (4) PRE. Hutchinson 143 (1) BOL, K, PRE; 149 (1) BOL, GRA, K, PRE; 663 (6) K, PRE; 1101 (15) BM, K, PRE; 1104 (15) K, PRE; 1624 (4) BM, K, PRE; 1748 (7) K, PRE. Hutchinson, Forbes & Verdoorn 75 (4) PRE. Hutton 1603 (14) K; s.n. (7) TCD.

Ihlenfeldt 1652 (14) PRE

Jacobsen 1340 (4) PRE: 1381 (7) PRE: 3468 (4) PRE. Jacobsz 1379, 1482 (4) PRE; 2603 (4) NBG, PRE; 313 (4) PRE; 664 (4) PRE. Jacot-Guillarmod 230, 298, 765, 780, 1546, 1792 (4) PRE; 3225 (4) K, PRE; 3824 (4) GRA; 7464 (15) PRE; 7610 (14) GRA; 7854 (4) GRA, PRE; 9842, 9886 (4) GRA, PRE; 10003 (14) GRA, PRE; 20776 (1) GRA, S.n. (15) GRA. Jacot-Guillarmod & Brink 29 (14) GRA; 41 (7) GRA, PRE, Jangle 156 (16) PRE. Johnson 102 (20) M, NBG; 1069 (7) GRA, K, PRE, Johnstone 543 (4) MO, PRE, S. Jordaan 97 (4) PRE; 655 (1) PRE. Joubert s.n. (14) S. Jules Verreaux s.n. (14) TCD. Junod TRV17326 (4) PRE.

Kapp 1 (14) PRE. Keet 873 (7) NBG; 1152 (19) PRE; s.n. (4) NBG. Kemp 1184 (4) MO, PRE. Kemsley 141 (7) GRA, NBG. Kerfoot 8102 (4) PRE. Kers 3240 (4) S). Killick 1071, 1585 (4) PRE; 1973, 2289 (13) PRE; 4225 (4) MO, PRE; 4485 (4) PRE. Killick & Strey 2386 (4) M. PRE. King 1 (14) BM, M, MO, PRE. Kluge 797, 1995 (4) PRE; 2044 (4) PRE. Knaf s.n. (14) PR. Krause s.n. (6) NBG; s.n. (1) M. Krynauw 789 (4) PRE. Kuntze s.n. (7) K; s.n. (14) K.

Laidler 372 (15) NBG, PRE. Lam & Meeuse 4272 (1) S; 4660 (19) MO. Lambinon & Reekmans 82/157 (4) PRE. Lansdell s.n. (14) PRE. Lanyokwe 97, (14) GRA. Laubert s.n. (6) S. Laubner s.n. (7) K. Lavranos 3700 (15) PRE; 11652 (14) PRE. Le Jolis s.n. (14) S. Le Munch 387 (4) K. Leighton 1855 (1) PRE; 21113 (12) BOL. Leitz 182b, s.n. (14) M. Letty 142 (14) PRE: 269 (4) PRE. Levyns 1515, 2371 (10) BOL: 2303, 2349, 2414, 2813, 5538, 6314, 6514, 6626, 6627, 8029, 9140, 9145, 9741, 9743, 15495 (15) BOL; 2325, 2550, 3763, 4443, 9603, 9644, 10833, 10873 (14) BOL; 3100 (6) BOL; 3101, 10291, 10832, 10867 (7) BOL; 5040, 7842 (19) BOL; 8236 (13) BOL; 8274, 9409, 9553 (4) BOL; 9511, 9674, 9726 (12) BOL; 9725 (1) BOL. Lewis 71 (1) NBG; 1795, 1796 (15) NBG; 1797 (15) NBG, PRE. Liebenberg 6574 (20) PRE; 7299 (4) K, NBG, PRE. Lindeberg s.n. (1) S. Linder 3980 (4) BOL, K, PRE. Lindley s.n. (14) S. Linnaeus 504.5 (6) LINN. Long 386 (15) K, PRE; 790 (14) GRA, PRE; 798 (7) K, PRE; 809 (20) PRE; 1061 (20) GRA, PRE. Louw 2373 (4) NBG. Louwrens A7763 (14) GRA. Lubke 274 (4) PRE, M; 1782, 2312 (7) GRA; 1861, s.n. (14) GRA. Lutjeharms 6818 (4) PRE. Lynes 1737 (20) BM; 1927 (14) BM.

