BULLETIN OF THE
Dipterists
Forum

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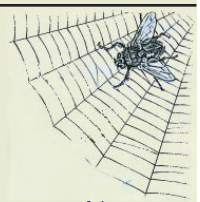

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Dipterists Forum Forum www.dipteristsforum.org.uk/index.php

Photographs: Front cover (top) *Poecilobothrus nobilitatus* (Dolichopodidae), pond at Chute Farm, Whipsnade, 2 Jul 2011, det J O'Sullivan **Alan Outen,** (bottom) *Subclytia rotundiventris* (Tachinidae) by R. Ivel, Clifton, Beds. 16 Aug 2011 **Alan Outen**, (above) *Cordilura albipes* (Scathophagidae) from Flitwick Moor, Beds, 15 Jul 2011, det Stephane Lebrun, **Alan Outen**. Other photographs as supplied by the authors or the editorial panel who would be pleased to receive illustrations for general purposes - many thanks for those already sent. If you want to catch the next front cover, please note that the orientation must be upright (portrait)

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Dipterists Forum Events

The following Newsletters and other special items are incorporated into the package for the printers after completion of the Bulletin. They are not to be found in any pdf version of this Bulletin and they have their own pagination. Please contact the Newsletter editors for full colour pdfs, back issues may also be found on DF website.

Hoverfly Newsletter #52
Empid & Dolichopodid Newsletter #17
Cranefly Newsletter #23
Flowers for flies - Judy Webb
Booking form for meetings
Guidelines for Bulletin contributions

Desktop publishing: Darwyn Sumner

Editorial

Biodiversity in the UK

I've been keeping my eye on the activities of the UK All Party Parliament Group (APPG) on Biodiversity which I mentioned in the last Bulletin.

The purpose of the All Party Group is to provide a forum for cross party Parliamentarians, senior policy makers, academics, leading industry figures, and other interested parties to have an informed discussion on all aspects of protecting biodiversity in the UK and abroad. "Our primary role is to make best use of this forum to help facilitate Parliamentary debate on biodiversity."

In November the APPG launched a members' inquiry into 'Planning & Biodiversity' (that's "members" as in Members of Parliament). Danny Stevens (APPG Secretariat) told us that "*The Government has committed to* "retain protection and improvement of the natural environment as core objectives for local planning." *It has also stated that the planning system will contribute to the overall objective of no net loss of biodiversity.*" and then detailed the Inquiry which APPG had initiated and which involved consultation with interested parties - that's us. Dipterists Forum aren't involved directly with this but we are part of a wider network, the NBN.

The NBN has made comments to the Inquiry, these were made under the following headings:

1. Support the local and national infrastructure that makes quality assured biodiversity data available to inform planning decisions.
2. Provide guidance on best practice regarding the use of biodiversity data in the planning process.
3. Monitor the use of biodiversity data in the planning process.
4. Enforce existing legislation for the conservation of biodiversity.
5. Publish clear objectives and targets for biodiversity conservation to inform local action.

The paper produced by the NBN is quite substantial and difficult to summarise but here's an example from #4. above which gives a bit of a flavour of it:

*The Localism Act 2011 enables planning permission to be granted by parish councils and [...] empowers community organisations to put forward development proposals under a Community Right to Build Order [...] [...] Government must stress that the NERC Act applies to parish councils, community groups and neighbourhood forums when operating in a planning context [...] [...] many Local Planning Authorities do not feel compelled to comply with the NERC Act 2006 (*basically this Act says *"have regard to the purpose of conservation of biological diversity in the exercise of their functions").*

The Localism Act lets every Tom, Dick and Harry grant planning permission, Tom knows the biodiversity conservation rules and mainly chooses to ignore them, Dick and Harry have never even heard of the rules.

So if you think you are doing your bit by providing records to be put onto the NBN Gateway, think again; most planning applications go through without any attempt to look at what might be there.

A nice authoritative document, well worth a read if you are concerned about conserving Diptera. If you want to find it, try the NBN website.

Meniscus & trickle midges

Henry Disney is currently scouting around for a new organiser for the Dixidae & Thaumaleidae recording scheme. See NBN eNews for details (request it from support@nbn.org.uk).

Citizen Science cuts

I'm told this term originated in the USA. Sadly we hear the news that the Federal Government has withdrawn funding for the American equivalent of our NBN, the NBII (National Biological Information Infrastructure), and it is to close down entirely in February. Read all about it at http://www.nbii.gov/portal/server.pt/community/termination_of_nbii_program/2057/termination_faqs/7650

The University of Birmingham, School of Biosciences has also announced that it will cease to host the 'Biological Recording and Species Identification' programmes (UCert, PG Cert, PG Dip, MSc and day schools).

Biological Recording in Scotland

Strictly speaking there's no Diptera news in this, but we've a trip to Scotland planned this year so the BRISC newsletter might be worth a read to find out what's going on north of the border: http://brisc.org.uk/newsletters/Pending/BRISCRecorderNews84.pdf

ALERC appointment

Financial support for an ALERC National Coordinator has been provided by ALERC and the NBN Trust. Tom Hunt (of Cheshire's LRC: Ecorecord) was recently appointed and whilst the LRC part of his job will only affect us Dipterists who are involved with their LRCs, he's also got to "facilitate the adoption of new biodiversity data management technology and encourage exchange of ideas" so this will see him working with online recording initiatives, I guess he'll bump into our iSpot man Martin Harvey before long. Tom can be contacted at tom.hunt@alerc.org.uk

NatureSpot

I believe there are a number of these initiatives, like iSpot but at a County level. I've just had the following from David Nicholls (Leicestershire & Rutland) "*Diptera are obviously a tricky group so in time it would be really helpful to get your views on which species can be identified from an image and which need examination.*" which really is an issue that warrants a view or statement from Dipterists Forum - maybe the comments from Martin Harvey and Roger Morris in the following articles say it all?

Views and reviews

Many thanks for the photographs, it's useful to have a selection handy when compiling the Bulletin, even if they don't all get used.

Please keep your eye open for things that might be of interest to the readers of the Bulletin. Books on Diptera are not published very often but there are other topics like conservation, recording and biodiversity that definitely interest us. Equipment for photography, microscopy, collecting and breeding too. Do drop a note to the editors.

Darwyn Sumner

Notice board

Diptera on iSpot

iSpot (www.ispot.org.uk) encourages people to learn about wildlife identification, by enabling novices and experts to meet online. It is developed by The Open University as part of the OPAL project, and was launched in mid-2009.

Since the launch over 16,000 people have joined iSpot, sharing nearly 90,000 observations of wildlife and helping each other to sort out which species is which. Advice on identification comes from the involvement of many experts and representatives, including from museums, recording schemes and societies (see www.ispot.org.uk/representatives), whose help is greatly appreciated.

One of the strengths of OPAL and iSpot has been the focus on taking natural history and environmental science to groups of people who may not previously have had the chance to participate in such activities. For iSpot, much of this work has been delivered by our team of Biodiversity Mentors, funded through OPAL in each of ten English regions, and by an Open University grant in Ireland, Wales and Scotland. They have been introducing iSpot and related wildlife activities to audiences from a very diverse set of backgrounds, many of whom have been thrilled to receive feedback on their observations from the other users on iSpot.

Among the experts active on iSpot are several who represent Dipterists Forum and its associated recording schemes. So far (for Britain), around 3,400 observations of roughly 360 species of Diptera have been posted on iSpot. Given iSpot's aim of encouraging 'beginners' to develop their interest in wildlife observation, the top ten most frequently posted species are probably not all that surprising, being mostly large, well-marked species that make themselves obvious to the casual observer: the hoverflies *Episyrphus balteatus*, *Volucella pellucens*, *Scaeva pyrastri*, *Volucella zonaria*, *Eristalis pertinax*, *Myathropa florea* and *Eristalis tenax*, plus the bee-fly *Bombylius major*, the tachinid *Tachina fera* and the muscid *Mesembrina meridiana*.

Although iSpot is not targeted at rare and unusual species, rather at encouraging people to take an interest in the wildlife they see around them, there have been many discoveries of species in places where they hadn't previously been recorded, and among the less common species posted on the site are several observations of the Hornet Robberfly, plus the hoverfly *Pocota personata* (www.ispot.org.uk/node/76428), the tachinid *Gymnosoma rotundatum* (www.ispot.org.uk/node/35669) and the rarely seen bot-fly *Gasterophilus pecorum* (www.ispot.org.uk/node/3874). Of course, not all photos of flies can be identified to species level, and one of iSpot's aims is to help people learn when they need to take specimens, and encourage them to join organisations such as DF to take their interest further.

We now have a data download system in place to allow the export of iSpot observation data for particular taxonomic groups into a spreadsheet format. We are starting to pass data on to those recording schemes that wish to receive it (including some of the DF schemes). Although iSpot is intended to help people learn how to identify wildlife rather than as an online recording system, we are keen to make the iSpot data available where relevant. Recording schemes receiving the iSpot data will have to make decisions about how much of it can be fully verified and imported into their main datasets, but we hope the spreadsheet export format will make this relatively easy to do, with hyperlinks back to the original observations in iSpot so that unusual records can be checked.

That's Chew Valley in Somerset, not Saddleworth, thanks Roger, Ed.

If you have any questions about how you can make use of iSpot to help encourage novice dipterists, or about representing a recording scheme and receiving data for your scheme, please contact Martin Harvey at the Open University (m.c.harvey@open.ac.uk).

Martin Harvey

Developing a Dipterists Forum database – records from the internet

Having spent a long while trawling the internet for hoverfly records, I have latterly turned my attention to other families because there does not seem to be an established mechanism for harvesting Diptera data from the internet. This has resulted in a database of 6,000 potentially identifiable photographs with location and date information attached. About 1,600 of these photographs have already been identified and it is proposed to incorporate these into the DF dataset. Apart from my efforts, I know that Matt Smith does the dame for the Tachinidae but there are photographs of many other flies that hithertohave not scrutinised and recorded.

Data that have been extracted have been placed on a spreadsheet

Found something interesting? Don't forget to tell the scheme organiser. Spare specimens would probably be appreciated too, pop them into a small store box throughout the year and take to Field Meetings or our AGM.

for uploading onto the DF dataset on RECORDER that I maintain. The data collected up until 20 December comprised:

Source	Positive ID	Partial ID
ISpot	500	1086
WildaboutBritain	44	80
Flickr	1067	3473
DF Website	1	0
Other sources	188	279

Specialists will be invited to check unidentified photographs and so the dataset might grow considerably. It is unlikely that all photographs will be identifiable but if half of the photographs are identified then that will be a fairly sizeable chunk of data. Ultimately, this will mean that the Dipterists Forum section of the NBN will include a file of data abstracted from websites.

Readers will doubtless question the merits of making this effort. It has involved several tens of hours of work (probably extending to hundreds) and for the most part the data involve relatively common species. Rarer or important species have been reported as part of this process. For example, several records of *Asilus crabroniformis* and *Bombylius discolor* have been secured. More noteworthy still is the tantalising record of the beefly *Systoechus ctenopterus* from the Chipstead Valley (Surrey) recognised by David Gibbs in 2010. It therefore seems logical to look upon the contributors to identification forums and to photographic sites as parataxonomists who make a valuable additional contribution to our knowledge of the British fauna.

There are notable additional benefits. For example, where important species have been photographed I have followed up the source and have sought more accurate data. This means that we are gradually gaining contact with a cohort of active field naturalists who might not otherwise submit diptera data. Several people have started to send data to the Hoverfly Recording Scheme on a regular basis as a result and I am hopeful that a wider group of casual recorders will be established. Perhaps some will become members of the Forum and will go on to more detailed recording.

Proposal for a central Dipterists Forum Dataset

In the past 6 months, the major backlog of Dipterists Field Meeting data has been computerised and now amounts to about 80mb of data. These data will go on the NBN in due course. However, Dipterists Forum has no formal means of absorbing and managing diptera records that are not captured by Recording Schemes.

This means that it is likely that considerable numbers of records are not readily available for initiatives such as new recording schemes or analyses for conservation initiatives. In addition, several Recording Schemes appear to be inactive and experience with the Hoverfly Recording Scheme has shown that in the absence of interaction between the scheme and potential contributors, the numbers of records submitted rapidly decline.

Now that the DF dataset is largely computerised, the process of incorporating new data is a much less daunting prospect. It therefore makes sense to extend our record-keeping activities to encourage recorders to submit all data to a central database that can be used to distribute records to particular schemes for validation. In the case of schemes known to be active, data would be forwarded directly to the scheme organiser. Where the scheme is believed to be inactive, or there is no scheme, data would also be retained within the Recorder format so that it can be accessed and used as required.

This approach brings several advantages:

- A data assembly initiative might encourage new recorders to submit records of species that are currently not covered by recording schemes.
- The database could form a repository for all diptera records so that members could be assured that data is not lost in the event of their death.
- The project might be used to develop a network of data translators – turning historic diary and museum data into an accessible dataset.

Where Recording Schemes fail to interact with DF, a new recorder can be appointed and the bulk of the data retrieved and validated.

Proposal

DF should establish a central database that is maintained and regularly updated on the NBN.

I am volunteering to maintain and input new data during the immediate future.

A data management initiative should be developed, with regular updates in the Bulletin and direct contact with active Dipterists to determine how many might be prepared to lodge full data with the Forum.

Roger Morris
7 Vine Street, Stamford, Lincolnshire

Conservation

News from the Conservation Officer

As the new Committee member elected to take on the conservation and BAP roles previously done so very capably by Barbara Ismay, I have much to learn but am looking forward tremendously to the task. Barbara, who will continue to act as Co-ordinator of '**Adopt a Species**' until I have my feet under the table, has already provided me with a lot of useful background material, and together with her husband John, has kindly offered me her full support. I very much hope I can build on Barbara's excellent work and initiatives to take forward the conservation of flies.

My background working for many years with Natural England and its predecessor bodies English Nature and the Nature Conservancy Council should help me with the legislative and bureaucratic processes. I am, however, fairly new to the magnificent world of flies, but from my experience at field and indoor meetings am confident that other committee members will provide me with the guidance and support I shall no doubt need. I also look forward to working closely with Duncan Sivell, who represents Buglife on the Forum.

As with Barbara before me, I should be very grateful if members could let me have any news pieces relevant to fly conservation, especially about those 35 species that are recognised as priorities for conservation in the UK Biodiversity Action Plan (BAP). If you think there are any policy or strategic nature conservation documents the Forum should express a view on, do please let me know.

I in my turn shall be particularly pleased to help Judy Webb our Publicity officer. Raising the profile of flies is critical to their effective conservation. With another hat on, I recently drafted a

webpage for the Devon Moth Group in which I said that making gardens more attractive to moths would help a wide range of other wildlife too, from bees and flies to birds. Symptomatically, the editor promptly edited out the word flies replacing it with ladybirds. I compromised by saying hoverflies! It's quite a challenge to increase the public appeal of flies to that of butterflies, moths or bumble bees, but we must try. My task starts at home, convincing my wife that the only good fly is not one squashed on the wall or stuck to the flypaper. I shall keep you informed….

Habitat protection for the pine hoverfly and the aspen hoverfly?

The Scottish Government is currently consulting on whether the aspen hoverfly *Hammerschmidtia ferruginea* and pine hoverfly *Blera fallax* should be added to Schedule 5 of the Wildlife and Countryside Act. This would make it an offence to damage or destroy or any structure or place harbouring these very rare flies, already listed as UK BAP priority species. The intent is to encourage better woodland management, such as continuous cover forestry rather than clear felling. In particular it will help to protect the small number of stumps and logs which are critical for their larvae. Since collectors usually search for larvae to breed through, and in the process destroy or severely damage their very restricted deadwood habitats, on behalf of the Forum I propose to respond supporting the scheduling. Of course, any *bona fide* researchers, such as Ellen and Graham Rotherhay who have done excellent research on the two species leading to vastly improved understanding of their ecology, will be able to apply for licences.

Robert Wolton
Conservation Officer
robertwolton@yahoo.co.uk
01837 810416

ADOPT A SPECIES

This scheme is hoping to find dipterists willing to conduct some research (field or desk based) on a fly species or group of species or in a certain area or habitat. Further details can be found in several Bulletins in 2007 and 2008 or on the Dipterists Forum webpage, where you can find it in the Forums section. This scheme is particularly for BAP, RDB or notable species or areas where these can be found. If you have any information you would like to share with fellow dipterists, then I would like to hear from you. Also, if you would like to take on a species or help threatened species by conducting some more general research, then please contact me.

News from 'Adopt a Species'

I would like to thank all of you who already adopted a species and have contributed to this or other Bulletins or kept me updated so that I could summarise your work.

Your work is very encouraging and I hope that some other dipterists might follow. Currently 16 of our 35 BAP species and 4 species with conservation status have been adopted. Thank you very much for all your hard work and good luck with your quests. I would very much like to receive updates on any of the adopted species, so please get in touch.

I have now handed the BAP and Conservation officer roles to Rob Wolton and hope members will continue to support Rob in this role. I have retained the 'Adopt a Species' role until Rob has settled in. Unfortunately due to illness I have been unable to chase members for updates.

Eggs of *Dorycera graminum*?

Dorycera graminum numbers on this dry grassland site in North Essex were down from the peak of 75 counted in 2010 to 55 on 17th May 2011. However, this was the first count in an early season and I may have missed the peak which is usually around two weeks later in early June.

D.graminum is easily found and counted here as both sexes assemble on fence posts and wire across the site. Behaviour includes courtship wing waving, and pairing. Females probe both posts and wire apparently seeking ovipositing sites, although eggs have not been detected before this year.

Previous attempts to sieve pupae in the spring from soil and turf near to the fence have been unsuccessful, so this year I removed part of the tops of posts where probing was noted. The posts are sweet chestnut erected about 30 years ago and showing some rot, cracks and beetle borings.

On one sample piece eggs could be seen in cracks and on another post with more decay which broke partly during sample removal, eggs were seen more easily. They were mostly stacked in clumps of at most 20 in redundant beetle borings and rotted areas no more than 6 to 8 mm deep below the surface. The eggs were 0.7 mm long, spindle shaped, creamy white and slightly yellow at the narrow end, and apparently smooth at lower magnification. Some moribund eggs, drunken and distorted, were also present, conceivably from 2010.

To attempt to rear the eggs the clumps, attached to their wood, were placed in 3" x 1" tubes and larger plastic containers with a piece of kitchen roll to hold added moisture. Chopped *Agrostis* leaves and shaken roots were added to the containers when larvae started to hatch after about a week.