MacOwen 103 (14) GRA, K, TCD; 103 (15) GRA; 103 (20) GRA, K, S; 3404 (6) K. Maguire 1127 (14) M; 1210 (15) NBG. Manson 147 (19) PRE. Marloth 1585, 2756, 6189, 6218, 10693, 10912 (14) PRE; 4273, 6862, 10964, 11283, 12724, 12785, 14128 (15) PRE; 5648 (6) B, PRE; 6244, 13044 (19) PRE; 9695 (10) PRE; 10817 (10) NBG, PRE; 11873 (4) PRE; 13012 (7) PRE; s.n. (6) PRE. Marsh 541 (1) K, PRE; 572, 1361 (14) PRE; 594 (19) K, PRE; 1327 (7) NBG, PRE, Marshall 130, 131 (15) NBG; 244 (15) NBG, PRE. Martin s.n. (6) GRA; s.n. (7) GRA; s.n. (19) GRA. Masson s.n. (1) BM; s.n. (20) BM. Matthews 284 (15) PRE; 1153 (15) NBG. Matthews & Van Rensburg 1036 (4) PRE). Mauve 5246 (4) PRE. Mauve & Hugo 167 (14) MO. PRE. McDonald 819 (14) NBG. PRE; 2125, (10) NBG. McDonald & Morley 1005 (15) BM, NBG, PRE. TCD, McKinnon s.n. (14) NBG, McKitterick 12 (7) GRA, McMurtry 335 (15) PRE. Meebold 9965 (4) PRE; 15151, 15153, 15159, 15160 (14) M; 15154 (6) M; 15158 (4) M; 15755 (7) M. Meinkauff s.n. (14) M; s.n. (19) M. Mellersh 617 (4) TCD. Mendes 866 (4) BM; 3803 (4) K, M. Merxmüller 591 (4) M. Meyer 106, 1061, 9429 (4) M. Michell 24 (14) PRE; 71, 326 (15) PRE; 333 (10) PRE. Miller 2701 (7) PRE; 3815, 3847, 4644 (4) K, PRE. Milton 2 (16) BOL. Mitchell, Panimenter & Spencer B4-11 (7) PRE, Mogg 3347, 7083 (4) PRE; 13220 (7) K, PRE; 17383 (4) PRE; s.n. (7) PRE, Moll 1228 (4) K, PRE; 2466 (7) K, PRE, Montgomery 18 (20) NBG. Morley 21 (12) PRE; 92 (14) M, PRE. Morris 396 (19) NBG. Mortensen 200 (14) C. Moss 5644 (16) BM; 5760 (1) BM. Moss & Rogers 1263 (4) BM. Mostert 1185 (4) PRE. Mudd s.n. (4) K. Muir 12, 240 (14) NBG, PRE; 14 (7) PRE; 2441 (12) PRE; 4469 (14) K. PRE; 4485 (15) BOL, PRE; 4496 (1) K, PRE; 4538 (14) PRE; 683 (14) BOL. PRE. Mund s.n. (19) BM, NBG. Munro s.n. (15) PRE. Museum Botanicum Hauniense s.n. (1) C; (6) C; s.n. (7) C; s.n. (14) C.

Nanni 123 (7) PRE. Nel 219 (4) PRE. Niven & Laubert s.n. (5) S. N.J.A. s.n. (6) S. Noel 322 (14) GRA; Noel s.n. (7) GRA; s.n. (19) GRA.

Obermeyer 258, 1078, TRV30044 (4) PRE. O*Callaghan 273, 752 (14) PRE; 708 (1) NBG; 839 (7) NBG, PRE; 1003 (7) GRA; 1425 (7) NBG. Oldenland, Herb. Schreb. s.n. (14) M. Oldevig-Roberts 120 (7) S. Olivier 118 (15) M. PRE; 593 (19) K, PRE; 2027 (7) PRE, 2988 (14)

94 Bothalia 33,1 (2003)

GRA; 3024 (7) GRA; 3197 (11) PRE; 3679 (15) K, PRE; 3797, 4555, 5194 (15) PRE; 5226, 5342 (14) PRE; 10318 (15) NBG. Onderstall 919, 1269 (4) PRE. Orchard 312 (14) C, MO, PRE, S. Osbeck s.n. (14) S. Osborne 126 (7) GRA.

Page 97 (15) PRE. Palmer 1094 (15) PRE; 1417 (4) GRA; 3932 (20) GRA. Pappe s.n. (6) NBG; s.n. (7) S; s.n. (14) GRA; s.n. (14) NBG. Parker 3843 (1) K NBG; 4109 (1) K, PRE. Parsons 60, 182 (7) NBG; 61, 112 (1) NBG; 169 (14) PRE; 172 (19) PRE; 322 (7) NBG, PRE. Paterson 1123 (7) GRA: 270 (20) GRA: 833 (15) BOL: 883 (15) PRE: TRV25754 (20) PRE. Paterson-Jones 697 (7) NBG. Pedro & Pedrogao 8095 (4) BOL, PRE. Peeters, Gericke & Burelli 387, 407 (4) MO, PRE. Pegler 234 (7) PRE. Penther 2891 (14) M, S, W; s.n. (14) M, S. Perold & Fourie 2256 (4) PRE. Peterson 1263 (16) BOL. Phillips 228 (4) PRE; 1294 (14) NBG; 1503 (15) NBG; 1622, 3363A, B (7) K, PRE; s.n. (14) NBG. Phillipson 624 (4) MO, UPS; 1140 (4) K, PRE. Pillans 2792 (14) BOL; 3779 (14) BOL, PRE; 3783 (1) BOL, PRE; 7689 (9) BOL; 8030 (14) BOL; 8513 (6) BOL; 9240 (1) BOL; 17158 (10) NBG; s.n. (1) GRA; s.n. (14) BOL, MO. Pole Evans 129, 986 (4) PRE; 4373 (6) PRE. Pons s.n. (4) PRE). Potts 1281 (14) PRE; BLF1300 (14) PRE; BLF288 (7) GRA, PRE; s.n. (14) NBG. Primos 41 (10) PRE. Prosser 2050 (4) PRE. Purcell s.n. (7) NBG; s.n. (14) NBG. Putterill s.n. (4) PRE.

Quickelberge A7758 (7) GRA.