The hatched larvae were off the otitid type (K G V Smith `Introduction to the Immature Stages of British Flies`, p. 196.) white, slightly segmented and tapering towards the head end, mouthparts just visible within.

Hatched larvae immediately dispersed from the wood and were seen moving on the sides of the tubes but appeared disinterested in the supplied roots and leaves. Larvae often rested in a semicircular position and when disturbed were able to flick themselves about 10 times their own length, as well as normal crawling.

Unfortunately after about 10 days the larvae had all died, apparently without feeding.

It first appears that neither rotted wood nor leaves and roots of grass (at least *Agrostis*), are the normal food of *D.graminum*.
David Scott

If any of you, who are good at rearing have suggestions for David, could you please get in contact with me, Barbara? I am happy to pass this on as this looks rather promising.

Contacting authors

If you wish to contact any of the authors, where an email address is not given, then please email me (Barbara Ismay) and I will forward this to them, or try and get in contact with them via the Dipterists Forum webpage. You can post a query or information for the author under Forum and there under 'Adopt a species' if you are a member of Dipterists Forum

Please contact me again if you have not heard from me in response to an email as we have managed to lose some emails or not received them in the past.

Barbara Ismay Co-ordinator of 'Adopt a Species'
e-mail: schultmay@insectsrus.co.uk or telephone: 01844-201433

The blog of RSPB's Mark Avery is worth looking at (www.markavery.info/blog). He has an incisive article in December 2011's British Wildlife where he speaks of our £2 billion going to English and Welsh farmers via CAP each year. He says *"I fear that some in the farming industry have forgotten the priveleged position they occupy"*.

Recording flies on RSPB nature reserves

Almost 2900 species of flies have been recorded on RSPB nature reserves, but we are sure there are more to be found. With habitats from salt marsh to mountain summits, RSPB reserves are a rich hunting ground for dipterists. Most reserves welcome visits from naturalists wanting to record wildlife, as long as the RSPB receives a copy of the records, but please contact the site staff before you visit because some areas may be sensitive to disturbance. Visits can also be arranged through Mark Gurney at the RSPB's head office (phone no.:01767 693426 or email: Mark.Gurney@rspb.org.uk). A species list for a site can be useful, but advice on habitat management is even more valuable, so if you are able to include details of habitat requirements for the species you find, or an assessment of the habitat at the site, please do so. You can find details of your nearest reserve at www.rspb.org.uk/reserves/area/.

B. Ismay

Dipterists Digest: 50 issues

Dipterists Digest volume 18 part 2 (December, 2011) was the 50th issue, so I thought it was appropriate to mark the occasion.

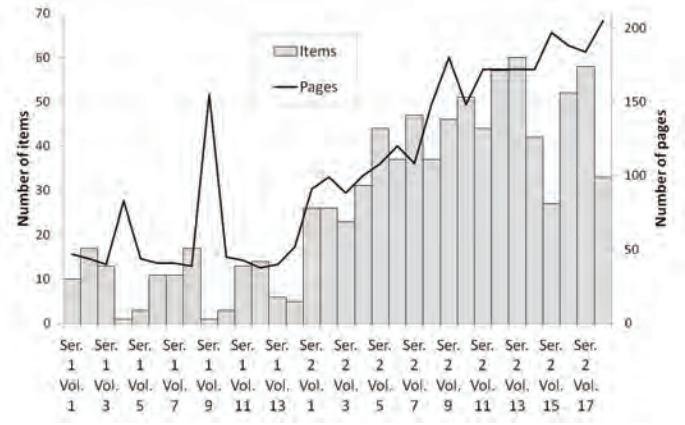
I hope that members know that the DF web-site includes a facility to view and search the contents of the Digest. You can look up any issue and see its contents (and front cover) and you can search for articles by author's name, Diptera family or words from the title. This is driven by a database which I update soon after each new issue is published. Therefore, it contains details of every item so far published – and it is that information on which I base this article.

First, a bit of history: Dipterists Digest was launched by Derek Whiteley in 1988 and it was both edited and published by him on behalf of the Central Panel of Diptera Recording Schemes. When Dipterists Forum was founded in 1994, the Digest became the journal of the new society. At this time Graham Rotheray took over as editor and he began the "second series". DF took over publication from Derek from volume 2 of the second series in 1995 and Peter Chandler became editor from volume 5 in 1998. The latest innovation has been the use of colour, with colour plates first appearing in 2004 (Vol. 11, No. 1) and a colour cover in 2010 (Vol. 17, No. 1). (Although Graham Rotheray's "Colour Guide to Hoverfly Larvae", 1st series, Vol. 9, 1993, was of course in colour).

There has been a general trend for the number of pages to increase over time and the latest volume, with 205 pages, is the biggest so far. In total, 3,404 pages have been published!

The 50 issues so far have included 866 items written by 268 different authors. Peter Chandler has been the most prolific author by far, producing 124 items – although that does include the updates to the checklist and Irish list, reports on the exhibits at Dipterists Day and occasional editorials which account for 61 of them. Even so, he is well ahead of David Gibbs, Martin Drake, Graham Rotheray and Alan Stubbs – all with counts in the 30s.

I categorise items by the family(ies) they cover and distinguish "lists" – such as catalogues of species from a site. Excluding the "lists", items about Syrphidae are by far the most popular, with 141 items, followed by Dolichopodidae (56), Tachinidae (49) and Tephritidae (31). Interestingly, Nematocera don't appear until 8th and 9th place with Limoniidae (22) and Tipulidae (20). Overall, 87 families have featured (out of about 108 in Britain - depending on exactly how they are split), so quite a high proportion of the fauna has received at least some sort of coverage. Items reporting species new to Britain are very popular, but there have been 27 papers which included descriptions of new species – 37 of them from 14 families.



We owe considerable gratitude to Derek Whiteley for initiating the Digest and to Graham Rotheray and Peter Chandler for all their work in keeping it going. We also depend on several others, behind the scenes, who are involved in proof reading, refereeing and distribution. I think that Dipterists Digest is a journal of which we can be proud. Long may it continue!

Stuart Ball

Members

Membership Matters

At the AGM in November 2011, John Showers was elected as the new Membership Secretary. Many thanks to Mick Parker, who has undertaken the role for a number of years. He is not stepping down completely and will continue to handle distribution of publications to new members.

All subscriptions, changes of address and membership queries should now be directed to John Showers at:

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Membership and Subscription Rates:

Members and Subscribers are reminded that subscriptions are due on 1st January each year. The rates are as follows:

Home

Dipterists Forum: £6 per annum. This includes the Bulletin of the Dipterists Forum.

Dipterists Digest: £9 per annum.

Please note that a number of people are still paying at the old pre-2005 rate so please check and amend your payments accordingly.

Overseas

Dipterists Forum and Dipterist Digest: £20 pa.

There is only this one class of membership. Payment must be made in Pounds Sterling.

John Showers

Correspondence

Keeping it rough

Dear All

Many thanks for the hard work you have put in on getting the [*Empid & Dolichopodid*] Newsletter publishable.

The Bulletin goes from strength to strength and you have done a fantastic job transforming it from a rough rag to a smart production. Can I make one small plea... In continuing to develop the Bulletin, please do try to preserve some of its 'seat of the pants' roughness and originality. It's a place for exchange of news, views, ideas and information (some of it tentative) and the more journal-like it becomes, the less likely it is to retain this flavour perhaps? A very polished production might put off some potential contributors?

Best wishes for 2012

Hope to see you in the field with net-full's of interesting flies during the year ahead

Cheers

Adrian [Plant]

Adrian hangs around in Wales (ed)

So here's some "rough seat of the pants" stuff especially for Adrian, an email thread I've done my best to sort out, with apologies to those who tried their best to keep it serious:

Peter Chandler 27th October

Martin refers in the recent Bulletin to variant spellings of the commonly used abbreviation of dolichopodids, more than once misconstrued when overheard by passers by. This inconsistency

Honey trap scandal: the myth that refuses to die "*If the bee disappeared off the surface of the globe then man would only have four years of life left.*" **Albert Einstein.** Falsely attributed by European beekeepers grumbling about cheap imports, this quote first turned up in 1994, 39 years after his death but was **"**quoted" again in Independent on Sunday 29/1/12. An enquiry has been launched via a worm hole in the space/time continuum.

is seen in the newsheets of the Empid and Dolichopodid Study Group that began to appear in 1986, where at least three spellings are used. The first March 1986 issue is headed "Dollies and Empids Study Group" while the second August 1986 issue has "Empid and Dolie Newsheet No. 2 and Dolies is used throughout that issue. Dolis is used in Newsheet No. 5 (March 1988). I haven't checked further to see if there are cases of more than one spelling in a single issue.

The question arises as to who first coined this abbreviation and when. I had assumed it would have happened within living memory. To Colyer & Hammond they were "long-headed flies", which as Martin points out is an erroneous translation, and Fonseca avoided giving them a common name.

However, I recently came across the following sentences, which show that doubt over spelling is not new:

"I have practically finished my overhaul of the year's captures among the Dollies."

"The Dolies and Empids I had better go through before submitting to you."

Any suggestions as to the author and date of the above? If you know of any other early uses of this or of other popular abbreviations of fly names please let me know.

Anthony Bainbridge 27th October

Peter's note has made me cast my mind back to the days when Roy Crossley and I started the group. I recall, as a relative newcomer against Roy's already substantial reputation, deferring to his

acknowledged expertise on the dolichopodids by assuming that we would call it D&E - which was both polite and alphabetic, and is actually slightly easier to say in a hurry. (I had tended to focus on the E's and had not mastered d'Assis-Fonseca at all.) There's no doubt we and other contributors did indeed become lazy in our spelling and in the order of the initials.

But 'dollies' is inexcusable.

Barbara Ismay 28th October

We came across other abbreviations:

Tachs, Strats, Sphaeros, empids, Tephs

May be more, but I can't remember all of them. A lot of these originated in acronyms used when data was first written down by hand and seems to be used nowadays in emails as well.

We recently saw the following used in a report (not by us):

a window-gnat Anisopodidae
a st. marks-fly Bibionidae
a small phytophagous fly Chloropidae
a parasitic conopid fly Conopidae
a diastatid fly Diastatidae
a meniscus midge Dixidae
a long-legged fly Dolichopodidae
a fruit-fly Drosophilidae
a predatory fly Empididae
a shore-fly Ephydridae
a heleomyzid fly Heleomyzidae
deer fly Hippoboscidae
a dance-fly Hybotidae
a lauxaniid fly Lauxaniidae
a crane-fly Limoniidae
a spear-winged fly Lonchopteridae
a phytophagous fly Opomyzidae
a hairy-eyed crane-fly Pediciidae
a scathophagid fly Scathophagidae
a marsh fly Sciomyzidae
a black scavenger fly Sepsidae
a lesser dung-fly Sphaeroceridae
a hover-fly Syrphidae
a parasitoid fly Tachinidae

I am not sure what to make of these.

Peter Chandler 31st October

Thank you Barbara for those comments.

I think most of the familiar abbreviations you list are, as you say, recent shorthand. Strats is a little older and empids is the original family abbreviation when it was Empidae. Mycets has been used (not by me) but gnats is shorter. The English names for flies only identified to family are presumably from some version of Recorder – I hope the site assessment didn't depend on that list.

Regarding Anthony's comment that Dollies is inexcusable, this was preferred in the English names for BAP species *Dolichopus laticola*, the Broads Dolly-Fly and *D. nigripes*, the Black-footed Dolly-Fly, from which it appears that Dolly-Fly is now the adopted family name, though I used long-legged flies in the Dipterist's Handbook..

The quotations I mentioned of 'dollies' are more than a century old as they come from letters written to Verrall by Colonel Yerbury. As Verrall's replies don't exist I can't say who used it first. A fuller dated list appears below, from which it appears he changed to one "l" in 1908, in one case deleting the second "l" after writing it. Continuity of usage rather than being newly re-invented is harder to confirm. Peter Dyte may know.

Red Dolly - What is it? (16/7/06) [in box sent for naming; presumably a brassy specimen]

Gravesend will be worth working for Dollies alone (30/7/06) [is it worth going now?]

There are one or two interesting dollies in my Studland lot (8/7/07)

I will bring my Dollies, Empids & other odds and ends to the next meeting of the Ent. Soc. (25/11/07)

I have practically finished my overhaul of the year's captures among the Dollies (13/10/07)

As regards the Dollies ... (13/1/08, with the second l crossed through)

The Dolies and Empids I had better go through before submitting to you (15/9/08)

The following account was written from the Haven Hotel, Sandbanks, Parkstone, Dorset, on 20/5/07, referring to a visit to a site known to both of them as 'The Green Pond' and heavily populated with gulls. Perhaps Mick knows it.

'I nearly came to unmitigated grief near there – I saw a specimen of Microdon and in attempting to catch it stepped on some Sphagnum and went in nearly up to my waist – Microdon of course escaped.'

No indication of his state of dress on return to the hotel.

There's clearly nothing new in dipterology

Alan Stubbs 31st October

It is so intuitive to shorten Dolichopdididae to doli/dolies that I suspect it has been re-invented a number of times since the Yerbury era. Correct English spelling or not I prefer one 'l' as an abbreviation of a scientific word rather than per a child's doll/dolly (and

innuendos of the latter form of spelling).

As regards family names, my generation (includes Peter) were weaned on Colyer & Hammond as the standard work (the first complete review of the British fauna written for the naturalist.) We have adopted some additional English names for families but there have been a number of introductory books etc that have devised their own names to create a mess. The family splits in recent decades have added to the complexity, and in any case Colyer & Hammond did not nominate an English name for all the smaller families. At another scale, I am not aware of common usage English names for higher groupings such as Nematocera, Aschiza and Acalypterates and Calypterates, whilst the Larger Brachycera Recording Scheme was devised in parallel to Macro & Micro moths (where actual size is not the sole criterion).

At one level this does not matter, but at the level of popularising flies and in catching the attention of the media (as per Buglife) it is relevant.

Of course were have further abbreviations, such as pips (Pipunculidae) and pezids (platypezids).

Peter Chandler 31st October

I thought that Buglife had endorsed (proposed?) having 2 'l's with the names of the BAP species.

Contrary to Colyer & Hammond I've never called them long-headed flies.

Alan Stubbs 2nd November

As it happens I don't think these English name spellings were my doing midst a rush to invent English names for hundreds of species. Anyway, I doubt whether innuendoes of having 2 'l's were spotted. This not the only BAP ambiguity of this type, and some scientific names have changed, so the BAP list of official names is not immutable.

I still hold the view that the most logical abbreviation of the family name is 'doli', and that spelling avoids other connotations. If DF were to decide that should be the official spelling I suspect others would adjust to that ruling.

Peter Chandler 3rd November

I think it is probably too late to expect that the spelling will be standardised, but regardless of spelling the pronunciation will always suggest that dolly is intended and that was clearly the intention by whoever proposed Dolly-Fly for the BAP species.

I was at Oxford again yesterday and now have copies of Yerbury's correspondence with Collin, in which Dollies are often mentioned so Collin may have provided the continuity of usage.

Roy Crossley 4th November

If Yerbury was the first to call them 'dollies' then it all makes sense.

Yerbury was an old soldier with an obvious soldiers highly sexist sense of humour, and at the time there could be no doubt what he was getting at by using that word. Do you want me to spell it out?

Better by far to stick entirely to the scientific names then we wont get mixed up in slightly risqué nineteenth century aspersions!

As an aside, the song 'Goodbye Dolly Grey' was the army's top of the pops in the Boer War - according to my long dec'd grandfather!!

Anthony Bainbridge 7th November

Yerbury obviously liked dollies with long legs. I wonder if his own were decorated in any way?

Peter Chandler 7th November

Roy is quite right about the song although according to the attached it first appeared in the Spanish-American War of 1898. Dolly was of course a familiar form of Dorothy. Yerbury had a lady friend called Dora Isaac, who was a nurse and in his diaries is referred to as Nurse Dora or Sister Dora. I suppose he may also have known her as Dolly. In the article by Graham Rotheray about Yerbury's activities in Scotland (Digest 1997 4, 92-99) he mentions that she joined him on one of his trips there. That was in 1913 and she sadly died in the following year aged 41, but he had often mentioned taking her to the theatre and restaurants from 1900 onwards, as well as a day trip to Brighton in 1903 (they went down by coach, dined at the Old Ship and returned by train). I haven't yet tracked down whether photos of either Yerbury or Dora exist, so can't comment on leg length. Dora was half German as her father Albert (an optician) was born in Cologne.

Alan Stubbs 2nd November

Do dolichopodids sing? Drosophilids for instance have song.

It is said that given enough monkeys and enough time one of them may reproduce the plays of Shakespeare.

Are you now suggesting that a dolichopodid can come up with a complete rendition of an 1898 song? If not, that scuppers your inferred spelling, and even if they could sing it, I doubt whether they could spell it.

I maintain my case for 'doli'.

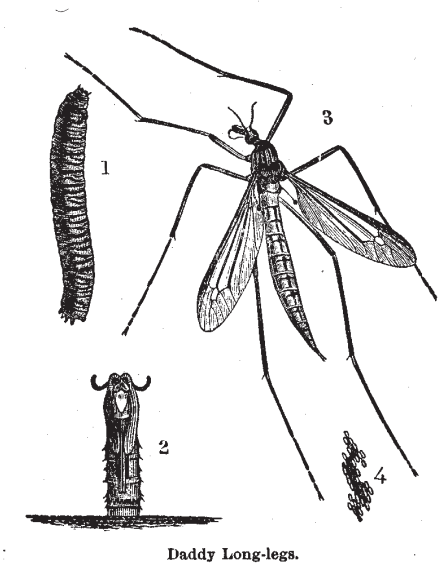
and there the Dolichopodid story seemed to end. However Peter then picked up the thread that Alan had started, in a similar vein, on Tipulidae:

Daddies & gad flies

Peter Chandler 10th November

Thank you Alan for those useful comments on singing dolis

John Henry Wood of Tarrington, who was Yerbury's cousin, also had a copious correspondence with Verrall and Collin. He seems to have always said dolichopids but was more colloquial with craneflies, always referring to them as daddies



Daddy Long-legs.

Yerbury rarely referred to craneflies but when he did so also called them daddies (e.g. a "*Ctenophora ruficornis*" ovipositing on a birch

stump was called a daddy, though a female). Wood was more into craneflies and often included them in boxes of specimens he sent to Verrall for checking, with comments such as "The little daddy I fancy must be *Cheilotrichia imbuta*". As that's a tiny yellow species, daddies wasn't just reserved for the big ones.