Raal & Raal 296 (4) PRE. Raitt s.n. (15) PRE. Ramsay 1592 (14) GRA. Rattray 382 (4) BOL; 909 (4) K; s.n. (4) PRE. Rauh & Schlieben 9788 (4) M, PRE. Rechinger A-4413 (4) M. Reed 35 (14) GRA. Rehm s.n. (1) M. Renny 179 (4) PRE; Rennie 480 (14) BOL, GRA; Rennie 2613 (4) BOL). Repton 6253 (4) PRE. Retief 12 (20) PRE; Retief 1194 (7) MO, PRE. Rivers-Moore s.n. (7) GRA. Rob & Fries 3395 (11) UPS, S. Roberts 1953, 1954, 1973, 2001, 3359 (4) PRE. Robinson 1873 (4) K, MO. Rodin 1305 (19) BOL, K, MO, PRE. Rogers 788, 21919, 23675 (4) PRE; 4724, 15512, 16701, 26983 (14) K; 16701A, 16705 (15) K, PRE; 16703, 17222 (15) K; 17281 (14) BM; 26574 (14) GRA. NBG. PRE; 26774, 26790 (7) PRE; 26987, 27017 (14) K, PRE; 27004 (19) BM, NBG. PRE; 27026 (14) PRE; 28018 (7) GRA; 28983 (19) GRA. NBG. Rosenberg & Rutherford 308 (12) NBG. Rourke 1204 (14) K, PR. PRE, S; 1505 (7) K, NBG; 3000 (14) NBG. Rutprum s.n. (14) S. Rycroft 1331 (1) NBG; 2135 (6) NBG; 3000 (19) S; 3117 (14) S; s.n. (1) K, S, TCD.

Salter 6220 (1) BM, K; 6352 (19) BM, BOL, K; 7051 (6) K; 9370 (14) BM; s.n. (14) BOL. Sankey 69 (4) K, MO. Savage Cat. 504.2 (1) LINN. 504.3, Sp. 161 (1) LINN. Scharf 1013 (11) PRE; 1023 (19) PRE; 1067 (11) PRE; 1069, 1522 (15) PRE; 1578, 2001, 2005 (19) NBG, PRE; 1596 (11) NBG, PRE; 1689 (19) K, NBG, PRE; 1958 (11) PRE; 1959 (11) K, PRE; 1961 (20) K, PRE; 1985, 2002 (19) PRE; 2033 (19) GRA, PRE. Scheepers 1831 (4) MO, PRE. Schlechter 1363 (1) C, PRE. Schlieben s.n. (4) M. Schmidel s.n. (14) M. Schmidt 41 (15) PRE; 567. 569 (14) M; 56 (4) PRE. Schmitz 8265, 8314 (4) PRE. Schonland 3054 (11) GRA, PRE; 3398, 3525 (14) GRA, PRE. Schrire 1869 (15) GRA; 1968, 1969, 2037, 2038 (20) GRA; 2083 (19) GRA. Schweickerdt 759 (4) PRE. Seutloali 94 (4) PRE. Shearing 870, 891 (15) PRE. Shumane P58 (20) GRA. Sidey 1219 (20) S; 1696, 1812 (14) MO, PRE, S; 2312 (15) MO, NBG, S; 3595, 3801 (14) PRE, S. Sieber 74 (14) BOL, HAL, M, NBG, P, PRE, S, W. Sieber & Zeyher s.n. (14) MO. Sim 1 (7) GRA; 20 (14) K; 68 (4) K PRE; 1380, 1499 (14) C, NBG; 1471 (4) K; 19659 (14) PRE; 2595 (4) K. Simon 657 (4) K, PRE. Simpson 97 (15) NBG, PRE. Smart 15512 (14) PRE. Smit 22 (4) PRE. Smith 4637, 4649, 5081 (14) PRE. Smuts 91 (4) PRE; 1134 (6) PRE; s.n. (7) NBG; s.n. (14) NBG. Smuts & Gillett 2162 (4) BOL, NBG, PRE; 2469 (4) PRE; 3179 (4) NBG. Smuts & Pole Evans 933 (4) BOL, K. Snyman s.n. (7) GRA. Sparrman s.n. (1) S; s.n. (7) M, S; s.n. (14) S; s.n. (19) M, S. Stam 43, 174, 429 (4) PRE. Staples 17 (4) PRE). Starke s.n. (15) NBG, PRE. Stayner 24 (14) GRA, PRE. Stephen 455 (7) PRE. Stokoe 1790, 1811 (10) PRE; 2542, 3199 (5) PRE, K; 2800, 2802, 22329 (5) BOL, K. NBG, PRE; 6335, 8677, 8977 (15) BOL; 8226 (16) BOL; s.n. (1) NBG; s.n. (5) NBG, PRE; s.n. (14) NBG; s.n. (15) NBG, PRE. Stopp 70 (7) M. Story 2071 (4) MO, PRE; 3109 (14) PRE; 3162 (20) PRE; 3559 (19) K, M, PRE; 3667 (4) PRE; 3778 (4) GRA PRE; 3885 (14) GRA; 3896 (4) GRA, PRE; 3899, 3900 (4) PRE. Strauss s.n. (12) NBG. Strey 715 (1) PRE; 6769 (7) PRE; s.n. (14) M. Sutherland 185B (4) TCD; s.n. (4) K; s.n. (7) K. Swartz s.n. (6) M. Symons 144 (4) PRE