It appears, however, that Yerbury had more affinity than Wood for mud. In referring to the site for a *Clinocera* species Wood says: "I know to an inch where the *Clinocera* swarmed – but a real beastly spot it is, a thin boggy surface much poached by cattle on a steep and slippery clay bank, a place that only Yerbury with his great boots and disregard for dirt could enjoy".

Gad flies is one of the names for Tabanidae cited by Colyer & Hammond. Yerbury urged Verrall to restrict the name gad flies to oestrids as he thought applying it also to horse-flies caused confusion.

Malcolm Smart 10th November

I suppose the J. Henry Wood you refer to was the man after whom the Promenade Concerts were named - perhaps he got an orchestra of crickets to accompany him while he promenaded around diptera sites singing 'Goodbye Dolly Grey' and waving his net at dollies and their (sugar) daddies!!

John & Barbara Ismay 11th November

Singing is actually quite widespread among Diptera! It occurs in fruit flies (Tephritidae), leaf miners (Agromyzidae) and grass flies (Chloropidae). It is used to attract mates! Reed gall flies (Lipara) were the first insects shown to have local dialects in their songs (by Kanmiya). We do not know a case of singing dollies, but surely they would win Diptera Come Dancing with their wing-waving and leg acrobatics! Perhaps, since many live on rather smelly mud, they have to make an extra effort to attract their dollie mates.

Alan Stubbs 15th November

Ah. 'Grass flies', new one to me but I like it.

Daddies(& sugar daddies). That term has been long extinct.

Who has the equipment to listen in to dolis?

Perivale Wood - a little extra

Darwyn Sumner 8th October

(to London Wildlife Trust/GIGL - London's LRC)

Hi Mandy, sorry to trouble you with something as mundane as a site, but can you shine any light on this place that might be useful to the Dipterists Forum people who have been asked to take a look at Perivale Wood LNR by Peter Edwards

I'm guessing it's in your patch.

Mandy Rudd 10th October

Yes, it's in our patch – as Peter mentions their data end up with us via London Natural History Society.

What would you like to know? I can ask my colleague to run you a free data search on the site if that's easiest? It'll at least give you an overview of the site as a SINC/LNR, the habitats recorded and the species data we hold already.

Let me know what you'd ideally like from us and we'll get on with it for you.

So there you are, you "Perivale Wooders", get a free site search from the UK's premier LRC before you go. I've passed details to John Kramer - he knows the ropes about these reports - they should prove useful on the day. (Ed)

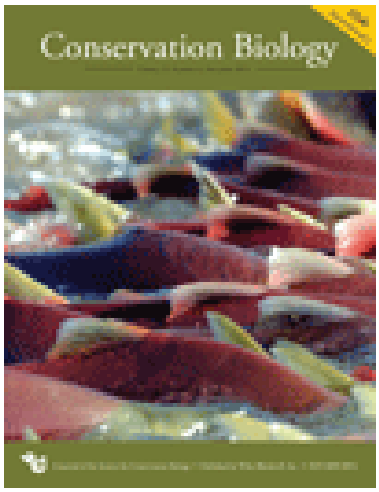
Perivale Wood is threatened by the HS2 rail link (Ed)

Review

Publications

Conservation & collections

Drew, Joshua. The Role of Natural History Institutions and Bioinformatics in Conservation Biology, Conservation Biology. Volume 25, Issue 6, pages 1250–1252, December 2011; Available (free) from: http://onlinelibrary.wiley.com/doi/10.1111/j.1523-1739.2011.01725.x/full#



Introduction: *Natural history institutions (museums, herbaria, zoos, aquaria, etc. whether privately owned or existing in conjunction with universities) and the collections they house play a large role in documenting species diversity. Historically, these collections were a rich source of data that ultimately fueled developments in biogeography and systematics and provided the intellectual framework for conservation biology ...*

via Martin Harvey

Books

Diptera - forthcoming

A proposed new RES Handbook on Moth Flies (Psychodidae)

Volunteers required for testing

Phil Withers' key to the British species of this family was published in 1989 as an issue of Dipterists Digest. Since then a number of additional species and an additional genus have been recorded in the British Isles, increasing the total from 89 to 99. Phil has now thoroughly revised his text and prepared a new handbook to be published in the RES Handbooks series in the style of the recent handbook on Lonchaeidae. Many of the genitalia figures in this new handbook will be those drawn by François Vaillant to illustrate the Palaearctic keys to this family, with others by Phil. A generic key to larvae to assist in recognition of the early stages will also be included.

Volunteers are sought to test the keys in this new handbook, which is likely to become available for testing during the first half of 2012. Please let me know (chandgnats@aol.com, 01225708339) if you are willing to participate in the testing and a copy of the text will be sent to you when it is available.

Peter Chandler

Diptera

A DIPTERIST'S HANDBOOK

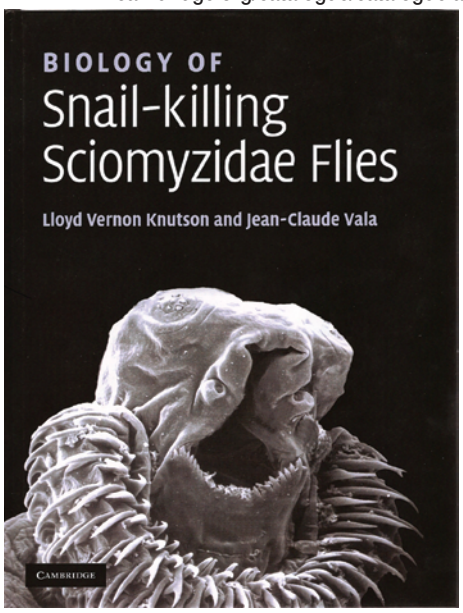
This are still a small number of copies available at the price of £32 if buying in person from Peter Chandler (at Dinton Pastures or at field meetings during 2012) or from Erica McAlister at the NHM, or at £36.68 including postage and packing within the UK

(contact Peter Chandler at chandgnats@aol.com).

Sciomyzidae

Knutson, L.V. & Vala, J-C 2010. Biology of Snail-Killing Sciomyzidae Flies. Cambridge University Press. 584 pp., £85, hardback. ISBN-10 0521867851 ISBN-13: 978-0521867856.

Available from Pemberley Books, Amazon.com or CUP at http://www.cambridge.org/catalogue/catalogue.asp?isbn=9780521867856



My first publication on flies was on the predatory habits of a sciomyzid fly. Ever since I have regarded Sciomyzidae is being among the more beautiful flies.

From the 19th Century onwards there were reports of their larvae feeding on dead snails and pupae being found in empty snail shells. However, it was not until Berg's report of 1953 that it was realised that some larvae of these flies were killing the snails. Since then his former students and others have transformed our knowledge by demonstrating that the larvae are now known, with a few exceptions, to be feeders on molluscs. Furthermore, while a few feed on dead snails or bivalve molluscs, most are now known to be predators or parasitoids of living molluscs, be they aquatic or terrestrial.

This book critically reviews our current knowledge, including larval habits and host/prey preferences, phenology, reproduction, development, enemies, population dynamics, evolution and much more. Keys to genera for each biogeographic region, along with a guide to the literature for species identification, a world checklist of species, and a chapter on methods will allow anyone to embark on the study of these flies. A critical review of the risks and results of attempts to use sciomyzids for the control of snails that are pests or are hosts of pathogenic helminths and a brief history of the study of these flies completes this extensively illustrated review.

While the extent of our current knowledge of these flies falls short of a medically important family such as mosquitoes, this is one of the fullest accounts of the biology of any family of flies yet to be published, and this transformation has occurred in the last half century. The book is destined to be the springboard for the next half century of research on these attractive and intriguing flies.

Henry Disney

Note: The book briefly refers to other flies whose larvae feed on molluscs, whether dead or alive. This includes Phoridae. The authors not only missed some references subsequent to my 1994 review of what was known about Phoridae but unfortunately they list *Megaselia tertia* (Brues) larvae as feeders on dead snails. *M. tertia* was synonymised with *M. fisheri* (Malloch) in the 1960s and the latter is the correct name for this fly (as cited in my book).

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Meetings

Reports

Autumn Field Meeting

Worthing & Eastbourne

8-15 October 2011

This meeting was originally advertised for the North Pennines and Cumbria, but a combination of factors led to a switch of location. Firstly, I could not find sufficient accommodation at sensible prices in either Barnard Castle or in the Egremont and Grange-Over Sands areas. Several fruitless days work led to the need for a re-think. Furthermore, the long-range weather forecast suggested that rain was expected; it is 'The North' and my abiding memory of Cumbria when I did teaching practice there was that it was always raining or snowing! The decision to change venues proved to be a wise move. On day one it was raining in Cumbria and sunny in Sussex (our fingers were crossed that it would stay that way and as luck would have it – it did).

I try to find closely approximated guest houses for the party to simplify gathering for supper and daytime events. This time I found accommodation for almost all the group in one guest house in Worthing – apart from Malcolm & Mary Smart + Mara: suitable dog-friendly accommodation is harder to find. Unfortunately, I was outposted at a separate guest house and upon arrival was told that I was being moved to an alternative venue several streets away on the Monday. To say I was not impressed might have been an understatement that was not improved by the electrics in my room blowing five minutes after my arrival. Needless to say I would not recommend the Marine Guest House! Meanwhile, Sea Lodge Guest House where the main party was billeted was excellent and is to be recommended.

Worthing is nicely placed for woodlands and our first day involved a relatively short foray into local woods, mainly on Chalk. Conditions were very dry and there were few fungi about, so I feared the gnatting would not be good. I was amazed therefore that the first day's haul comprised 82 species together with three species of Platypezid caught by Malcolm Smart, while the scarce species *Agathomyia woodella* was found by both Ken and Peter in different parts of Angmering Park woods. At Fontwell Wood Alan caught several of the drosophilid *Hirtodrosophila trivittata*, a recent addition to the British list that develops in oyster mushrooms and is becoming widespread in the south, although it was not found again during this week. Cumbria, meanwhile, was still being rained on!

Our second day involved a longer journey, firstly to Burton and Chingford Ponds, a Sussex Wildlife Trust reserve that had been very productive during the summer field meeting at Plumpton in 2006. There were plenty of gnatty things about and the 39 species recorded included a recent addition to the British list *Phronia forcipula*. My eyesight is failing so I simply hoover everything that moves – creating a mass of struggling flies in the pooter that was mainly noteworthy for the numbers of mosquitoes. The haul here included 72 male and 46 female *Limonia nubeculosa*! Thank goodness for the Merrifield MkX pooter, which has the power to intercept flies in flight.

The party dispersed after this locality and visited Ebernoe Common and The Mens separately. Ebernoe Common was particularly productive and I was pleased to present Peter Chandler with a fine hillock of gnats. Two areas of Ebernoe Common were visited. Peter, Andrew Halstead and Ken & Rita Merrifield secured a very respectable 55 species from Willand Wood. Alan and I visited a northern area and obviously struck different habitat as the overall total from two areas visited by different parties at Ebernoe was 71 species – a respectable haul. Five species of platypezids including *Paraplatypeza bicincta* were also recorded. The Mens produced 40 species of gnat.

Assembly at Petworth park L-R: Ken Merrifield, Chris Spilling, Matthew Oates, Malcolm Smart, Peter Chandler. Photo: RogerMorris

Assembling on the third morning it emerged that we had taken the gnat list to 116 and there was a backlog developing. Bearing in mind how dry the woods were, and how few fungi were seen, this was amazing. Petworth Park was our main venue on the third day. Here, we joined up with Matthew Oates who many readers will know from his contributions to broadcasts on purple emperor butterflies. The park did not look particularly productive with little ivy and heavily grazed grasslands, but amazingly my net was filled with dancing flies – lots of gnats. Working over fallen timber and along walls produced a fair haul that was replicated by Andrew Halstead, Chris Spilling and Alan Stubbs (who also did moderately well for craneflies beside a lake). Peter fared less well, due to venturing into the more exposed areas, but the group as a whole generated 54 species of gnat. After leaving Petworth Park, the party went to a lake at Woolbeding House. Finally, we visited a pond on Woolbeding Common at which 38 species of gnats were recorded. Neither Woolbeding sites was frightfully inspiring but as the day was drawing to a close we might have had more success earlier in the day.

Day 4 involved a change of venue, from Worthing to Eastbourne via Amberley Wildbrooks and other sites en-route. Amberley is a lovely grazing marsh with what seem to be nice ditches. It might be appealing in the summer but this autumn it was unproductive and the *Salix* woodland was singularly unimpressive. I got nets full of mosquitoes but little else! Andrew and Peter, meanwhile, managed to fill their wellies with sludge due to the very soft ground. However, the drier fringes of this carr produced 16 species of gnats, including the most south-easterly record yet by 2km of the polypore-feeding *Ditomyia fasciata* (caught by Chris Spilling) that is surprisingly unrecorded from Kent, and the uncommon wetland species *Rymosia britteni*.

Our second proposed venue was a site near Brighton but it proved a big challenge to find. Frantic telephone messages finally assembled part of the group at Chalk Hill; which proved to be the local dog-emptying zone. This very dry woodland eventually produced 35 species of gnats. By the time we arrived at Eastbourne I was

convinced that Sussex has too many dogs! Needless to say, this day did little to improve the lists but the gnat list had proceeded above 140 and was starting to look respectable.

Eastbourne is nicely placed for a variety of habitats, especially chalk grasslands, which are not terribly productive in the autumn.

Admiring the geology at Birling Gap: Photo Ken Merrifield

However, our first call at Birling Gap (day 5) proved very productive. A small ash/ sycamore woodland yielded a huge haul of gnats for Alan and Peter: it was the only cover for miles around and the gnats fled there for cover from the wind! Remarkably, the list for this site reached 48 species, though more than half of the gnats sampled were *Brevicornu griseicolle*. It produced the uncommon *Mycetophila* species, *M. caudata* and *M. deflexa*, as well as two recent additions to the British list *M. sigmoides* and *M. sublunata*, the latter only first recorded as British on this year's summer meeting in Devon. The first three mentioned also turned up at other sites during the week. My attention here was focussed on the geology of the Gap itself. This is a key geological site that is established in planning case law because residents had tried to force the National Trust to defend the former Coastguard cottages and other properties that it owns there. NT policy is to allow coastal processes to continue and to accept loss of property. Thankfully the inspectors recognised the value of the geology, which attracts numerous school groups as we saw on the day of our visit – 3 school parties were there looking at the 'head' deposits.

Coastal erosion and "head" deposits at Birling Gap: Photo Ken Merrifield

After Birling Gap we explored Friston Forest where the combined total for the two areas visited was 51 species of gnats. Notably, one-upmanship developed with two members being overheard discussing the size of their gnats: 'my gnats are bigger than yours'. We then headed to Cradle Hill which overlooks the Cuckmere Valley. This too is an important coastal location because a partnership between the National Trust, the Environment Agency, East Sussex County Council and Natural England had hoped to re-establish tidal influences over the seaward end of the valley, but had been stopped by objections from a small group of residents living above the floodplain who objected to the creation of unsightly mud! Our vantage point at Cradle Hill offered great views of this valley and some para gliders – just out of reach of our nets! The hawthorn scrub was heavily ivy-covered and there were plenty of bees in attendance, including a few *Colletes hederae*.

By day 6 (Friday) the gnat list had reached 155 and together with 41 craneflies. We had one full day's recording to push the lists even further, but by now it was clear that Sussex had been extremely productive for gnats. This last day proved to be exceptional.

Selwyn's Wood (1). Now what has Malcolm been up to? Mirth and frivolity at Selwyn's Wood. Photo: RogerMorris

Selwyn's Wood (2) R-L: Rita Merrifield, Ken Merrifield, Malcolm Smart, Alan Stubbs & Peter Chandler. Photo: RogerMorris

Our first venue on the Friday was Selwyn's Wood, a Sussex Wildlife Trust reserve that was difficult to find because the entrance was obscure; thankfully we all made it. As with the rest of the week conditions were lovely and sunny but as usual there were

lots of brambles. By this time I had come up with a new definition of woodland: a large bramble patch with pits, hollows and the odd tree, all of which conspire to impede the gnat-hunter! Notwithstanding the obstacles this site produced the remarkable total of 92 species of gnats which is the highest ever number of gnats recorded for a site on a single visit.

Park Wood L-R Alan Stubbs, Andrew Halstead, Malcolm Smart & Peter Chandler. Photo: RogerMorris

My definition of woodland was confounded by Park Wood, our second venue. This is a fantastic woodland; probably the best of the week from an old woodland perspective. It comprises oak standards with hazel and hornbeam coppice, on clay. What was more pleasing was the relative lack of brambles! It also contains a fantastic dry gnat stream with overhangs and clouds of gnats dancing in the sunlight, together with lots of trichocerids and *Limonia nubeculosa*. I had a big job sorting the catch – which I frequently refer to as 'black grot' but am reminded that it is hardly an appropriate term for interesting Diptera that have died in the name of science! The gnat total reached a fantastic 80 species but was dwarfed only by the Selwyn's Wood haul. Malcolm Smart, meanwhile, took a very late female *Rhingia rostrata* and Andrew Halstead added the second site of the week for *Agathomyia woodella*. The final visit of the day for most was Abbot's Wood, where the dry woodland produced 44 species of gnats including *Keroplatus testaceus*, which had already been found at five other sites during the week. With three productive sites visited in the day, the combined gnat total was 127 species, boosting the week's records substantially.

Alan at The Crumbles looking for the remaining insect.

Crumomyia nitidia (Sphaerocerid) from Park Wood (Chris Spilling)

My party returned via Sovereign Harbour, which was once 'The Crumbles', a major shingle structure that was destroyed in the late 1990s in preference to a modern yacht harbour and associated development. This piece of coast is a testimony to the desperate need for the Habitats Directive in the late 1980s. What was once one of the rarest habitats in Europe (vegetated coastal shingle) is now an abomination of modern buildings not far different from what has happened to the Mediterranean coast of Spain!

The meeting came to an all-too sudden end, with members dispersing on the Saturday morning. But, we had reached a total of at least 200 gnats (plus a *Boletina* of uncertain identity) and had good hauls of heleomyzids too. It was also noteworthy that the snail-killing fly (Sciomyzidae) *Pherbellia scutellaris* was obtained for an unusually high proportion of sites in W Sussex. Platypezids were not numerous during the week but totalled 7 species (*Callomyia amoena*, *Agathomyia unicolor*, *A.. woodella*, *Protoclythia modesta*, *P. rufa*, *Platypeza consobrina* and *Paraplatypeza bicincta*) – this was lower than some meetings such as the highly successful trip to the Isle of Wight in 2005. Drosophilids were well recorded, with at least 14 species. Craneflies (41-43 species) on the other hand, were well down on historic levels and several other families were noticeably absent. Sepsids in particular were virtually absent and this suggests a potentially worrying trend. Remarkably few flies were seen at ivy despite the sun, especially the muscid *Mesembrina meridiana* and scathophagids; insects attending the flowers were often restricted to a few social wasps. And what about the weather in Cumbria? Well the rain ended mid-week so we might have managed a couple of days field work had we gone there!

Part of the group (PC, AS, AH & RM) gathered for a final morning's gnatting at the RSPB's newly acquired Broadwater Warren reserve. Despite lovely morning sunshine it was cold and the gnats took a long while to mobilise. Thus, by midday they were just moving as the party finally dispersed. This is a strong reminder that autumnal entomology is heavily temperature-related and one needs to think carefully about aspect when choosing a site to record. Peter continued until mid afternoon with a visit to Hoth Wood, where there was actually a wet muddy path and deep gnatty gulleys, providing records of 54 species.

As a general reflection on autumn meetings, field work is pretty well over by 15.30 so it is generally only possible to do two sites properly in a day; that suits me nicely as the autumn field meeting is an excellent way of finishing the field year. I rarely generate much

more than 175 records directly but have evolved into a reasonably efficient gnat hunter so I act as a parataxonomist to aid Peter and Alan's recording schemes. We take our time getting out in the morning and are back by a sensible time in the afternoon. What is more, all you have to do is hoover and sort – the end result is a really impressive list of flies. I'm totally hooked on this meeting and would make it my priority if I had to choose between the meetings on the year's calendar. The field year comes to an end with happy memories of golden woodlands, a happy band of dipterists, good food and nice landscapes – it is definitely my favourite!

This meeting was extremely useful because Eastbourne proved to be a really nice venue and looks suitable for a spring meeting. Both Guest Houses (Cambridge House and Sea View) were excellent and to be recommended. The nearby pub (The Marine) was also good for its food, although it was a little expensive. I may re-organise the proposed venues for field meetings to make possible a meeting here in May 2013.

Participants:

Peter Chandler, Andrew Halstead, Ken & Rita Merrifield, Roger Morris, Matthew Oates, Malcolm & Mary Smart (+ Mara), Chris Spilling, Alan Stubbs.

ANNUAL MEETING

Saturday 26th November 2011

Dipterists' Day 2011 at the Manchester Museum

People began to assemble at the Museum from about 9.30 am. Coffee was available so those not involved in preparing for the day could talk, or view the exhibits, and the entomological books displayed by Ian Johnson and his wife, from Pemberley Books.

A start was made at 10.30, when our Indoor Meetings Secretary and Chief Organiser for the day, began his opening presentation of information necessary to survive the day. No sooner had Malcolm mentioned the word 'fire alarm', than with almost perfect timing the fire alarm went off, and so we all left the building. The many conversations that had started before the meeting, continued until we were told that it was safe to return. Apparently a smoke alarm had gone off in a kitchen at the other end of the building. We began again at 10.55 and Malcolm handed over to Dmitri Logunov, the Keeper of Zoology at Manchester Museum who welcomed us to the Manchester Museum and its entomological collections. For an excellent account of the Diptera Collections see the excellent article by Dmitri in Bulletin 70, Autumn 2010. He would welcome the use of the collection by any interested dipterists. There are still some 4,500 specimens to be identified, and plenty of data to be gathered.

Time Flies - Diptera in Amber, David Penney

The first talk was given David Penney and entitled 'Time Flies -Diptera in Amber'. We learnt that about 98% of the arthropods in Amber are Diptera and that the resin which traps them takes about 2 million years to change first to copal and then to amber. Amber is found all over the world and Flies from the Lower Cretaceous epoch, 135-125 Ma, have been found in Lebanon, and on the Isle of Wight. 800 species have been found in Baltic Amber from the Eocene (ca. 45 Ma) and, in all, about 5,000 species are known. The most stunning pictures shown by David were those using X-ray tomography to produce a 3D picture of very high resolution, while the specimen remained embedded and invisible to light rays of longer wavelength. So many behavioural events are frozen in time, parasitic nematodes emerging, phoresy by mites and pseudoscorpions, even egg-laying, so that a picture can be built of the lives and palaeohabitats of these fossil flies.

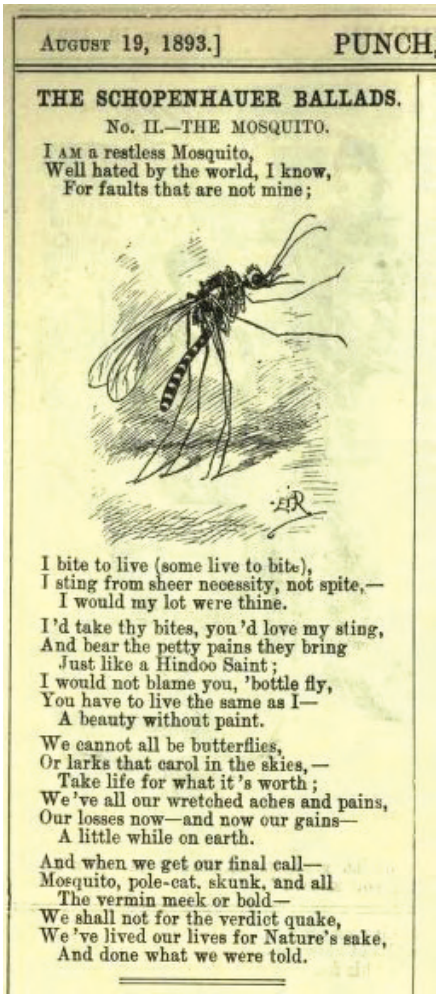
Good samples of ancient DNA have yet to be consistently be extracted from Diptera in amber.

(Account by JK)

After one or two questions it was time for a coffee, or to resume conversations.

Mosquitoes & their Recording Scheme, Jolyon Medlock

We reconvened and at 12.10, Jolyon Medlock told us about 'Mosquitoes & their Recording Scheme'. Jolyon told us something of the taxonomy of the Culicidae and said that there were some 33 species recorded in Britain, the females being the more easy to identify. Being blood-feeders they are very important vectors of diseases such as West Nile virus dengue fever and malaria, and it is probable that this will increase in the future. It is therefore very important that we learn as much as possible about the distribution and ecology of the Culicidae. In addition to his talk, Jolyon, with Erica McAlister, gave us a very useful workshop on the mosquitoes on Sunday morning at the museum.



August 19, 1893.] PUNCH,

THE SCHOPENHAUER BALLADS.

No. II.—THE MOSQUITO.

I am a restless Mosquito,
Well hated by the world, I know,
For faults that are not mine;

I bite to live (some live to bite),
I sting from sheer necessity, not spite,—
I would my lot were thine.

I'd take thy bites, you'd love my sting,
And bear the petty pains they bring
Just like a Hindoo Saint;
I would not blame you, 'bottle fly,
You have to live the same as I—
A beauty without paint.

We cannot all be butterflies,
Or larks that carol in the skies,—
Take life for what it 's worth;
We've all our wretched aches and pains,
Our losses now—and now our gains—
A little while on earth.

And when we get our final call—
Mosquito, pole-cat, skunk, and all
The vermin meek or bold—
We shall not for the verdict quake,
We've lived our lives for Nature's sake,
And done what we were told.

We now have a mapping scheme linked to the NBN Gateway project. So get involved and send in your data. (see Bulletin cover cover for contact details).

Since, due to the fire alarm, the programme was running late, Malcolm re-organised the talks so that our next speakers, and our final ones before lunch, were students **Peter Wing** and **Nathan Medd**. Peter has an interest in flies of medical or veterinary importance but particularly Oestridae, Calliphoridae and Sarcophagidae. Peter's short talk looked at the question 'Does larval diet influence adult body mass in flesh-eating flies?'

In order to answer this question Peter measured parts of the exoskeleton to produce a quantitative index for larval growth of these maggots, which can be compared for different larval diets.

Nathan Medd then gave a talk on The diurnal activity patterns of British Hoverflies. Nathan's study area is in Hampshire and he related flight periods to the air temperature. From his data he hypothesized that smaller flies were active earlier, ie. at a lower air temperature.

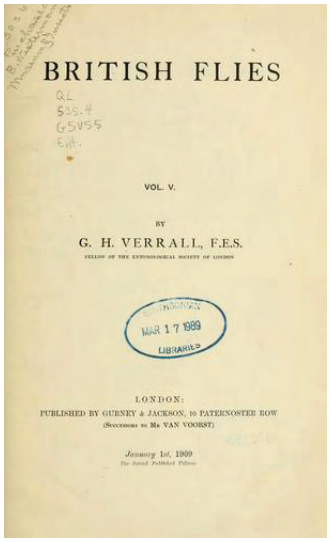
It being about 1.10 pm, we then broke up for lunch.There is an excellent café at the museum which sells plenty of good things to eat.

We reassembled at 1.45am and for the formal part of the meeting. For a report on the proceeds of the AGM, please read the Minutes in this Bulletin. At about 2.45 pm the programme of talks then continued

The next talk, **The G. H. Verrall story,** a centennial appreciation, was given by Adrian Pont. George Henry Verrall died one hundred years ago, on 16 September 1911, and so it was important that the Dipterists Forum marked this occasion. The full text of the talk will be published in the Dipterists Digest and the summary below is based on the text by Adrian.

The G. H. Verrall story, Adrian Pont

Adrian decribed the work done by Verrall under four headings: his researches and publications on the Diptera; the impetus he gave to dipterology in this country; the collection of Diptera that he formed, and finally, his employment of his nephew J. E. Collin as his amanuensis and successor in dipterology.



Verrall was born on 7 February 1848, the youngest of seven children, in the Sussex town of Lewes. He had an interest in natural history from an early age, and entomology took hold when he was about 14 years old and from the age of 18 he focussed entirely on the Diptera. He collected all over Britain and the Verrall-Collin collection which he founded is without doubt the most comprehensive and important collection of British Diptera ever formed. Adrian described Verrall's social background and activities before describing his work in dipterology.

Verrall described 6 genus-group names and 77 species-group names. At the time, his pre-eminent reputation rested on his series of papers on Syrphidae in the 1870s, Tipulidae in the 1880s, and Dolichopodidae in the 1870s and again in the 1900s, his two check lists of British Diptera of 1888 and 1901. He also wrote the two monumental volumes of *British Flies* of 1901 and 1909.The first, in 1901, was designated as volume 8 and dealt with the Platypezidae, Pipunculidae and Syrphidae. It also included taxonomic catalogues of all the European species of these families. The second volume, in 1909, was designated as volume 5 and dealt with the families of the so-called lower Brachycera, from the Stratiomyidae onwards. These are embellished with beautiful line drawings of structural details and even of whole flies by J. E. Collin.

Adrian briefly summarised Verrall's achievements as follows:

- his publications, especially his monographs and the two check lists
- his collection, including the invaluable collections of Kowarz and Bigot (and also including his list of the contents of the Bigot collection)
- the annual gathering of entomologists known as the Verrall Supper
- the purchase of Wicken Fen
- and, finally, his nephew James Edward Collin
- and, of course, his vast and detailed knowledge of the Diptera which he was happy to share through prolific correspondence with fellow-dipterists at home and abroad.

iFly ?

Our afternoon coffee break was then taken. The next speaker at 3.40pm was Keith Alexander who gave a talk about Diptera and decaying wood. Keith has generously provided a summary of his talk, below.

Talking rot: Deadwood for Dipterists, by Keith Alexander – A Summary

English Nature Research Report No. 467 (Alexander 2002) lists 730 GB Diptera species as dependent on decaying wood (saproxylics); these are from 68 families. 33% of the species have conservation status, either RDB (143) or Nationally Scarce (99). This is therefore a major assemblage of Diptera and one of considerable conservation importance. And yet we don't really understand the ecology of most of them; they develop in association with the decaying wood of trees in a wide variety of situations – woodlands, wood pastures, parklands, orchards, hedgerows – favouring open-grown trees and/or close-grown trees.

As individual trees develop and age, the quantities, types and locations of decaying wood vary. Open-grown trees provide the most varied habitats as they are able to grow old and retrench, whereas close-grown trees die young due to canopy competition. The two main types of heartwood decay are white rot, which removes lignin and leaves the white cellulose, (caused by bracket fungi such as *Ganoderma*, *Inonotus*, etc) and red or brown rot, where the cellulose is removed, (caused by *Laetiporus sulphureus*, *Phaeolus schweinittzii*, etc); Diptera tend to favour the former but a few develop in the latter. Aerial dead branches on living trees are a specialist habitat for Diptera. Rot-hole formation requires branch loss and so is associated with open-grown trees rather than close-grown trees – the main bracket fungus causing rot-holes is *Polyporus squamosus*.

Recent recording using flight interception traps has begun to demonstrate just how many of the rarer saproxylic flies are associated with open-grown trees. Even in a large Irish woodland, the richest

catch of fungus gnats came from the opened up compartments rather than the shady areas. A great advantage of trapping is that sampling takes place over 24 hours, as opposed to the daytime visits by dipterists with nets. Flight traps can be used in a much more targeted way than Malaise traps.

What determines the faunal composition of saproxylic flies at a particular site? The key 'four dimensions' are: age structure of the trees (decay succession); density of the trees (open-grown v close-grown); how many trees (viability); ecological continuity (site history). Advice provided to land managers about the habitat requirements of flies, for conservation, should aim to cover these aspects, as far as possible: eg open-grown trees required to encourage rot-hole formation, the most important trees are 100 years old or more, the species needs need x trees per xx hectares, etc, etc. It is no use just saying it is a woodland species of fly and needs deadwood. This is woefully inadequate!

Keith Alexander

International Symposium on the Syrphidae, Stuart Ball

At about 4.15pm Stuart Ball told us about some of the highlights of the 6th International Symposium on the Syrphidae held at Glasgow Aug 2011. Stuart's own paper was about the changing pattern of recorder behaviour – something which has a crucial bearing on the interpretation of data from the recording schemes, where a constant sampling effort would be ideal. Stuart described some statistical methods for taking account of sampling variations. Recruitment and training are the keys to the obtaining of good quality data and the Hoverfly Recording Scheme have led the way with this.

Malcolm thanked the final speaker, and closed the Afternoon Session. After receiving our instructions regarding the most suitable pub in which to assemble, and the location of the Sangam Indian Restaurant for our Dipterists' Supper, we escaped from the building just before the doors closed at 5.00pm. The Sangam provided about 20 members with a very tasty and enjoyable meal.

On Sunday morning, 27th November about 20 members again assembled at the Museum for a workshop on the Mosquitoes, led by Jolyon Medlock, and assisted by Erica McAlister. We were introduced to the most useful keys, and the most useful features to use for their identification. Erica then demonstrated the best way to mount these fragile flies using card points. (The prize for the most desirable gadget was awarded, by Erica, to the EntoBall, with which anything is possible.) Collecting methods were next, and how to breed, slide-mount and examine the larvae. The morning concluded with talk about the invasive species to look out for in the UK as climate-warming continues, causing the northward migration of populations from southern Europe.

Some of us also spent a useful hour, during the morning, in the collections room, at the Manchester Museum, examining some of the flies stored there. It is well worth a re-visit.

John Kramer

Minutes of the Annual General Meeting

Manchester Museum at 2.00pm on Saturday 26th November 2011.

Chair: Martin Drake. About 46 members were present.

1. Apologies were received from Judy Webb, Barbara Ismay, Roger Morris, and by e.mail Simon Hayhow, and John O'Sullivan.

2. Minutes.

The minutes of the AGM held at the Oxford University Museum of Natural History, at 2.00pm on Saturday 27th November 2010 and published in the Spring Bulletin 2011, were accepted unanimously by the meeting as a correct record.

Matters arising.

There being no matters arising, the Chairman introduced the Secretary's Report.

3. Secretary's Report to the AGM:

I am pleased to tell you that we continue to flourish and the current membership of the Forum is now 396 (2010 - 382) with 346 subscribers to the Dipterists Digest (2010 - 340). With some 30 renewals still pending, the target of 400 members should be reached by the end of this year. Thanks to our Publicity Officer and all the others who have helped to represent the Forum at public events, but special thanks are due this year to Mick Parker, our retiring Membership Secretary, for all of his efforts over the past 8 years, to recruit new members and to retain our current subscribers.

a) The Recording Schemes

In addition to recording, and making records available, the 17 Recording Schemes and Study Groups are also proactive in encouraging the development of identification skills through the workshops and through the availability of up-to-date, user-friendly keys.

Newsletters have been produced during the year by the Hoverfly RS (Nos. 50&51) Cranefly RS (No.21&22) Fungus Gnats RS (No.5), Tephritid RS, and Empidids & Dollies RS (No. 16).

Can I, as usual, urge anyone not currently involved in a Recording Scheme or Study Group to make contact with a scheme organiser. There are plenty of interesting things to enjoy, and plenty of people willing to help you.

b) Committee Meetings

The Committee met on three occasions throughout the year; 5 March, 6 July and 12 November 2011. Much of our effort has gone toward maintaining a full and successful programme of events, details of which can be read in the Bulletins Nos. 71 & 72, and the Newsletters of Recording Schemes.

c) Activities

i) Field Meetings: It is an aim of the Forum to record the distribution of Diptera throughout the UK and a major contribution to this are the Field Meetings. During 2011, the 3 main meetings were: Spring at Abergavenny, 12-15 May 2011, Summer at Exeter University, 2-9 July 2011, and the Autumn meeting at Worthing & Eastbourne, from 8-16 Oct.

Our thanks are due to Roger Morris who has done an enormous amount on behalf of us all. Not only has he done all of the organisation for the Field Meetings, sorting out suitable accommodation,

obtaining permits to visit sites, and writing up the reports for the Bulletin, but he has entered the data collected from all post 2000 field meetings onto the Recorder database. This has consisted of over 68,000 records in all, a landmark event, so that they are now available for Landowners, and the NBN Gateway. Can I also draw your attention to Roger's detailed report on the Records from 1987 – 2011, on the last pages of this year's Autumn Bulletin, No. 72.

Another important area of Roger's work, and also in the Bulletin, has been to increase our awareness of the need for Safety at Field Meetings. It is something that we all need to think about while doing our own field work, perhaps on our own in the middle of a grimpen mire, or pushing through the bracken in a deer forest.

ii) Indoor Meetings:

DF Identification Workshops, 4 – 6 March, 2011, at Preston Montford.