Taylor 2953 (7) PRE; 3413 (20) NBG, PRE; 3802, 8166 (1) NBG, PRE; 3803 (12) PRE; 4042 (6) K, M, PRE; 4143, 8367 (7) NBG; 5281, 7987, 7988 (14) PRE; 6211, 9871 (6) NBG, PRE; 6996 (1) NBG, PRE, S; 7123 (1) K, MO, PRE; 7562 (15) K, PRE; 8998 (7) PRE, S; 9356 (15) K, MO, PRE; 9879, 9905, 10144 (7) NBG, PRE; 10022 (1) PRE, K; 10171, 10249 (7) MO, NBG, PRE. Teague 312 (4) BOL, K, Theron 282 (7) PRE; 631 (14) PRE; 1076 (7) PRE, K; 2097 (14) PRE; 2182 (4) PRE; 2216 (14) M,

PRE. Thoday 13 (14) NBG; 42 (14) NBG; 52 (14) BOL, NBG; 100, (16) C, NBG, PRE; 212, 212A (10) BOL, K. NBG; 214 (15) BOL, NBG; 216 (1) K, NBG, PRE. Thode 993 (19) K, MO, PRE; 1120 (14) PRE; 1639 (4) PRE; A292 (4) K, MO, PRE; s.n. (4) NBG. Thomas s.n. (4) GRA. Thompson 1427 (15) NBG; 1856 (19) PRE; 1874 (20) PRE; 2005, 2151 (15) NBG, PRE; 3300 (14) PRE; 3303 (14) MO, PRE; 3318 (14) K, MO, PRE; 3374 (15) PRE; 880 (19) K, PRE; 903 (20) NBG, PRE. Thunberg s.n. (14) UPS. Topper 122 (1) NBG. Torre & Perreira 12683 (4) C. Toughton 156 (14) GRA. Trauseld 435 (13) PRE; 860 (13) PRE. Trimity College s.n. (14) TCD. Tyson 77 (10) GRA; 1449 (19) K, NBG, PRE; 2178 (20) K, NBG; s.n., TRV17233 (7) PRE.

Ueckermann 7793 (19) PRE.

Vahrmeijer & Tolken 252 (7) PRE. Van Breda 131 (15) PRE; 639 (15) K; 679 (14) PRE; 754 (15) K, PRE; 823 (1) PRE; 1037 (7) PRE; 1633 (1) PRE; 1664 (14) PRE. Van Dam TRV 23994 (14) PRE. Van der Merwe 964, 1225 (14) PRE; 1108 (12) PRE; 1186 (1) PRE; 2420 (15) K, PRE; s.n. (4) K, PRE. Van der Schifff 4478, 4836, 5592, 5845, 6191 (4) K, PRE; 5376 (4) PRE. Van der Walt 250 (15) PRE; 409 (14) PRE. Van der Westhuizen 144, 147 (7) PRE. Van der Zeyde s.n. (4) MO. NBG, S. Van Jaarsveld & Sardien 10998 (20) PRE. Van Niekerk 95 (19) NBG. Van Rensburg 151 (14) PRE; 442 (15) PRE; 443 (1) K, M, NBG, PRE; 444 (14) PRE; 459 (7) NBG; 2147 (7) PRE; 2148 (14) PRE. Van Schalkwyk 46 (7) PRE. Van Vuuren 1632 (4) PRE. Van Wilgen 163 (14) PRE. Van Wyk 390 (14) K, PRE; 1545 (7) PRE, PRU; 1736 (12) M, PRE; 1920 (14) PRE, PRU; 2517 (7) PRE, PRU; 2899 (4) PRE, PRU; 3167 (7) PRE, PRU; 6704 (4) PRE, PRU. Van Zinderen Bakker 12 (6) NBG. Van Zyl 3366 (15) NBG, PRE. Venter 7123, 11013, 12721 (4) PRE. Victor 400 (15) PRE; 498 (14) PRE. Vlok 1235 (15) PRE. Von Gedow 473 (14) GRA.

Wager TRV10772 (4) PRE. Wahl 42 (19) NBG; Wall 30, s.n. (15) S; s.n. (6) S; s.n. (7) S; s.n. (14) S. Wallich s.n. (19) K. Walsh s.n. (6) NBG. Walters 1 (15) NBG; 247, 1084 (6) C, NBG; 384, 1017 (15) K, M, NBG. PRE; 1769 (15) NBG; 2143 (14) NBG. Ward 549, 2167, 4374, 5735, 7211 (7) PRE; 6944, 6954 (4) PRE; s.n. (14) TCD. Watt & Brandwijk 1851 (4) PRE. Wawra 3 (14) M, PRE. Weigend 2341 (7) M. Wells 2998 (14) PRE; s.n. (14) MO. Welman 798 (4) PRE. Weltz 743 (15) NBG. Wendelberger 406 (15) PRE. Werdermann & Oberdieck 476 (14) B, PRE; 691, 693 (14) B, PRE; 1129 (4) K, PRE; 1560 (4) B, K, PRE; 1560 (4) K. Werger 1054 (4) PRE; 1806 (4) MO, PRE. West 181 (13) PRE; 358 (20) GRA: 485. 1392 (4) PRE. Westfall 716, 756 (4) PRE. White 95 (14) GRA; 5230 (1) PRE. Wild 1352 (4) K, PRE. Willemse 805 (12) NBG, PRE. Williams 440 (1) K MO; 1008 (7) K, M; 1451 (14) NBG; 2624 (14) MO, PRE. Williamson s.n. (20) TCD. Wilman s.n. (14) PRE. Wilms 2277 (7) K. Wirminghaus 178 (14) GRA. Without collector 170 (15) TCD; s.n. (7) TCD; s.n. (10) TCD. Wolley-Dod 1575 (1) PRE, K; 1878 (6) BOL, K; 2927 (1) BM, K. Wood 12179 (4) NBG; 1712 (7) BM, BOL, K; 4036 (4) BOL, K, GRA; 5786 (7) M, MO; 6592 (7) S; 6601, 11405 (4) BM, K; 9139 (7) BOL, NBG; s.n. (4) NBG, K; s.n. (7) PRE. Worsdell s.n. (7) NBG. Wright 1029 (4) PRE; s.n. (1) K, MO, P, TCD; s.n. (6) C, P; s.n. (14) K; s.n. (14) TCD. Wurtz 208 (20) NBG; 1210 (15) NBG; 1211 (14) NBG; 1567 (1) NBG; 2121 (19) NBG.

Youthed 663 (20) GRA.

Zantovska 120 (15) PRE. Zeyher 34 (14) NBG; 38 (19) BOL; 41 (11) G, S; 42 (6) NBG; 44 (11) G, NBG, S, W; 117.11 (4) S; 156 (20) BM, BOL, K, NBG, TCD; 277 (19) BOL, K, NBG, TCD; 405 (7) TCD, GRA; 1025 (11) BM, NBG, TCD; 3777 (7) NBG; 3778 (19) NBG; 3779 (20) BM, MEL, NBG, S W; 3780 (11) G, P, S, W; s.n. (6) NBG; s.n. (14) C, K, MO, P; s.n. (19) S. Zietsman 323 (4) PRE. Zietsman & Zietsman 427, 512 (4) PRE.

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REFERENCES

- ACOCKS, J.P.H. 1988. Veld types of South Africa, edn 3. Memoirs of the Botanical Survey of South Africa No. 57: 85–87.
- ADANSON, M. 1763. Familles des plantes. Vincent, Paris.
- ANGIOSPERM PHYLOGENY GROUP (APG). 1998. An ordinal classification for the families of flowering plants. Annals of the Missouri Botanical Garden 85: 531–553.
- ARCHANGELSKY, D.B. 1971. Palynotaxonomy of the Thymelaeaceae s.l. In L.A. Kuprianova & M.S. Jakovlev, *Pollen morphology*: 104–234. Academy of Science, USSR, Komarov Botanical Institute, Leningrad (in Russian).
- ARNOLD, T.H. & DE WET, B.C. (eds). 1993. Plants of southern Africa: names and distribution. *Memoirs of the Botanical Survey of South Africa* No. 62: 1–825.
- BALL, J. 1878. Spicilegium Florae Maroccanae. *Journal of the Linnean Society* 16: 281–742.
- BARTLING, F.G. 1840. Passerina incurva. Linnaea 14: 404.
- BENTHAM, G. & HOOKER, J.D. 1880. Genera plantarum, vol. 3. Cramer, Weinheim.
- BERGIUS, P.J. 1767. Descriptiones plantarum ex Capitae bonae Spei. Salvius, Stockholm.
- BEYERS, J.B.P. 2001. The Cape genus *Lachnaea* (Thymelacaceae): a monograph. *Strelitzia* 11: 1–115.
- BIGGS, C., KNİGHT, R., RAITT, L.M. & KEATS, D. 2001. The use of vegetation indicators for the assessment of coastal erosion in Milnerton South Africa. 27th Annual Conference of the South African Association of Botanists. RAU, Johannesburg. Unpublished.
- BOISSIER, P.E. 1838. Elenchus plantarum novarum. Lador & Ramboz, Genève.
- BOISSIER, P.E. 1842. Voyage botanique dans le midi de l'Espagne 2. Paris. BOISSIER, P.E. 1846. Diagnosis plantarum orientalium novarum 1. Herman, Leipzig.
- BOLUS, H. & WOLLEY-DOD, A.H. 1904. A list of the flowering plants and ferns of the Cape Peninsula, with notes on some of the critical species. *Transactions of the South African Philo*sophical Society 14: 207–373.
- BOND, P. & GOLDBLATT, P. 1984. Plants of the Cape flora: a descriptive catalogue. *Journal of South African Botany* Suppl. vol. 13. National Botanic Gardens of South Africa, Claremont.
- BREDENKAMP, C.L. & VAN WYK, A.E. 1996. Palynology of the genus *Passerina* (Thymelaeaceae): relationships, form and function. *Grana* 35: 335–346.
- BREDENKAMP, C.L. & VAN WYK, A.E. 1999. Structure of mucilaginous epidermal cell walls in *Passerina* (Thymelaeaceae). *Botanical Journal of the Linnean Society* 129: 223–238.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2000. The epidermis in Passerina (Thymelaeaceae): structure, function and taxonomic significance. Bothalia 30: 69–86.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2001a. Leaf anatomy of the genus *Passerina* (Thymelaeaceae): taxonomic and ecological significance. *Bothalia* 31: 53–70.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2001b. Taxonomic significance of inflorescences, floral morphology and anatomy in *Passerina* (Thymelacaceae). *Bothalia* 31: 213–236.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2002a. Systematics of Passerina truncata and a new subspecies monticola. Bothalia 32: 65–71
- BREDENKAMP, C.L. & VAN WYK, A.E. 2002b. Passerina quadrifaria (Thymelaeaceae): a new species from the southern Cape and Little Karoo in South Africa. South African Journal of Botany 68: 304–307.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2002c. A new species of Passerina from Western Cape, South Africa. Bothalia 32: 76–79.
- BREDENKAMP, C.L. & VAN WYK, A.E. 2002d. Taxonomy of the Passerina filiformis complex (Thymelaeaceae). Bothalia 32: 29–36.
- BREYNE, J. 1678. Exoticarum plantarum centuria. David-Fredericus Rhetius, Gdansk.
- BRUMMITT, R.K. 1992. Vascular plant families and genera. Royal Botanic Gardens, Kew.