The Beginners Workshop was an 'Introduction to Fly Families' (led by Stuart Ball & John Ismay), and the Advanced Worrkshop was 'Fungus-feeding Flies' (led by Peter Chandler & Judy Webb.)

Discussion has started in sub-committee about the presentation of the **Beginners Workshops** on a more local scale, following the success of Roger Morris and Stuart Ball with Courses and Workshops - for the Hoverfly Recording Scheme.

The OPAL grants, successfully bid for by Stuart and Roger, have enabled us to purchase sufficient microscopes and illuminators for the Forum to run its own courses at a variety of local venues. The equipment has been put to excellent use by them and has resulted in the recruitment of many new members to the DF. Our approach to workshops and courses, and the training of course leaders, is under review and we hope to make much more progress in this area.

If you have opinions about the costs of Workshops, or possible venues in your district, please e.mail me, or let another member of the committee know.

The DF, through the Hoverfly Recording Scheme, was pleased to support the **International Syrphid Symposium** which was held at The Hunterian Museum, Glasgow in August 2011. Stuart will say more about this later, as part of the day's programme.

d) Publications

The stock of printed **Starter Packs (previously given to new members)** is now exhausted but it is now available as a pdf to everyone via the website. The role of the Starter Pack has been to a large extent, taken over by the excellent new edition of **A Dipterist's Handbook,** edited by Peter Chandler which was presented to the forum at the Annual Meeting last year, and there are still copies available for purchase.

e) Publicity and Recruitment

As previously announced, Judy, our Assistant Bulletin Editor and our Publicity Officer, who has done so much to make the Forum more widely known and to increase the membership, is unable to be with us today and she writes to the meeting as follows:

I'm finding that I am short of time for the publicity work and therefore need to retire from the Publicity Officer post at the 2012 AGM in order to concentrate on my role on the editorial panel for the Bulletin.

I also feel that recruitment to the DF would benefit from the input of a younger person who would be engaged with modern social media and would therefore be better able to interest younger people in the Diptera.

If you feel that you can help by continuing this important work, please let a member of the committee know.

We do our best to support new members, and we are offering mentoring to new members**.** New members may be offered a mentor in their region, or, failing that, mentor-support for them via e.mail, if they so wish.

Alas, our hopes to interest more people in Diptera through a Photography Competition have sadly come to nothing, and very few entries were submitted so that the competition proposed for this year has had to be cancelled. Discussions are continuing and a more popular way forward may be found via the Website.

f) DF Website

The Forum website continues to be a very effective way for us to reach people. The Committee has discussed a number of problems which have occurred and it is hoped to re-design and develop the site much more in the future. Our website manager is a very busy person and the chief problem is a shortage of time.

Progress is being made on the scanning of past publications in order to create searchable pdfs so that eventually all of the past publications, Digests, Bulletins and Newsletters will be available on the Website.

g) Conservation Issues

It is through the conservation of species and habitats that we make our records count.

Barbara Ismay had previously announced at the 2010 AGM that she will resign from the post of Conservation Officer as from today, so thank you Barbara for all of your hard work. Martin will say more about this later. Unfortunately Barbara is not able to be with us today but has written to thank those who have contributed to the BAP species project and the 'Adopt-a-species' scheme. Barbara has led on the 'Adopt-a-species' scheme, with a view to improving our knowledge of data-deficient species and anyone interested in helping with this project, perhaps by studying a rare fly at a site near their home, should discuss their interest with her.

We are fortunate that Rob Wolton has expressed his willingness to continue this important work and Barbara wished him well in his future role.

h) Future Meetings

i)Next Indoor:

Spring Workshops at Preston Montford – Fri 2nd Sun. 4th March 2012

The Beginners Workshops – Introduction to Diptera–Led by Roger Morris and Alan Stubbs

Advanced Workshop: Dolichopodid Flies, Led by Martin Drake, with assistance from Roy Crossley and Alan Stubbs

ii) Next Field Meetings:

New Forest – 12-13 May 2012

Lagganalia Centre, Kingussie, Speyside. 22-29 July 2012

iii) The next AGM will be held at Bristol Museum and the probable date, contingent on other entomological meetings, is Sat Nov 24th 2012

John Kramer Secretary

4. Treasurer's Report – Howard Bentley

Copies of the audited accounts for the calendar year 2010 were distributed, and the treasurer apologised for having failed to publish them in the Bulletin. He expressed his thanks to Tony Pickles and Mr. A. S. Harmer for once again acting as auditors without payment. Tony has kindly agreed to continue his work for the Forum next year. We are neither a company nor a charity, so the production of audited accounts is not a legal

necessity; nevertheless we consider them important and intend to continue the present practice of producing them annually. The main sources of our income continue to be subscriptions for membership of the Forum and for the Dipterists Digest, and our main expenditures remain the printing and distribution of the Digest and the Bulletin. In 2010 the former brought in £6002 and the latter cost us £6084. These remain broadly in balance, having a slight surplus in some years and a slight deficit in others. Discrepancies with membership figures arise because some members pay in advance for their membership, and some in arrears, though most now pay by bank transfers early in the year. In future the accounts are likely to be more complicated, as field courses are now being financed through the Forum's bank accounts. Formerly Roger Morris, our field meetings secretary, ran these through his personal finances, but changes in his circumstances mean that this is no longer possible. Payments for accommodation, and income from members attending, are rarely finalised within one calendar year, and this will make interpretation of our accounts more difficult. The treasurer intends to produce a balance sheet for each course as and when its financial arrangements are completed. As mentioned by other speakers OPAL grants were obtained in 2010 and 2011. The 2010 accounts appear to show a surplus remaining from the 2010 grant, but this has in fact been spent this year. We usually finish each year with a small surplus, but costs are rising. So far we have been able to keep our printing costs down by judicious "shopping around", but this cannot continue indefinitely. Although we have no immediate plans for an increase in our very modest subscription rates, we are keeping these under constant review.

Howard Bentley Treasurer

5. Dipterists Digest Editor's Report

Two issues were produced during the year, as the second part of volume 17 was delayed until February and Volume 18 part one appeared at the end of June. I am pleased to report that no problems have been experienced with the printing of these issues. We have continued to include a variety of colour plates, and colour printing of the cover illustration has now become a regular feature.

It was announced in the first of these issues that the second part of volume 18 would commemorate the centenary of the death of George Verrall, who died on 16 September 1911. Contributions relating to his work were invited and an account of his life based on the talk by Adrian Pont, to be given following the AGM, will be included. It is hoped that this issue will be ready to go to the printers in December, although distribution will not now be practicable until the new year.

At the same time sufficient material has been offered to fill the first part of volume 19, so we are in a good position at present from that point of view, although two long papers have helped to achieve this. It may also be possible to publish that issue before the end of January.

More short notes are needed to fill space where papers are an odd number of pages, and any contributions of papers and notes are always welcome. I am grateful to all those who have contributed, but would appreciate it if papers submitted could as far as possible follow the instructions to authors. The layout of recent issues should be consulted to assist with this. Much editorial time is taken up with changing A4 to A5, altering fonts and font sizes and changing references to the Digest format, which should have journal titles without abbreviation.

Reports of exhibits for the annual meetings have been incomplete in recent years and I would like to remedy that by providing more comprehensive coverage in the Digest of what is exhibited. Notes on exhibits, where possible as email attachments, are requested.

As promised in this year's Bulletins, the 1998 checklist has now been updated to include all additions and changes reported in the Digest. I thank Stuart for putting a pdf of this update on the Forum website. I also thank Mike Pugh for proof reading and Roy Crossley for distribution.

Peter Chandler Dipterists Digest Editor

6. Any other business

There was none.

7. Chairman's Vote of Thanks to retiring members

Martin thanked Mick Parker on his retirement from the post of Membership Secretary. When Mick took over in 2003 the membership stood at 271. It is probable that without his efforts the membership would not have increased to the level that it is today. Mick represented the Forum every year at AES and BENHS exhibitions and worked assiduously to maintain our existing list of subscribers.

Barbara Ismay was also warmly thanked for all of her hard work, of a very high quality, on the BAP species, during her time as Conservation Officer. A vote of thanks was proposed to both retiring Officers that was supported unanimously by the meeting.

8. Election of Officers: See details below

The Chairman proposed that the Officers listed below be elected en bloc and this was seconded by Alan Stubbs and accepted unanimously.

The Officers and General Committee to be elected, or re-elected this year, 2011 were as follows:

Office	Officer
Chair	Martin Drake (Elected 2010)
Vice Chair	Stuart Ball
Secretary	John Kramer
Treasurer	Howard Bentley
Membership Secretary	John Showers (Proposed)
Field Meetings Secretary	Roger Morris
Indoor Meetings Secretary	Malcolm Smart
Bulletin Editor	Darwyn Sumner
Assistant Editor and Publicity Officer	Judy Webb
Website Manager	Stuart Ball
Conservation/BAP Officer	Robert Wolton (Proposed)

Committee Members

1. Chris Spilling (Proposed re-election)
2. Erica McAlister (Proposed re-election)
3. Duncan Sivell (Proposed election)
4. Barbara Ismay (Proposed re-election)
5. Mick Parker (Proposed re-election)
6. John Ismay (Elected 2010)

Post 6 was elected in 2010 and is therefore due to stand for re-election in 2012.

9.Chairman: Thanks to hosts and formal closing of the Annual General Meeting.

There being no other business, Martin brought the AGM to a close at 2.40pm. He thanked Dmiti Logunov, Curator of Arthropods, our host at the Museum, and Malcolm Smart, out Indoor Meetings' Secretary, for all of their hard work to make the meeting so successful and enjoyable.

John Kramer Secretary

Forthcoming

Diptera Identification Workshops 2012

Preston Montford Field Studies Centre
Friday 2th - Sunday 4th March 2012

Beginner's Workshop – Introduction to Diptera

Led by Roger Morris and Alan Stubbs

Arrive Friday in time for supper at 6.30pm - depart 4.00pm Sunday.

This is an introductory course on the Identification of Fly Families. It is designed to help people getting started with identification and recording of this fascinating group of insects which are very varied in their behaviour and can be found in nearly all habitats. They can also be used in the assessment of the quality of many different types of habitat.

The course is aimed at absolute beginners and will guide them through many hurdles, both as a group and as individuals. Each attendee gets individual help and will work using a microscope on their own individual set of specially prepared flies which are examples of the major Dipteran families found in the UK. A set of keys has been specially produced for this course! Each attendee leaves with their own set of keys plus advice on how to collect and pin flies for identification and for retention as voucher specimens.

All materials and equipment (microscopes, lights etc.) will be supplied by the Field centre.

If you are new to the delights of Diptera study and feel the need for a formal introduction, this is the workshop for you! If you know anyone who has an interest in flies but needs to know more, it's also just the thing for them too!

Advanced workshop – Dolichopodid Flies

Led by Martin Drake with assistance from Roy Crossley and Alan Stubbs

Arrive Friday in time for supper at 6.30pm – depart Sunday afternoon.

Elegance, poise and charm are not terms one would normally use to describe flies, but there is scarcely a nondescript species among the Dolichopodidae. They have long been studied by British dipterists, having caught the attention of G.H. Verrall whose early publications undoubtedly started the interest in the 19th century. There are now just over 300 species in the British Isles, and the rate of discovery of new species suggests that there are quite a few more to find. Within the Empid and Dolichopodid recording scheme, the Dolichopodidae have recently been given a higher profile; so the time is now right to run a workshop to stimulate more interest, sort out the problems with the published keys and provide some new test keys.

Most Dolichopodidae are associated with wetter habitats, especially seepages, fens, water margins and wet woodlands. A few are found on barnacle-covered rocks on the coast, and there are suites associated with tree trunks, canopy foliage and dry grasslands. The habitat affinities of wetland species have been studied in Europe so we know more about their ecology than you might expect for a group of fairly small flies, and a review of the rarity status in Britain was published in 2005. The combination of up-to-date statuses and good understanding of their habitat requirements makes the family among the more useful ones for assessing the value of wetlands.

Fancy legs and occasionally marked wings are used by the males in courtship. This behaviour can be seen easily in some common species, for instance *Poecilobothrus* at garden ponds and puddles, and *Chrysotus* buzzing females like the more familiar *Eristalis nemorum* hoverflies. A wide field of studies is open to keen observers here. The early stages are also under-worked and only a small proportion has been described. Nearly all are predators with the exception of leaf-mining *Thrypticus*.

The workshop is aimed at those who have some experience with smaller flies. It will concentrate on identifying adults using existing keys and some new ones where the RES Handbook (d'Assis Fonseca, 1978) causes problems. About 30 species have been added since the Handbook and recognising these will be covered. An introductory talk will discuss the natural history of the family.

Reference specimens will be provided but please bring any that you have collected yourself. If you have your own microscope and lamp, then please bring them along. The centre does have some, so don't feel that you cannot attend if you don't have them.

Fees & Booking Procedure for either workshop

Dipterists Forum members:

Single Room Resident: £168 full board accommodation
Shared Room Resident: £148 full board accommodation
Non-resident: £90 incl. packed lunches & evening meals

Non Dipterists Forum members (fees include one year's DF membership):

Single Room Resident: £248 full board accommodation
Shared Room Resident: £228 full board accommodation
Non-resident: £170 incl. packed lunches & evening meals

To book a place on either of these workshops please contact:

Preston Montford Field Centre, Montford Bridge, Shrewsbury, SY4 1DX
Tel: 01743 852040 Fax: 01743 851066
Email: enquiries.pm@field-studies-council.org

You will be requested to complete a booking form and to pay the full course fee in advance. **Please make sure that you note that you indicate you are a DF member on the booking form in order to secure your members discount**

Organiser: Malcolm Smart
malcolmsmart@talktalk.net

Spring Field Meetings

New Forest
12-13 May 2012

This meeting will provide an opportunity for members to visit several of the Inclosures and bogs around the New Forest. Members are asked to contact Roger Morris for assembly details. Those members wishing to stay are asked to make their own arrangements. Roger's party will probably be based in Bournemouth and it is anticipated that a group will form around this locality. It may be possible for early bookers to find accommodation in a common venue – which Roger will organise if he is given advance warning in March.

Roger Morris

Perivale Wood, Middlesex
19 May 2012

Also a request for subsequent recording on the site

This site at Greenford, Middlesex is the property of the Selborne Society and is the second oldest nature reserve in Britain. It overlaps the 1km squares TQ1583 and TQ1683 and comprises 11 hectares of ancient woodland, old permanent neutral pasture and a small pond. The pond, which is about 110 years old, has a shallow inflow for 6-8 months of the year from a nearby hay meadow country park and borders a 0.5 hectare area of carr woodland, which is wet throughout the year. The reserve does not have public access so can only be visited by arrangement.

We have been asked to visit by Peter Edwards, who has been involved with the management of the site for many years. There has been no thorough entomological assessment of the site and it is considered to be under-recorded for Diptera. It has, however, been visited by several dipterists over the years and in particular Ken and Rita Merrifield have recorded 186 species of Diptera on occasional visits to the wood over the past 15 years.

The Selborne Society is particularly interested in determining whether there is any dipterological interest in the pond and is willing to fund expenses incurred by dipterists who are able to make subsequent visits for this purpose spread over the year. This meeting will be an opportunity for anyone who is likely to be able to make such visits to assess the area's potential and familiarise themselves with the site.

John Kramer will be recording craneflies on this meeting, which will also be attended by Ken & Rita Merrifield and myself. We expect to meet at the locked gate to the reserve at 10.30a.m. The gate is at TQ163836 and situated between 36 and 38 Sunley Gardens.

If you would like to come please let me know (chandgnats@aol.com, 01225708339). If there is poor weather on this date a visit will be arranged for a later date.

Peter Chandler

The Great Fen Project
16th - 17th June 2012

The Great Fen project is an initiative aimed at re-wetting a large area of former farmland between Woodwalton and Holme Fens. Our meeting will provide an opportunity to visit both these fens and also to explore the re-developing fen. It is hoped that a meeting at this venue will become a regular event as Dipterists Forum is encouraged to participate in documenting the changes that occur over the coming decades. Our meeting is an inaugural and exploratory event. We have arranged that our lunch will be taken in the Rothschild bungalow at Woodwalton Fen. Please contact Roger Morris for final assembly details

Roger Morris

Bridlington for soft rock cliffs and chalk wolds
29 June – 1 July 2012

This is a special meeting of Dipterists Forum and Yorkshire dipterists and provides a rare opportunity to explore the soft cliffs and seepages of the coast between Flamborough Head and Scarborough. We also hope to explore a range of slumping cliff systems and nearby chalk grasslands on the Yorkshire Wolds. The meeting will be centred upon Bridlington which offers suitable B&B accommodation at sensible prices. Participants are asked to contact Roger Morris who will supply a list of possible guest houses – late participants will be expected to book their own accommodation, although Roger will book places for those who contact him in March.

Roger Morris

Summer Field Meeting

Lagganalia Centre, Kingussie, Speyside
22-29 July 2012

We have reserved accommodation for 20 members – ten in single rooms and a further ten in twin rooms. At the time of writing (December) all of the single rooms have been booked and vacant spaces remain for seven participants in shared rooms. If you want a room, it might be worth choosing your room-mate so make contact with Roger to find out who wants a room. It may be possible to arrange additional accommodation in nearby Kingussie, or if there are sufficient members interested in another lodge at Lagganalia.

This fantastic opportunity will allow us to explore the Spey valley at a slightly later time than we have previously visited. It is a wonderful part of the world where one barely has to leave ones doorstep before encountering the specialities of this very special valley. If you have not visited it is well worth the effort.

Roger Morris

Autumn Field Meeting

Bangor, North Wales
13-20 October 2012

This meeting provides an excellent opportunity to visit the woodlands and coastline of the North Wales mainland and the dune systems of Anglesey. It will be based in the University town of Bangor for the majority of the week, although consideration will be given to being based in Llandudno for part of the time.

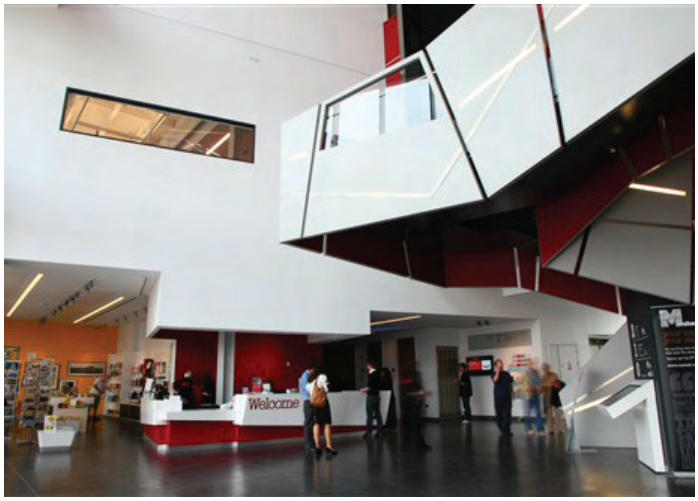
The autumn field meeting is a great chance to relax with friends at a more leisurely pace. Those of us who do groups that are most active in the summer act as parataxonomists and feed specimens to Alan Stubbs and Peter Chandler. It is quite the most excellent way of winding down for winter whilst making a real contribution to the cranefly and fungus gnat recording schemes.