- BRUMMITT, R.K. & POWELL, C.E. 1992. Authors of plant names. Royal Botanic Gardens, Kew.
- BUBANI, P. 1897. Flora Pyrenaea, vol. 1. Ulrico Hoepli, Milan.
- BUNGE, A.A. VON. 1833. Enumeratio plantarum, quas in China boreali. Collegit St Petersburg.
- BUNNIGER, L. 1972. Untersuchungen über die morphologische Natur des Hypanthiums bei Myrtales- und Thymelaeales-Familien. Beiträge zur Biologie der Pflanzen 48: 79–156.
- BURDET, H.M. 1979. Cartulae ad botanicorum graphicem. XV. Candollea 34: 203, 204.
- BURMAN, J. 1739. Rariorum Africanarum plantarum. Henricum Boussiere, Amsterdam
- BURMAN, N.L. 1768. Flora indica: cui accedit series zoophytorum indicorum, nec non prodromus florae capensis. Cornelius Haak, Leiden; Johannes Schreuder, Amsterdam.
- CAFFERTY, S. & BEYERS, J.B. 1999. Proposal to reject the name Lachnaea conglomerata (Rhamnaceae). Taxon 48: 171, 172.
- CAMBESSÈDES, J. 1827. Enumeratio plantarum quas in insulis Balearibus. Bélin, Paris.
- CLUSIUS, C. 1601. Rariorum plantarum historia. Joannem Moretum. COATES PALGRAVE, K. 1977. Trees of southern Africa. Struik, Johannesburg.
- COSSON, E.S. & DURAND, E.M. 1856. Passerina microphylla. Bulletin de la Société de France 3: 744.
- COUTINHO, A.X.P. 1909. Thymelaea broteriana. Boletim da Sociedade Broteriana 24: 145.
- DESFONTAINES, R.L. 1798. Flora Atlantica, vol. 1. Desgranges, Paris. DOMKE, W. 1934. Untersuchungen über die systematische und geographische Gliederung der Thymelaeaceen. E. Schweizerbart sche Verlagsbuchhandlung (Erwin Nägele) G.M.B.H., Stuttgart.
- DRÈGE, J.F. 1843. Zwei pflanzengeographische Documente. In E. Meyer, Besondere Beigabe zur Flora 2: 44–229. Reginsburg, Leipzig.
- DRÈGE, J.F. 1847a. Standörter–Verzeichniss der von C.L. Zeyher in Südafrika gesammelten Pflanzen. *Linnaea* 19: 584–591.
- DRÉGE, J.F. 1847b. Vergleichungen der von Ecklon und Zeyher und von Drège gesamelten südafrikanischen Pflanzen (so weit dieselben noch vorhanden) mit den Eksemplaren von Zeyher's neuesten Sammlungen, welche derselbe zum Verkauf stellt. Linnaea 20: 183–258.
- DUBY, J.E. 1828. Aug. Pyrami de Candolle, Botanicon gallicum 1. Mme ve Bouchard-Huzard, Paris.
- ENDLICHER, S.F.L. 1847. Generum plantarum supplementum quartum, part 2. Fr. Beck, Vienna.
- FARR, E.R., LEUSSINK, J.A. & STAFLEU, F.A. 1979. Index nominum genericorum (plantarum). Scheltema & Holkema, Utrecht.
- FEDER, N. & O'BRIEN, T.P. 1968. Plant microtechnique: some principles and new methods. *American Journal of Botany* 55: 123–142.
- FISCHER, F.E.L. VON. & MEYER, C.A. 1839. Passerina vesiculosa. Bulletin de la Société des naturalistes de Moscou: 170.
- FORSSKÅL, P. 1775. Flora aegyptiaco-Arabica. Möller, Copenhagen. FORSTER, G. 1786. Floralae insularum australium prodromus. Göttingen.
- GANDOGER, M. 1913. L'herbier africain de Sonder. Bulletin Société Botanique de France. Paris: 414–422.
- GILG, E. 1894a. Thymelaeaceae. Die natürlichen Pflanzenfamilien 3,6a: 216–245.
- GILG, E. 1894b. Studien über die Verwandtschaftsverhältnisse der Thymelaeales und über die 'anatomische Methode'. Botanische Jahrbücher 18: 489–574.
- GMELIN, J.F. 1791. Systema naturae, part. 1. Georg Emanuel Beer, Leipzig.
- GMELIN, J.F. 1792. Systema naturae, part. 2. Georg Emanuel Beer, Leipzig.
- GOLDBLATT, P. & MANNING, J. 2000. Cape plants. A conspectus of the Cape flora of South Africa. Strelitzia 9. National Botanical Institute, Cape Town and Missouri Botanical Garden.
- GREUTER, W., MCNEIL, J., BARRIE, F.R., BURDET, H.M., DE-MOULIN, V., FILGUEIRAS, T.S., NICOLSON, D.H., SILVA, P.C., SKOG, J.E., TREHANE, P., TURLAND, N.J. & HAWKS-WORTH, D.L. 2000. International Code of Botanical Nomenclature (Saint Louis Code). Koeltz Scientific Books, Königstein.
- GRIFF, W. 1854. Notulae ad plantas asiaticas 4. Bishop's College Press, Calcutta.
- GUNN, M. & CODD, L.E. 1981. Botanical exploration of southern Africa. Balkema, Cape Town.

- GUSSONE, G. 1843. Flora Siculae Synopsis 1. Tramater, Naples.
- HANSEN, B. & WAGNER, P. 1998. A catalogue of the herbarium specimens from Captain Cook's first and second expeditions housed in the Copenhagen Herbarium (C). *Allertonia* 7: 352, 353
- HARVEY, W.H. 1868. Order CV. Thymeleae. In J.D. Hooker, *The genera of South African plants* 2: 323–327.
- HEINIG, K.H. 1951. Studies in the floral morphology of the Thymelaeaceae. *American Journal of Botany* 38: 113–132.
- HILLIARD, O.M. & BURTT, B.L. 1987. The botany of the southern Natal Drakensberg. Annals of the Kirstenbosch Botanic Gardens 15. National Botanic Gardens, Cape Town.
- HILLIARD, O.M. & BURTT, B.L. 1988. Notes on some plants of southern Africa chiefly from Natal: XIV. Notes from the Royal Botanic Garden Edinburgh 45: 77–94.
- HOLMGREN, K., HOLMGREN, N.H. & BARNETT, L.C. 1990. Index herbariorum. International Association for Plant Taxonomy, New York Botanical Garden, New York.
- HUTH, E. 1898. Passerina passerina. Helios 11: 135.
- IUCN Species Survival Commission 2000. IUCN Red List Categories.

 As approved by the 51st Meeting of the IUCN Council. Gland,
 Switzerland.
- JACKSON, B.D. 1917–1918. Linné–Tulbagh correspondence in possession of the Linnean Society, being a fifth contribution towards the history of the Linnean collections. *Proceedings of the Linnean Society* 130, Suppl. 1–13: 5–13.
- JUSSIEU, A.L. DE. 1789. Genera plantarum. Viduam Herissant & Theophilum Barrois, Paris.
- KARELIN, G.S. & KIRILOV, I.P. 1842. Passerina ammodendron. Bulletin de la Société des naturalistes de Moscou 15: 444.
- KARNOVSKY, M.J. 1965. A formaldehyde-glutaraldehyde fixative of high osmolality for use in electron microscopy. *Journal of Cell Biology* 27: 137a.
- LAMARCK, J.P.A.P.M. DE. 1792. Tableau encyclopédique et méthodique. Botanique (Illustration des genres) 2. Pancoucke, Paris.
- LAMARCK, J.P.A.P.M. DE & CANDOLLE, A.P. DE. 1805. Flore Française, vol. 3. Agasse, Paris.
- LAMARCK, J.P.A.P.M. DE & CANDOLLE, A.P. DE. 1815. Flore Française, vol. 6. Desray, Paris.
- LAPEYROUSE, P.P. DE. 1813. Histoire abrégée des plantes Pyrénées. De l'Imprimerie, de Bellagarigue, Toulouse.
- LEANDRI, J. 1930. Recherches anatomiques sur les Thyméléacées. Annales des Sciences Naturales Botanique 12: 125–237.
- LEVYNS, M.R. 1950. Thymelaeaceae Reichb. In R.S. Adamson & T.M. Salter, *Flora of the Cape Peninsula*: 594–603. Juta, Cape Town.
- LINNAEUS, C. 1737. Hortus Cliffortianus. Amsterdam.
- LINNAEUS, C. 1753. Species plantarum, edn 1. Salvius, Stockholm.
- LINNAEUS, C. 1754. Genera plantarum. Salvius, Stockholm.
- LINNAEUS, C. 1755. Centuria I. plantarum. Höjer, Uppsala.
- LINNAEUS, C. 1756. Centuria II. plantarum. Höjer, Uppsala.
- LINNAEUS, C. 1760. Amoenitates academicae 6. Godofriedum Kiesewetter, Leipzig.
- LINNAEUS, C. 1767. Systema naturae, edn 12, 2. Salvius, Stockholm.
- LINNAEUS, C. 1771. Mantissa plantarum altera. Salvius, Stockholm. LINNAEUS, C. 1784. Systema vegetabilium, edn 14 (edn J.A. Murray).
- Dieterich, Göttingen.
- LINNAEUS, C. fil. 1782. (*1781*) Supplementum plantarum. Impensis Orphanotrophei, Brunswick.
- LITARDIÈRE, R.V. DE. 1938. Passerina corsica. In J.I. Briquet, Prodrome de la flore corse 3: 6. Lechevalier, Paris.
- LODDIGES, C. 1816. Catalogue of plants. Plummer & Brewis, London.
- LUBKE, R.A. 1998a. Dune Thicket. In A.B. Low & A.G. Rebelo, Vegetation of South Africa, Lesotho and Swaziland: 15. Department of Environmental Affairs & Tourism, Pretoria.
- LUBKE, R.A. 1998b. Terrestrial ecology. In R. Lubke & I. de Moor, Field guide to the eastern and southern Cape coasts 26: 402–343. University of Cape Town Press, Rondebosch.
- LUBKE, R.A. & VAN WIJK, Y. 1998. Terrestrial plants and coastal vegetation. In R. Lubke & I. de Moor, *Field guide to the east-ern and southern Cape coasts* 21: 289–343. University of Cape Town Press, Rondebosch.
- MABBERLEY, D.J. 1989. The plant-book. Cambridge University Press, Cambridge.
- MAGALLÓN, S., CRANE, P.R. & HERENDEEN, P.S. 1999. Phylogenetic pattern, diversity, and diversification of eudicots. Annals of the Missouri Botanical Garden 86: 297–372.
- MARLOTH, R. 1925. *The flora of South Africa*, vol. 2,2. Darter, Cape Town.