Roger Morris

Dipterists Day and AGM 2012

Bristol, Princes Wharf
24 November 2012

This is scheduled to be held in the new Mshed Museum at Princes Wharf, Wapping Road, Bristol BS1 4RN (http://mshed.org/about-us/) on 24 November 2012 with access to the entomological collections on Sunday 25 November.



Full details of the programme will be posted on the DF website (http://www.dipteristsforum.org.uk/) when available and will be formally announced in the Autumn 2012 DF Bulletin (no. 74)

Organiser: Malcolm Smart
malcolmsmart@talktalk.net

8th International Congress of Dipterology 2014

10-15 August 2014

The 8th International Congress of Dipterology will be held at Potsdam near Berlin, Germany, on 10–15 August 2014. The organizing committee consists of Marion KOTRBA, Netta DORCHIN, Fritz GELLER-GRIMM, Frank MENZEL and Joachim ZIEGLER.

Only about 30 minutes from the bustling German capital with its museums, shops, theatres and pubs, the small town of Potsdam harbours famous tourist attractions such as Frederick the Great's famous Sanssouci Palace with its beautiful park area, Filmpark Babelsberg, the Biosphere, and the cozy streets of the "Dutch Quarter".

The congress will take place at the Kongresshotel Potsdam, once the construction site of Count Zeppelin's famous airships and today a modern and spacious building beautifully situated on the banks of Lake Templiner. The hotel offers accommodation for all participants as well as all scientific sessions under one roof, with ample space for relaxation, socializing, and discussions during coffee breaks. There is also the restaurant / beer garden "Seekrug" immediately next to it. Check out these locations at 52°22'22"N 13°00'54"E.

The scientific programme will include broad-scope sessions covering large taxonomic groups (e. g., Nematocera, Orthorrhapha, Eremoneura, Calyptrata, acalyptrate families) as well as general topics (e. g., higher level Phylogeny, Morphology/Anatomy/Ultrastructure, Physiology, Biodiversity, Conservation, Evolutionary Biology, Behaviour, Diptera of Economic Importance, Diptera in Forensic Entomology, Diptera in Medical Entomology, and Fossil Diptera).

We plan to combine the congress with special public outreach activities, the most important of which is the award-winning exhibition "Flies" ("Fliegen – Mouches") originally created by Christophe DUFOUR and Jean-Paul HAENNI (Neuchâtel). The exhibition will be presented at the Museum of Natural History in Berlin during the congress. Our congress logo is largely based on the design of this exhibition. We also plan other activities, such as a fly-related art exhibition, promoting a fly for "insect of the year", a celebration of the 250th anniversary of the birth of the founder of European dipterology Johann Wilhelm MEIGEN (1764–1845), a Diptera postage stamp, public talks, press conferences, etc.

We have already secured substantial financial support from the Senckenberg Foundation (as can be seen from the logo) and will apply for additional funds from other scientific foundations and sponsors.

With ample input from the dipterological community worldwide this will become yet another wonderful congress. Keep up-to-date as our new website (www.icd8.info) takes shape and do not hesitate to contact us with ideas, wishes and suggestions.

Dr Marion KOTRBA (Munich)
e-mail: marion.kotrba@zsm.mwn.de

Events Calendar 2012

Dipterists Forum & selected meetings

Check the Dipterists Forum website for changes and meetings added after publication of this Bulletin, www.dipteristsforum.org.uk)

18-19 February 2012, BENHS Regional Meeting at the FSC Preston Montford, Shropshire. 'Invertebrate Challenge' (part of Shropshire Entomology Day on Sat 18 Feb.). Identification workshops for various insect groups on Sun 19th. Contact Pete Boardman, (pete@field-studies-council.org) or see : www.benhs.org.uk

2-4 March 2012, DF Identification Workshops. Beginner's workshop on 'Introduction to Diptera', Advanced Workshop on 'Dolichopodid Flies'. Preston Montford Field Studies Centre, Shrewsbury. Details in this issue and posted on the DF website and on FSC website: http://www.field-studies-council.org/prestonmontford/

10 - 11 March 2012, Hoverfly Identification Course. Yarner Wood, Woodland Classroom, Devon. Costs : £30~40 depending on number of participants (12 maximum). For further details or to secure a place contact: mattprince1969@gmail.com

17 March 2012, BENHS AGM and Presidential Address plus talks, tours and discussions. University Museum of Natural History, Parks Road, Oxford OX1 3PW. See: www.benhs.org.uk

28 April 2012, AES Members' Day and AGM 2012. The Manchester Museum, The University of Manchester, Oxford Road, Manchester, M13 9PL, UK. http://www.amentsoc.org

10 May 2012, Insect-Fungus Interactions, .Insect Ecology Special Interest **Group Meeting.** Rothamsted Research, Harpenden, Herts, AL5 2JQ. Contact **Michael Bonsall: michael.bonsall@zoo.ox.ac.uk**

12 May 2012, BENHS Regional Meeting at Elvedon, Thetford, Norfolk 'Breckland Invertebrates'. Contact: Claudia Watts (CWatts@royalparks.gsi.gov.uk) or See: www.benhs.org.uk

12-13 May 2012, DF Spring Field Meeting to the New Forest, (to be confirmed).. Contact Roger Morris (7 Vine Street, Stamford, Lincolnshire, roger.morris@dsl.pipex.com

19 May 2012, Perivale Wood, Middlesex. One day Field Meeting to record Diptera for the Selbourne Society in this site with ancient woodland pasture and pond. Contact Peter Chandler (chandgnats@aol.com, tel: 01225708339)

15-17 June 2012, Identifying Flies Course. Flatford Mill Field Centre, Suffolk. Tutor Martin Harvey. Contact: enquiries.fm@field-studies-council.org

16-17 June 2012 The Great Fen Project The Great Fen project is an initiative aimed at re-wetting a large area of former farmland between Woodwalton and Holme Fens. Our meeting will provide an opportunity to visit both these fens and also to explore the re-developing fen. Contact Roger Morris (7 Vine Street, Stamford, Lincolnshire, roger.morris@dsl.pipex.com

29 June – 1 July 2012, Bridlington for soft rock cliffs and chalk wolds. A special meeting of Dipterists Forum and Yorkshire Dipterists. Contact Roger Morris (7 Vine Street, Stamford, Lincolnshire, roger.morris@dsl.pipex.com

25 June - 1 July 2012, National Insect Week. See website: http://www.nationalinsectweek.co.uk

18-20 July 2012, Ento'12 - the National Science Meeting of the RES, Venue: Anglia Ruskin University, Cambridge. See http://www.royensoc.co.uk

22 - 29 July 2012, DF Summer Field Meeting 2012 Lagganalia Centre, Kingussie, Speyside. See this issue, early booking recommended. Contact Roger Morris (7 Vine Street, Stamford, Lincolnshire, roger.morris@dsl.pipex.com

17-20 August 2012, Identification of Hoverflies. Preston Montford Field Centre, Shrewsbury. Tutors Stuart Ball and Roger Morris. See FSC website: http://www.field-studies-council.org/prestonmontford/

13-20 October 2012, DF Autumn Field Meeting to Bangor, North Wales Contact Roger Morris (7 Vine Street, Stamford, Lincolnshire, roger.morris@dsl.pipex.com

3 November 2012, BENHS Annual Exhibition. Imperial College, London See: www.benhs.org.uk

24 November 2012 Dipterists Day and AGM 2012, Bristol. M Shed Museum at Princes Wharf, Wapping Road, Bristol BS1 4RN (http://mshed.org/about-us/) with access to the entomological collections on Sunday 25 November. Full details of the programme will be posted on the DF website (http://www.dipteristsforum.org.uk/) when available.

BENHS Dinton Pastures Open Days in the Pelham-Clinton Building, Hurst, Reading. Open 10:30-16:00 on second and fourth Sunday in each month except April to September when only on the second Sunday of each month (except for August when there are no Open Days). We encourage you to bring along your pinned flies and use the Diptera Collections and library for identification. Other Dipterists are usually present meaning good chat and assistance with identifications may be possible. The grid reference for Dinton Pastures is SU 784718, turn left off the B3030 driving North from Winnersh. The site is about 15 minutes walk from Winnersh station, which has trains running on a half-hourly service from Reading and Waterloo. See: www.benhs.org.uk

April-Sept/Oct 2012 The Northants and Peterborough Diptera Group hold meetings every weekend from end of April until some time in September/October. Contact John Showers on: ShowersJohn@aol.com

Hoverfly Newsletter
Number 52
Spring 2012
ISSN 1358-5029

2011 may have been a year when hoverflies were often especially hard to find in numbers, but, as the many articles below will testify, it was certainly a vintage year for Syrphidology in several other ways. The Glasgow Symposium was a truly memorable event, and the year saw the publication of **The Natural History of Hoverflies** by Graham Rotheray and Francis Gilbert and the new atlas, and as we enter 2012 we have the start of the Big Hoverfly Watch, the Wild*Guide* and a new edition of British Hoverflies to look forward to.

It has been a pleasure to receive so many articles in a timely manner without the need for me to chase authors or try to solicit contributions. My thanks to you all. Please try to keep up this momentum!

Articles and illustrations (including colour images) for the next newsletter are always welcome. Copy for **Hoverfly Newsletter No. 53** (which is expected to be issued with the Autumn 2012 Dipterists Forum Bulletin) should be sent to me: David Iliff, **Green Willows, Station Road, Woodmancote, Cheltenham, Glos, GL52 9HN, (telephone 01242 674398), email:davidiliff@talk21.com**, to reach me by 20 June 2012.

The hoverfly illustrated at the top right of this page is a female *Leucozona glaucia*.

Ron Payne: a tribute

Ted Levy
9 Chilton Grove, Yeovil, Somerset, BA21 4AN

I want to pay tribute to Ron; his obituary appeared in the Autumn edition of the Bulletin of the Dipterists Forum. He was entirely responsible for my early interest in hoverflies, which has been our obsession ever since. Originally I had been interested on Lepidoptera, but was also a keen birdwatcher prior to 1971. When Ron moved to Westcliff-on-Sea, he joined the South Essex Natural History Society, based in Southend, and as a committee member, I felt it appropriate to welcome him by tentatively collecting hoverflies locally, and showing an interest! Some of my specimens were scarce or interesting, so my enthusiasm grew, and we had several sessions at his home, determining and seeing his vast collection, which occupied shelves all around his study! He was always most kind and patient, and lent me identification keys and guides which were most helpful!

As at that time I was a sub-editor of The South Essex Naturalist Journal, which we were upgrading from news sheets to a better publication. I received several articles from Ron about hoverflies in our region. I also joined him when he led a coach trip of dipterists to Lakenheath in Suffolk in June 1971, and it was at this time that I began recording and collecting hoverflies in earnest.

When Ron moved to Somerset we lost touch, except by brief correspondence, but when we began our research for Somerset Hoverflies he helped with advice and records, though by then he had more interest in grasses. When we became interested in finding *Eristalis cryptarum* on Dartmoor he proved to have collected widely there, and had a specimen of that species. Later during book research we were permitted access to his collection in Bristol Museum and species were added to the Somerset list.

Dave and I were fortunate to meet Ron Payne and we have forty years of interest in hoverflies to thank him for, though probably his enthusiasm must also have rubbed off on many dipterists in his lifetime.

6th International Symposium on the Syrphidae, Hunterian Museum, Glasgow, 5-8 August 2011

Roger Morris
7 Vine Street, Stamford, Lincolnshire PE9 1QE

The first International Symposium on the Syrphidae was held in Stuttgart in 2001 and was a tremendous success. It attracted a broad spectrum of delegates from across the world and there were at least ten UK delegates. Since then, numbers of UK delegates have declined to a bare handful: just four went to the last symposium at Novi-Sad (Serbia) in 2009. Yet, hoverflies are a very popular group of insects that boast one of the most active recording schemes and are the subject of several UK research themes. It was therefore greatly encouraging that UK attendance at this latest symposium involved at least 22 delegates, several of whom provided a presentation or a poster.

The conference ran over three days and was split into six sessions with a total of 41 presentations listed in the abstracts volume. In addition, at least 27 posters were presented. Unfortunately several delegates, including Dieter Doczkal and Valeri Mutin, were unable to attend in the end and so the list was somewhat shorter. There is often a silver lining to such clouds and on this occasion it allowed Nathan Medd and Kirsten Miller, MSc Students at Imperial College, to tell us more about their work on hoverfly activity patterns and aspects of the ecology of *Microdon myrmicae*. For me, these were obvious highlights to the conference because they showed that a new generation of hoverfly enthusiasts might just be developing. Hopefully we will hear much more from both Nathan and Kirsten in coming years.

A wide spectrum of issues was covered, ranging from autecological studies to morphometric analyses and genetic studies that throw important light on the taxonomy and classification of particular genera. Studies of hoverflies in Latin America are clearly gaining momentum and it was interesting to hear of work in Brazil, Columbia and Puerto Rico. For me, the following highlights represent the parts of the conference that most grabbed my attention:

Ellen Rotheray ***Restoring endangered hoverflies: Case study of the pine* Blera fallax *and aspen* Hammerschmidtia ferruginea *hoverflies in Scotland***. Ellen's work has thrown considerable light on the ecology of both species and gives reason to hope that conservation measures for both species will lead to maintenance of more robust and sustainable populations. Remarkably, it appears that *Callicera rufa* and *Blera fallax* larvae vacate water-filled cavities in the winter to avoid being entombed in ice, whereas *Myathropa florea* remain *in-situ* and suffer major losses through freezing. Most noteworthy, however, was the degree to which *H. ferruginea* was demonstrated to disperse – at least 5 km. in one case. Translocation of *B. fallax* also provides encouraging signs that populations are capable of dispersal and there is now evidence of a new breeding site established about a kilometre away from the initial introduction site.

Rob Wolton ***Adult and larval behaviour of the ant-eating hoverfly,* Microdon myrmicae**. Rob regaled us with his studies of the population of *M. myrmicae* on his farm in North Devon. Rob has managed to observe most of the stages in this hoverfly's life cycle. Eggs are laid in batches of up to three at the mouth of the nests of the ant *Myrmica scabrinodis* (and possibly other species). First instar larvae are postulated to feed on buccal pellets and other detritus within the nest but subsequent instars are predacious upon the ant larvae. Adults show little sexual selection or courtship. Males have been demonstrated to live for up to 18 days in the wild, whilst females have lasted 20 days in captivity. Rob also raised the possibility of a new genus – Mogodon (referring to a sleeping pill) – coined by his family to describe their response to his constantly talking about this fly!

Maarten de Groot ***The effect of altitude on species composition and seasonal dynamics in hoverflies in beech forest***. Using a mixture of netting and malaise trapping, Maarten demonstrated substantial changes in the peaks of abundance and species richness on the north side of Mount Krim in Slovenia. Although the results were consistent with what might be expected, they nicely illustrated the differences in timing of hoverfly species richness and abundance. For me, the most striking point was that there were multiple peaks in abundance at the various points up the mountain with peaks in May, July and August, with multiple peaks at lower altitudes and less pronounced peaks at higher altitudes.

Menno Reemer & Gunilla Stahls ***Phylogeny and classification of the Microdontinae***. For me, the most striking part of this presentation was the remarkable range in form amongst the Microdontinae that highlighted the extreme uniformity in the European fauna and extensive diversification in the tropics. So far, the DNA of 80 of the 400 species of Microdontinae has been analysed. A significant proportion of the described species are known from the type specimens only, suggesting that there is

considerable scope for adding to our knowledge of these remarkable flies.

Gunilla Stahlls *et al.* MtDNA *CO1 haplotype distribution patterns in the eastern Aegean area (Greece)*. This study focussed on the genus *Merodon* which is one of the most dominant genera in the eastern Mediterranean. The larvae are associated with a variety of 'geophytes' or bulb-forming plants. Separation of the European and Asian continental plates during the Miocene means that populations of certain *Merodon* species might be expected to have been isolated over differing timeframes and signals for these differences were sought in the Mitochondrial DNA CO1 gene. Evidence from a small suite of islands showed that these differences could be detected in some but not all species.

Catalina Gutierrez-Chacon & Padu Franco *Syrphids in the coffee-growing region of the Columbian Andes: occurrence in relation to landscape context*. This study investigated the highly crenulated landscape of the Columbian Andes with coffee plantations on steep slopes that are capped by relatively undamaged forest. Given the intensity of sampling, the numbers of hoverflies recorded was remarkably low – a total of 896 specimens from 88 hours netting, 2,856 hrs. malaise trapping and 960 hrs. of van-Someren-Reydon traps. This effort yielded 79 species from 19 genera. Results pointed to an increase in species-richness as the landscape contained more forest, but some genera seemed to occur exclusively in the coffee plantations.

John Smit ***A survey of the hoverflies of the Lagua Blanca Natural Reserve in Paraguay***. John's talk explored a survey of three habitats, wet Atlantic forest, dry forest and the extremely hot Cerrado (a thermophilic scrubby habitat). His talk was noteworthy for the extremely low numbers of hoverflies recorded; at times equating to about one specimen per hour of effort! His conclusion was that Paraguay was not worth visiting for hoverflies (I formed a similar view for some other parts of South America after getting better results but still poor numbers). The results also conformed to the findings in Columbia where malaise trapping was found to be a relatively poor method of recording hoverflies in the tropics.

Several taxonomic studies were relevant to the UK fauna. Work on *Dasysyrphus* by Michelle Locke on the Nearctic fauna (some of which is Holarctic) suggests that we will see several important splits in *D. venustus*; a situation previously reported from work by Dieter Doczkal. We also learned from Zorica Nedeljkovic that *Chrysotoxum festivum* comprises two species: a paler yellower northern species that occurs in Scandinavia and seems to be represented in British collections; and a darker species that appears to be confined to the Mediterranean. Finally, and most importantly, work on the genus *Pipiza* was presented by Ante Vujic, Hans Bartsch, Rune Bygebjerg and Gunilla Stahlls. In this work it seems that there are no obvious new species and that *Pipiza bimaculata* and *P. fenestrata* have new names; a key to the European fauna was provided in a poster. We eagerly await the publication of these changes.

The split between professional and non-vocational UK participants was marginally weighted towards the non-vocational component. Overseas participation was primarily from academic institutions. A strong contingent from the Netherlands, Germany and Scandinavia was augmented by teams from Novi-Sad (Serbia) and Spain (Alicante). Delegates from further afield, including Canada, Columbia, Brazil, Russia, Romania, the Czech Republic and Ukraine, contributed to the overall complement of over 70 delegates.

The Williston Diptera Research Fund and four UK societies (the British Entomological & Natural History Society (BENHS), Dipterists Forum, Glasgow Natural History Society and the Malloch Society) provided financial support for the Symposium. This made it possible for us to assist seven overseas delegates with part of their costs: Catalina Gutiérrez Chacón (Columbia), Dr Pavel Laska (Czech Republic), Dr Grigory Popov (Ukraine), Augusto Montoya (Puerto Rico), Mirian Morales (Brazil), Dr Martin Speight (Rep. of Ireland) and Dr Carmen Stanescu (Romania). We were also able to subsidise student attendance and to provide conference literature including a special edition of the new atlas of British Hoverflies. This atlas, jointly authored by Stuart Ball, Roger Morris, Graham Rotheray and Kenneth Watt is the first to combine all UK data, including those data held by the Scottish Hoverfly Mapping Project; it is a huge improvement on the atlas produced in 2000 and is based on almost double the number of records (745,000).

Unfortunately, although we approached all of the UK professional entomological and ecological societies for sponsorship, only one actually responded and informed us that it would not be able to help; the others simply did not answer! We were amazed to get no response from the Royal Entomological Society (RESL) and pursued this both through their on-line communication system and through a direct letter to the President. The former yielded no response, whilst the latter led to initial contact from

Professor Stuart Reynolds, but we heard no more from the Society itself.

The fact that four amateur societies felt it appropriate to support the conference and the RESL chose not to respond speaks volumes for the nature of entomological outreach in the UK, and this despite four of the five organisers being Fellows of the RESL. Clearly, hoverflies are regarded as an 'amateur' pursuit even though they attract significant funding under pollinator programmes across the world.

Work by the HRS to train new hoverfly recorders is clearly paying dividends, and it was immensely gratifying to meet up with several alumni of HRS training courses. Three presented posters and Rob Wolton presented his impressive work on *Microdon myrmicae*. We hope that by the time this symposium returns to the UK there will be many more alumni of the *Introduction to hoverflies* course. Training new syrphidologists was part of the theme we developed for Recording Scheme presentations. Our presentations explored some aspects of our experience and evolution of teaching techniques, and looked at some of the trends in hoverfly recording and its implications for data analysis.

Organising this event was a major undertaking and it has drawn upon the efforts of five organisers: Stuart Ball, Francis Gilbert, Geoff Hancock, Roger Morris and Graham Rotheray. The organisers were greatly assisted by the sponsoring organisations and would like to give special thanks to all five sponsors.

Offers from Canada and Russia to host the Seventh Symposium were put to the audience and it was concluded that the next symposium would be at Novisibirsk in 2013. This venue in Siberia is difficult to reach and the meeting will therefore be followed by an extended field trip to the Altai Mountains (we heard about these from John Smit in 2007). Anybody intending to go should make sure that they get an invitation from Anatolii Barkalov in good time to make it possible to apply for a Russian visa. Initial investigations of flights suggest that there are no direct services and that at least one and possibly two changes are required. Flight costs are difficult to judge but it looks as though there will be little change from £1,500. Start saving now!

The post symposium trip visited Rowardennan Research Station. This is a fantastic place that lies on a wooded peninsular on the eastern shore of Loch Lomond. Hoverflies were sparse but, with over 30 hoverfly specialists working this area, a respectable list was compiled. The list has still to be completed but so far 47 species have been reported. We tend to take our fauna for granted and it was therefore noteworthy that the Dutch contingent was pleased to see *Leucozona glaucia*, which is extinct in the Netherlands.

Species recorded from Rowardennan: *Baccha elongata*, *Cheilosia antiqua*, *C. bergenstammi*, *C. fraterna*, *C. longula*, *C .scutellata*, *C. vernalis*, *Chrysogaster solstitialis*, *Chrysotoxum arcuatum*, *C. bicinctum*, *Dasysyrphus albostriatus*, *Didea fasciata*, *Epistrophe grossulariae*, *Episyrphus balteatus*, *Eriozona syrphoides*, *Eristalis nemorum*, *E. pertinax*, *Ferdinandea cuprea*, *Ferdinandea ruficornis*, *Helophilus pendulus*, *Leucozona glaucia*, *L. lucorum*, *Melangyna compositarum*, *Melanostoma mellinum*, *M. scalare*, *Meliscaeva auricollis*, *M. cinctella*, *Myathropa florea*, *Neoascia podagrica*, *Orthonevra nobilis*, *Platycheirus albimanus*, *P. clypeatus*, *P. fulviventris*, *P. nielseni*, *P. occultus*, *P. peltatus*, *Scaeva selenitica*, *Sericomyia silentis*, *Sphegina clunipes*, *S. elegans*, *S. siberica*, *Syritta pipiens*, *Syrphus ribesii*, *S. torvus*, *S. vitripennis*, *Xylota jakutorum*, *X. segnis*

Hoverfly Recording Scheme update

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What a bumper year for hoverfly enthusiasts. A new atlas has been produced; the 6th International Symposium on the Syrphidae was held in Glasgow in August; we have delivered the text and illustrations for the forthcoming WILD*Guide* '*Britain's Hoverflies*' to the publishers; and we have finalised the statuses of hoverflies listed in the National Status Review published by JNCC. At the moment we are working on a revision of Alan Stubbs and Steven Falk's '*British Hoverflies*', which we hope will emerge in 2012. What is more, we have also run numerous training events to recruit new hoverfly recorders in the past year and will be running further courses in Glasgow, Bristol, Exeter, London and Bangor this winter. Many of these venues are dependent upon us providing the microscopes and we are very fortunate to have secured an OPAL grant to buy 13 microscopes for running courses at venues that are not normally equipped

with microscopes. This is a great advance and we are keen to take bookings for the winter of 2012-2013.

Production of a new atlas is just one part of the Recording Scheme. It was generated from a dataset of three quarters of a million hoverfly records – double the number that was available ten years ago for the first atlas. How many will there be in 2020 when we produce a more comprehensive volume? Do keep the records rolling in and watch out for opportunities to get involved as we are starting to embark on new initiatives. The one that is potentially very exciting is the start of 'Big Hover Watch' which we are modelling on the 'Big Garden Birdwatch' that the RSPB holds each year. The concept is new and consequently we want to recruit a small group of people to trial it in 2012 in advance of preparing literature and publicity for a bigger launch in 2013. If you are interested in taking part, please let Roger know – details are posted in this issue of the Newsletter.

We also hope to launch a new version of the website in 2012. Stuart has been hard at work developing a new format which we hope will improve our interface with hoverfly enthusiasts. In the meantime, the mapping facility on the existing website has been repaired and is now fully functional with up- to- date records available.

2011 was not the greatest year for hoverflies but it has yielded some exciting new records, most notably reports of *Callicera rufa* from Shropshire and Bedfordshire. These reports prompted a further one for 2009 from Nottinghamshire to be reported. What is going on with this species? It is rarely seen as an adult in Scotland, and there have been no indications of a gradual southward spread into northern England, so we seem to be looking at major jumps from an unknown source. However, there are potentially good reasons for this change and it seems likely that creation of new habitat as conifer plantations are felled plays some part in the process. We know from work by the Malloch Society that *C. rufa* is moving into felled plantations in Scotland, so why not elsewhere? Hopefully recorders will be sufficiently motivated to look for this species in pine woods elsewhere. Nigel Jones (2011) explained his discoveries in Shropshire in the last Bulletin. If you know of a hill top with exposed Scots Pine trees it is worth a look as *C. rufa* seems to be hill-topping. However, the other two sites are lowland localities with conifers and so it is entirely possible that this species will turn up in many more places.

We were also greatly excited by a possible *Syrphus admirandus* caught by Roger Morris at a site in Lincolnshire. In the end we have concluded that the specimen is not this species (thanks to Hans Bartch's excellent guide to the Swedish fauna (Bartsch, 2009). At the moment it must be logged as a very odd *Syrphus ribesii* but this seems highly unlikely. More work is needed to determine quite what it is.

Hoverfly atlas

Hopefully, by the time this newsletter reaches readers those who have made a significant contribution to the atlas over the past 10 years (50 or more records) will have received their copy via the Biological Records Centre. If it has not arrived, it should do soon after. Blame Roger for the delay – he has bitten off too much this year and has had a problem sorting out the address list.

Hoverfly WILD*Guide*

We seem to be regularly reporting delays, and again we have to report a delay. We finally delivered text and photos to WILD*Guides* in October and hoped that the book would come out in March or April 2012. Unfortunately that is not the case as the process of formatting our product has raised a number of issues. We are now revising our approach to identification to see whether we can come up with a format that meets the consistently high standards that WILD*Guides* prides itself on. The problem is how to produce a key that is not a key? We have tried several approaches and they all have their drawbacks. So, at the moment we think we may see the final product in July 2012. We are terribly sorry about the delays which really amount to us underestimating the work involved and the extra effort required once the draft was produced.

Data from websites

Over the past few years the numbers of photographers posting excellent photos of hoverflies on websites such as Flickr and WildaboutBritain has increased tremendously. Roger regularly trawls these sites and extracts usable data. The numbers seem to be rising exponentially and in the last 6 months some 1500 have been extracted. In all, it looks as though we will gather somewhere between 1500 and 2000 records per year from this medium, which amounts to perhaps as much as 10% of the yearly totals. It is quite amazing what people manage to find, but making a firm identification is far more challenging. If you post photos on the web and use a pseudonym, please can you let Roger know so that we don't generate multiple datasets for the same person.

In memoriam – Hans Bartsch

Many of our more enthusiastic readers will know of Hans Bartsch through his fantastic volumes on the hoverflies of Sweden. Sadly, Hans died of pneumonia in April this year. We had the great pleasure of his company at several of the Hoverfly symposia and have very happy memories of those times; he will be greatly missed for his infectious enthusiasm and kind nature.

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Jones, N., 2011. Astonishing discoveries of *Callicera rufa* in England. *Hoverfly Newsletter* **51**: 4-5.

Syrphus nitidifrons Becker (Diptera Syrphidae) – a second UK record

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I visited Denny Wood in the New Forest, Hampshire, between 31 May and 3 June 2011 to look for speciality hoverfly species.

On 1 June I caught a hoverfly in an opening along a path in broadleaved woodland near the edge of the wood and adjacent to heathland. The hoverfly was hovering at eye-height, and I expected it to be *Parasyrphus punctulatus*, of which I had already caught several in this opening, exhibiting the same behaviour. When confined in a tube, I determined that it was clearly not this species, although superficially similar, and indeed not any other species with which I was familiar.

The hoverfly, a female, was of a similar size to *P punctulatus*. The abdomen was black, with narrow paired orange spots on tergites two to four, the pair on tergite two being slightly broader. The sternites showed dark central spots. The face was yellow down to the base of the facial prominence, and below this the mouth edge was black. The frons was shining black without dusting.

Because of its overall similarity to *P punctulatus* I initially took this specimen through the *Parasyrphus* key of Stubbs and Falk 2002. It quickly ran to *Syrphus nitidifrons* due to the paired spots on tergites 2 and 3 and its black, shining frons. However, seeing that this was a species not yet discovered in the UK (at the time of publishing of Stubbs and Falk) I tried to work the hoverfly through other keys in this book (*Syrphus*, *Eupeodes*) but without any satisfactory conclusion.

I collected the specimen and subsequently passed it to the Bedfordshire County Hoverfly Recorder John O'Sullivan, who identified it as *Syrphus nitidifrons*, using Stubbs and Falk 2002, Van Veen 2004 and Parker 2010, and also having had the benefit of seeing the first UK specimen of this species at the BENHS Annual Exhibition in November 2010. This first specimen had been collected in Dorset on 10 May 2010 by Mick Parker (Parker 2010). The identity of the New Forest specimen was subsequently confirmed by Dr Martin Speight at the 6th International Symposium on the Syrphidae in Glasgow in August 2011.

This species occurs in parts of western Europe and now appears to be colonising the UK. The paucity of records may possibly be due at least in part to its arboreal habits and a flight period restricted between the months of April and June.

Acknowledgements
My thanks to John O'Sullivan for identification of the specimen, and for commenting on a draft of this paper. Thanks also to Martin Speight for confirmation of the identification.

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A further record of *Callicera rufa* Schummel, 1842, in Central England

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Following my report of the discovery of *Callicera rufa* in lowland England in the previous Hoverfly Newsletter (Jones, 2011), I received an email from John Szczur in Nottinghamshire. It turns out that John had found a single female *C. rufa* on the edge of the National Trust's Clumber Park on 31 May 2009, pushing back the English discovery date by two years. John had captured and determined the fly correctly, but in some disbelief that it could be *C. rufa*, he had

withheld from making the record known, in case it was an incorrect determination. John has emailed photos of the specimen to me and it is clearly *C. rufa.*

John's *Callicera* was taken from *Rhododendron ponticum* flowers, from which it was nectaring. The site is described by John as:

The principal habitats within 100m radius of the capture site are:
a) Grass Heath; established on a clear-felled conifer plantation with rotting cut stumps still evident in 2009.
b) Mature conifer plantations; composed, in the main, of Scots and Corsican pines, with clear-fell and restocked areas.
c) Mature mixed woodland; with numerous huge trees, including Scots Pine, many of which were originally growing in an arboretum/parkland setting but today are growing alongside a significant amount of regeneration.
d) A tree line of mature Yew.

We now know of four sites, in three counties, in central England (Bedfordshire, Nottinghamshire and Shropshire) for *C. rufa.* I noted in my previous report that it seemed unlikely that *C. rufa* would not be present in more areas, and this has already proved to be so! This new record adds to the conviction that *C. rufa* is likely to be quite widespread in England. The peak season would appear to be May, with the presence of adults lasting into late June. I'll repeat my previous call for dipterists to make a special effort during May to visit areas with cut conifers and mature or standing dead trees. The best place to search is probably at height on dead and live tree trunks, possibly targeting hill and ridge top areas with these features.

Good hunting!

Reference

Jones, N. 2011. Astonishing discoveries of *Callicera rufa* in England. Hoverfly Newsletter **51**. 4-5.

Pelecocera tricincta locally numerous

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Pelecocera tricincta is usually encountered singly though is reputed on occasion to be locally frequent. 31 August 2011 on Chamberlayne's Heath, Dorset, was just one such occasion. Along a length of gravel track through wet heath I noticed several and, finding that once I "got my eye in" they were quite easy to spot, I decided to do a quick count along a measured stretch of track between two readily identifiable and fairly permanent posts (with a view to perhaps repeating the exercise on future occasions). Because they were fairly sedentary there was little risk of double counting.

Over a distance of 103 metres I noted 23 individuals. All of these were on yellow flowers of, in descending order of frequency, *Hypochoeris radicata, Crepis capillaris, Potentilla erecta* and *Hypericum perforatum*. On several plants of the first two species I noticed that the flies seemed to be avidly feeding on pollen (see photo). Elsewhere on the heath I encountered a pair in cop. - again on a flower of *Hypochoeris* and, knowing where to concentrate my search, found them to be fairly widespread on this and other, neighbouring, heaths. However no other spot produced more than one or two individuals.

Pelecocera tricincta feeding on pollen
(photo: Ian Cross)

An early record of *Eristalis similis* from Britain

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Aside from generating a large dataset for the new Atlas, study of undetermined syrphids in the collection of the Norwich Castle Museum collection during 2011 produced a record of *Eristalis similis* (Fallen) in Britain well before its addition to the British list (as *Eristalis pratorum* (Meigen)) by Falk (1990).

A male of this species, taken in Norwich on 18 July 1942, was the star of a small and otherwise unremarkable collection of hoverflies made by R.M. Stuart Brown. No other details accompany the specimen save for the information that it was taken on hawthorn on a wet day. The specimen's accession number is NWHCM : 2000.150.43.

Raoul Stuart Brown was a student in London during the early 1940s, but his home was in Norwich. With the encouragement of Ted Ellis, Keeper of Natural History at the Museum at that time, he made a collection of insects of all orders. It was entirely fortuitous that he collected the *E. similis*, which remained unrecognised until the Museum's miscellaneous collections were examined critically in 2011. It is quite likely that earlier specimens may remain overlooked in other museum collections.

Since its addition to the British list from Warwickshire, *similis* has been recorded infrequently but widely with further records from Derbyshire, Yorkshire, East Sussex, Leicestershire and Somerset.

Reference Falk, S.J. 1990. *Eristalis pratorum* (Meigen,1822): a new British hoverfly. *British Journal of Entomology and Natural History*. **3**, 139-141.

R. M. Stuart Brown's *Eristalis similis* specimen (photo: Tony Irwin)

Some recent, and one not so recent, records from Buckinghamshire (VC 24)

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Cheilosia griseiventris: on the evening of 2 September 2011 I was running a mercury vapour light at Prestwood Picnic Site, a chalk grassland and scrub nature reserve near High Wycombe (SU866991). A *Cheilosia* appeared on the sheet under the light. I've never seen a *Cheilosia* at light before, and it seems likely that this one was disturbed from the adjacent vegetation rather than being attracted to the light. It proved to be *Cheilosia griseiventris*, apparently the first record for the county judging by the online recording scheme maps. It was swiftly followed by a second record the next day, from a disused railway line north-west of Aylesbury (SP709201), this time from flowers during the day (the line has developed a good mix of open, flower-rich turf and species-rich hedges).

Cheilosia nigripes: I have a *Cheilosia* specimen from 1995 that had sat in my collection over the name "*Cheilosia albitarsis s.l.*", and although I'd made a note that the legs seemed too dark for *albitarsis* I hadn't been able to find a better match. I recently sent the specimen to Roger Morris who has determined it as *Cheilosia nigripes*. According to the recording scheme online maps this is also a new record for the county. The specimen is a female and was collected from Homefield Wood Wildlife Trust reserve (SU812869) on 6 May 1995, in a woodland-edge chalk grassland meadow, typical habitat for *C. nigripes*. Many thanks to Roger for his help with this and other specimens.