- MEISNER, C.F. 1840. Passerina L. Linnaea 14: 390-404.
- MEISNER, C.F. 1857. Ordo CLXVII. Thymelaeaceae. In A.L.P.P. de Candolle, *Prodromus systematis naturalis* 14: 493–605. Masson, Paris
- MEYER, C.A. 1843. Remarques sur les genres de Daphnacées sans écailles périgynes, et exposition des caractères des ces genres. Annales des Sciences Naturelles, Seconde Série, Botanique 20: 45–53.
- MILLER, P. 1768. The gardener's dictionary, edn 8. London.
- MIQUEL, F.A.W. 1863. Annales musei botanici lugduno-batavi, vol. 3. C.G. van der Post, Amsterdam.
- NORLINDH, T. & WEIMARCK, H. 1958. Beiträge zur Kenntnis der Flora von Süd-Rhodesia X. *Botaniska Notiser* 3: 623–631.
- PALMER, E. & PITMAN, N. 1972. Trees of southern Africa, vol. 3. Balkema, Cape Town.
- PAU, C. 1887. *Notas bótanicas à la flora espanôla* 1. Escuela Tipográfia del Hospicio, Madrid.
- PAU, C. 1904. Passerina granatensis. Carta botánicos 1: 7.
- PAU, C. 1922. Passerina argentata. Memóries del Museu de Ciències naturals. Barcelona. Ser. Bot. 1., No. 1: 67.
- PAU, C. 1925. Passerina pubescens (L.) Loscos var. virgata. Memóries del Museu de Ciêncies naturals. Barcelona. Ser. Bot. 1., No. 3: 33.
- PETERSON, B. 1958. Beiträge zur Kenntnis der Flora von Süd-Rhodesia 10. *Botaniska Notiser* 111: 625–628.
- PETERSON, B. 1959. Some interesting species of *Gnidia. Botaniska Notiser* 112: 465–480.
- PHILLIPS, E.P. 1944. Notes on some genera of the Thymelaeaceae. Journal of South African Botany 10: 61–66.
- PLUKENET, L. 1700. Almagesti botanici mantissa. Sumptibus autoris, London
- POIRET, J.L.M. 1804. Encyclopédie méthodique (Dictionaire Encyclopédique de botanique) 5. Paris.
- PRESL, C.B. 1844. Botanische Bemerkungen. Gotlieb Haase Söhne, Prague.
 RADFORD, A.E., DICKISON, W.C., MASSEY, J.R. & BELL, C.R.
 1974. Vascular plant systematics. Harper & Row, New York.
- RAFINESQUE, C.S. 1836. Flora telluriana, part 4. Auctore C.S. Rafinesque, Philadelphia.
- RAMOND DE CARBONNIÈRES, L.F.E. 1800. Passerina dioica. Bulletin de la Société philomatique de Paris 41: 131.
- REBELO, A.G. 1998. Fynbos Biome. In A.B. Low & A.G. Rebelo, Vegetation of South Africa, Lesotho and Swaziland: 62–74. Department of Environmental Affairs & Tourism, Pretoria.
- ROUY, G.C. 1910. Flore de France, vol. 12. Foucaud, Rochefort.
- ROYAL BOTANIC GARDENS, KEW. 1997. *Index kewensis* on Compact Disc. The Word Factory and System Simulation Ltd., Oxford University Press, Oxford.
- SAMPAIO, G.A. & DA SILVA, F. 1913. Herbário Português. Tipografia Costa Carregal, Porto.
- SAVAGE, S. 1945. A catalogue of the Linnaean Herbarium. Linnean Society of London.
- SCHOUSBOE, P.K.A. 1800. lagttagelser over Vextriget: Marokko. K.H. Seidelin, Copenhagen.
- SCHRADER, H.A. 1810. Passerina tartonraira. Neues Journal für die Botanik 4.1: 89.
- SENNEN, F. 1924. Passerina imbricata. Annales de la Société linneene de Lyon: 69.
- SIEBOLD, P.F. & ZUCCARINI, J.G. 1846. Florae japonicae familiae naturalis, vol. 2. Lugdini batavorum, Leiden.
- SIM, T.R. 1919. Flowering trees and shrubs for use in South Africa. The Speciality Press of South Africa, Johannesburg.
- SMITH, C.A. 1966. Common names of South African plants. *Memoirs of the Botanical Survey of South Africa* No. 35: 589. Botanical Research Institute, Pretoria.
- SMUTS, L.M. 1996. *Passerina paludosa*. Information system for endangered plants, Cape Nature Conservation, Stellenbosch.
- STAFLEU, F.A. & COWAN, R.S. 1986. *Taxonomic literature*, vol. 6. Scheltema & Holkema, Utrecht.
- STEARN, W.T. 1973. *Botanical Latin*, edn 2. David & Charles, Newton Abbott.
- STEUDEL, E.G. VON. 1841. Nomenclator botanicus, edn 2. Cottae, Stuttgart.
- STORY, R. 1952. A botanical survey of the Keiskammahoek District. *Memoirs of the Botanical Survey of South Africa* No. 27: 1–184. Department of Agriculture, Pretoria.
- TAKHTAJAN, A.L. 1997. Diversity and classification of flowering plants. Columbia University Press, New York.
- TAN, K. 1980. Studies in the Thymelaeaceae II: a revision of the genus Thymelaea. Notes from the Royal Botanic Garden Edinburgh 38: 189–246.

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