Ferdinandea ruficornis: a species I hadn't encountered before, from the disused railway line north-west of Aylesbury (SP709201), swept from flower-rich vegetation on 24 June 2011.

Ferdinandea ruficornis male (photo: Nigel Jones)

Rhingia rostrata: after many years of checking the abdominal markings of *Rhingia campestris* in vain, I finally had my first encounter with *rostrata* on 5 September 2010 at a flowering Buddleia in the walled garden at Hughenden Manor (National Trust; SU861954). On 29 May 2011 another one was among brambles at the edge of the disused railway line north-west of Aylesbury (this time at SP711214).

Rhingia rostrata female (photo: David Iliff)

Spread of *Chrysotoxum verralli* into Gloucestershire

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Chrysotoxum verralli, previously unrecorded on the western side of England, has spread into Gloucestershire since 2006, and now appears to be well established there.

Of the eight British species of *Chrysotoxum*, five - *C. arcuatum*, *C. cautum*, *C. elegans* (typical form), *C. octomaculatum* and *C. verralli*, are somewhat difficult to distinguish from one another at sight; these are the species in which the yellow markings on the tergites predominate over the black, and which consequently are excellent mimics of the social wasps (*Vespula* sp.). Before the year 2006, only two of these five, *C. arcuatum* and *C. cautum*, had been recorded in Gloucestershire, *cautum* frequently and *arcuatum* represented by only a very small number of records from the far west of the county, where it is apparently at southern extremity of its range.

C. verralli was not known from Gloucestershire before 2006, and was considered to be species of the south and east of England – I had encountered it on visits to Essex. It was described by the Recording Scheme organisers as having suffered a substantial decline in the 1990s. The first Gloucestershire record was of two females at The Mythe, near Tewkesbury, on 25 August 2006, by Martin Matthews. It has since been recorded in the county in every subsequent year apart from 2008, with records from four sites in 2011(an otherwise very unproductive year for hoverfly recording in Gloucestershire). Even in 2008 it was probably observed: on two occasions that year I saw in my garden a *Chrysotoxum* of the wasp-mimic type that was smaller than typical *C. cautum*, but on both occasions it evaded capture or close observation. John Phillips and Martin Matthews reported similar sightings that year.

The full list of Gloucestershire records of *Chrysotoxum verralli* is as follows:
25/8/2006 The Mythe (SO8834) 2f Martin Matthews
11/7/2007 Woodmancote (SO9628) 1f David Iliff
17/8/2009 Pope's Hill (SO6841) 1f John Phillips
1/7/2010 Blakeney Straits (SO6508) 1f Maris Midgley
26/6/2010 Pope's Hill (SO6814) 1m John Phillips
28/6/2011 Prior's Park, Tewkesbury (SO8931) 1m David Iliff
30/6/2011 Prestbury Hill (SO9924) 1f David Iliff
16/7/2011 Hartpury Orchard Centre (SO7825) 1m Anthony Taylor, det. David Iliff

In the period 2009 to 2011 Gloucestershire records of *Chrysotoxum cautum* have been atypically sparse. Could this have any connection with the spread of *verralli*?

Male and female *Chrysotoxum verralli* in Gloucestershire in 2011 (photos: David Iliff)

The hoverflies of a Devon hedge

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Last year I decided to see how many species of plant and animal I could find in a single hedge on our farm in north Devon. It has become absorbing and exacting work, and an excellent way to improve my natural history skills. I'm hugely grateful to all those experts who are helping me with identification – without them I would still be struggling with the basics. My hope when I started was to demonstrate that the lowly hedge really is a habitat worth conserving in its own right - with 1300 species identified so far, the sheer diversity of life in the hedge I'm looking at has surpassed all my expectations and I think proves the point.

It's a fairly typical Devon hedge, about 90m long, running along one side of the farm lane. It has an earth bank down the centre, and on the lane side a shallow ditch with tall herbs, nettles, umbellifers and brambles, on the 2m margin between this ditch and the tarmac. On the other side a lightly grazed herb-rich margin grades into semi-improved pasture. The shrub layer is species-rich, with hazel, blackthorn, hawthorn and grey willow predominating. The hedge is well connected to other habitats, with a thin strip of streamside woodland at the bottom, a similar hedge on the other side of the lane, and a small farm pond close by. The farm has Soil Association organic registration.

As may be expected the majority of species recorded are insects, with the three big orders being Lepidoptera, Diptera and Hymenoptera. The Malaise trap I've been loaned picks up a considerable diversity of parasitic wasps, but even so I think the flies are likely to lead the way in terms of species richness. And of these, the most diverse family appears to be the hoverflies (Syrphidae), very small flies like midges excluded (I have an alcoholic soup of these, if anybody wants to have a go?). This may partly, of course, be explained by hoverflies tending to be more conspicuous and easily caught than many other flies, and because I am more familiar with them than other families.

In 2011, I recorded 75 species of hoverfly from the hedge. All of these were using the hedge for something, if only as a resting place while searching

more widely for mates or breeding sites. Most were seen feeding, in particular on the umbellifers. The succession of these flowers from late April through to October proved my most fertile hunting ground. Large quantities of flowering hemlock water dropwort *Oenanthe crocata* growing out of the ditch were a particular draw, although the smaller numbers of flowering stems of cow parsley *Anthriscus sylvestris*, wild angelica *Angelica sylvestris* and hogweed *Heracleum sphondylium* were equally attractive. The well-known value of umbellifers to flies makes me think about the loss of habitat that must result from all the mid to late summer roadside verge cutting that takes place. Other flowering plants in the hedge that were used extensively for feeding by the hoverflies included grey willow, blackthorn and bramble.

How many of the species are actually breeding in the hedge is something I must try and look into. A few species are most unlikely to have been doing so, for example *Anasimyia contracta* which must have come from the nearby pond, and there are no suitable rot holes in the hedges for *Criorhina berberina* and *C. floccosa*, or sap runs for the single *Ferdinandea cuprea* found on a hogweed flower. The behaviour of others suggested that a careful search would reveal their larvae – for example, females of many of the 12 *Cheilosia* species were seen flying low down amongst the herbage, searching, I suspect, for places to lay their eggs.

Although I did not record numbers, the most numerous hoverflies were certainly *Syritta pipiens, Platycheirus albimanus* and *Melanostoma* species, although *Eristalis* species were abundant at times too and it was a very good year for *Rhingia campestris*. There were a few species I expected to see but did not: *Leucozona glaucia, Helophilus trivittatus* and *Pipiza noctiluca*. I searched too for *Platycheirus ambiguus* which I also know to occur on the farm, but without success. Indeed, I did not see any of these species anywhere on the farm or nearby all last year. I wonder whether they were hit badly by the harsh winter weather. Fortunately, the summer drought that has affected much of the rest of England did not hit us here; to the contrary it has been a remarkably wet and soggy year, April excepting.

There were some nice surprises too. On 9 April I caught the first of several *Melangyna arctica* on a dandelion flower, while three days later I spied a strange looking *Syrphus* resting on a dock leaf in the ditch which turned out to be a male *Parasyrphus nigritarsis*, not, I think, recorded before from the vice county. Later I caught another *P. nigritarsis* in the farm polytunnel. In mid-April I observed a female *Cheilosia nebulosa* flying low among the primroses and other plants at the base of the bank. In June a *Xanthandrus comtus* was feeding on a hemlock water dropwort flower – this species also turned up in the polytunnel.

One intriguing question is how important is it to diversity of hoverflies using a hedge that there should be a combination of ditch, shelter-giving shrubs and flower-rich margins present? I suspect that it is the combination of these features (and probably others) that accounts for the high diversity I found. Perhaps agri-environment schemes like Environmental Stewardship in England should be tailored to reflect this?

Big Hoverfly Watch – an experimental project – volunteers sought

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For a long while we have wanted to develop a monitoring project that might provide a snapshot about the status of Britain's hoverflies. Ideally we would like to develop hoverfly transects similar to those used for butterfly monitoring. However, the difficulty of identifying hoverflies makes this more challenging and therefore we think there may be scope for doing something more along the lines of the Big Garden Birdwatch.

We therefore propose to pilot an idea in 2012 to develop a network of recorders who would visit their favourite site on two separate days once in each of two pre-arranged dates and to develop a list for that site for that date. We realise that there will be differences in the skill-base of recorders but we think there is scope for distilling the differences between complete and incomplete lists. Once we have a big group contributing, any differences in recording skills are likely to be evened out by the size of the dataset.

Proposed method:

The event will take place over two periods:

- Thursday 10 May to Sunday 20 May
- Wednesday 20 June to Saturday 30 June

During this week, participants are invited to visit a favoured site, one they want to visit regularly. During this visit they would spend a maximum of two hours between 10.00 and 13.00 recording hoverflies. The choice of weather and time is important – we know that in general hoverflies are most active in the morning – in May timing between 10.30 and 12.30 is probably about right but is weather-dependent. In late June they will fly earlier so a 10 a.m. start may be more appropriate. However, this is also potentially dependent on the latitude – flies may fly a bit later in more northerly locations so 11.00 to 13.00 may be more suitable.

This project is open to recorders of all abilities. We need to get a feel for the ability of the recorders in order to analyse the data. Our analysis of existing Recording Scheme data suggests that there are several major steps in recording confidence and this will inevitably have a bearing on how many species and the numbers of specimens recorded.

Where recorders have limited experience and want to collect specimens and send them to the HRS for identification we will be happy to take material specifically for this project – material should be forwarded to Roger Morris, 7 Vine Street, Stamford, Lincolnshire PE9 1QE. If participants are unhappy about collecting specimens for determination they are encouraged to get voucher photographs and to send them to Roger for an identification.

Data will be assembled from electronic returns and the results outlined to participants through an e-group newsletter. As this is a pilot we really need feedback on the practicalities involved. So we hope that this will be an interactive project that will be of interest to everybody.

If the initiative is successful, we hope to develop an extensive network of recorders across the whole of the UK, but in this first year a foundation group of maybe 50 participants would be sufficient to explore the practicalities of such an initiative. Ultimately, who knows how many recorders might be generated?

Advertisements placed on the Yahoo Hoverflies group, DF and HRS websites yielded an immediate response from widely differing localities in England and also from Ireland. This is encouraging as it looks as though this initiative will recruit a range of recorders who are new to hoverfly recording. This is great, but we do need to have an input from a group of experienced recorders – all welcome.

Recorder name	**Date visited**
Site Name	**Time visited**
Brief site description	**Grid ref**
Weather conditions	

Recorder experience	10+ years		3-10 years		1-3 years		novice	
	Takes specimens		Photo record only		Field ID only			
	Yes/no		Yes/no		Yes/no			

Species	Number recorded*	Species	Number recorded*

*optional

Since placing advance notice of the idea on the Yahoo Hoverfly Group and the Recording Scheme website, we have had a good response and more than 20 volunteers have been recruited.

Xanthogramma pedissequum group

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In *Hoverfly Newsletter* 50 ; spring 2011 (accompanying *Bulletin* 71), Martin Speight provided a key to *Xanthogramma* which included *dives* and *stackelbergi* as previously unrecognised species split from *X. pedissequum* : at least one of these extra species occurs in Britain.

The ideal is to have characters with unequivocal yes/no answers. Seemingly life is not so simple, making some couplets long and in places tortuous to allow for variation, including the segregation into male and female couplets. The major limitation in Speight's key is that the split between the true *pedissequum* and the other two species is based on a character which is unreliable and often difficult to interpret, and supplementary descriptive characters are not given for *pedissequum*. Thus I have teased out the characters for comment and provide a more pragmatic key.

The membrane between the tergites and sternites. Because the tergites are wider than the sternites, one has to view the ventral side of the abdomen. Ideally, segments 3 and 4 would have a completely yellow membrane in the extra species and mainly dark in *pedissequum* but there are exceptions

Second basal cell: percentage bare of microtrichia. Each species has a range of percentages. which overlap. Taking the figures given in Speight's key there are some useful thresholds, especially if playing on the safe side (I have allowed 10% margin). Thus if the second basal cell is 0-15% free of microtrichia, *pedissequum* is the only qualifying species. Above 50% bare qualifies as *dives*.

Number of yellow spots on the pleura. This can range from 1 to 5. The minimum is a single vertically elongate yellow spot about the front of the pleura, perhaps the most frequent state among *pedissequum* s.l. in Britain: such specimens qualify as *pedissequum* s.s.. So far, so good. If there is a second spot, in principle the specimen still qualifies as *pedissequum* s.s.. If there are more than two spots, any of the three species could be entailed.

Wing apex clear of darkened. The degree of darkening is minor so this character may be overlooked at casual glance. It is a feature of *dives* though not always present.

Stigma and costal cells colour. The second costal cell is yellow (or grey) in *dives* but almost clear in *stackelbergi.*

Shape of tergite markings. There are some differences between species, though variation occurs. Illustrations will be needed.

Lower squama marginal hair colour in females: dark in *dives*, yellowish in *stackelbergi*. (probably variable: *pedissequum* can be either).

Pragmatic key to *Xanthogramma pedissequum* Group

1. Side of thorax with a vertical yellow strip, otherwise black. **pedissequum**

- Side of thorax with a vertical yellow strip plus one or more additional yellow markings.....2

2. Wing below the stigma with any darkening confined to the cell immediately below (i.e. not crossing the next vein, R_{2+3}.
   [Male tergites 3 and 4 with yellow bars pinched in width about the lateral margin; tergite 2 yellow markings usually with the posterior margin angled obliquely forward (rather than backwards as a triangular wedge). Female frons with the median stripe usually narrowed or pinched-out in front (about the top of the lunules situated above the antennae).].....
   ..... **stackebergi**

   Wing below stigma with a dark patch continuing below R_{2+3}.
   .[Male tergites 2 and 3 with yellow bars that maintain their width at the lateral margins; tergite 2 triangular. Female frons with median stripe rather variable but often expanded in front to extend along the outer side of the pair of lunules.].....3

3. Apex of wing with faint darkening.at the apex. Second basal cell at least 50% bare of microtrichia.
   [Male tergite 2 with the yellow bars often pointed at inward end, but not always.]..... **dives**

- Apex of wing with no hint of darkening.. Second basal cell at most 15% bare of microtrichia (the % difference can be less extreme, but safe figures are chosen).
  [Male tergite 2 with the yellow bars usually rounded at their inward end.]
  .....**pedissequum**

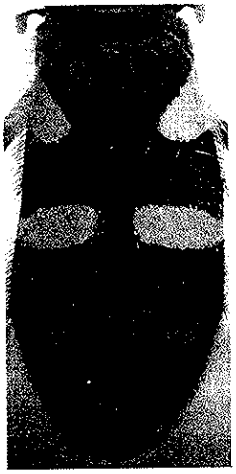
On this basis it should be possible to recognise *stackelbergi* as being distinct from *pedissequum* without recourse to the often fraught interpretation of

the colour of parts of the membrane between the tergites and sternites. The membrane character (explained above) should still be checked. If in the slightest doubt, vouchers will needed for verification.

Unfortunately *dives* does not always have the wing darkened at the apex and even then the marking can be faint. As yet I am not aware of a British specimen. If a clear-winged specimen were to escape recognition, it would run to the commonest species, *pedissequum*, where a misidentification would not be too serious.

All existing data already lodged with the Hoverfly Recording Scheme will have to be treated as *pedissequum* s.l. (broad interpretation of that species). Where vouchers for any of those records can be checked, data can be resubmitted as *pedissequum* s.s. (strict sense) or as one of the 'new' species if there is no room for ambiguity of characters. If *dives* is reported, verification will be essential.

Male *pedissequum* s.s.
Tergite 2 spots triangular
Tergites 3 & 4 spots uniform width to outer end



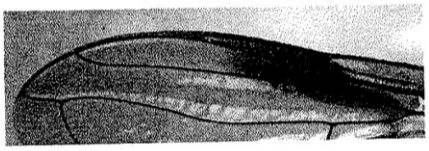
Male *stackelbergi*
Tergite 2 spots oblique
Tergite 3 & 4 spots pinched at outer end

Female *pedissequum* s.s.
Frons stripe wide in front

Female stackelbergi
Frons spots tapered in front

pedissequum s.s. pleura
Vertical strip on mesopleuron only

5 spot pleura
Vertical strip on mesopleuron
Spot below at top of sternopleuron
Spot in front of propleuron
Spot in front of haltere
Spot below the latter



pedissequum s.s.
Dark area decends below R_{2+3}

stackelbergi
No darkening below R_{2+3}

Where does Marmalade come from?

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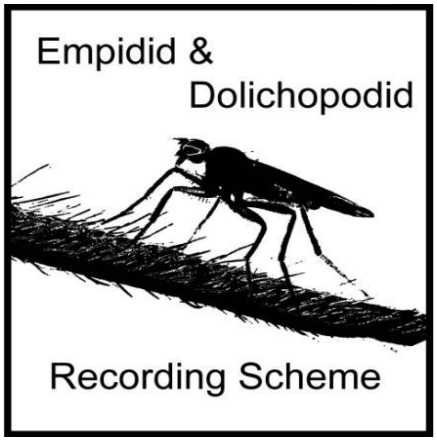
Most readers of the Bulletin will probably have heard *Episyrphus balteatus* referred to as the Marmalade Hoverfly. Despite the name being somewhat whimsical, there seems to be no criticism of it, even among the most serious of dipterists. Perhaps that's because the transparent orange colour and strand-like bars of the fly's abdomen actually fit the title rather well. Certainly, this is a name to catch the imagination of our fellow-citizens, particularly children: and it's good to have a common name for such a common insect.

My question is: does anybody know who first coined the name? For helping to spread awareness, at least, the originator should be the toast of hoverfly-enthusiasts everywhere!

(Editor's comment: I first encountered the name in **Hoverflies of the Sheffield Area and North Derbyshire** by Derek Whiteley, published by the Sorby Natural History Society in 1987, as Sorby Record Special Series No. 6 (ISSN 0260-2032). In this excellent book three species are illustrated by line drawings, and each of the three is captioned with its scientific name accompanied by an imaginatively chosen English name. The three are *Episyrphus balteatus* (The Marmalade Fly), *Rhingia campestris* (The Heineken Fly) and *Helophilus pendulus* (The Footballer). The only other English names for hoverflies that are used in the book are the few well-established ones such as Drone Fly and Narcissus Fly. I do not know whether these names were coined by Derek Whiteley himself or by others. Perhaps, Derek, if you read this, you, or anyone else who knows, would let the newsletter know who was the author of these names).

The Marmalade Fly *Episyrphus balteatus* (photo: David Iliff)



Newsletter No. 17
Spring 2012

Editorial

Adrian Plant & Martin Drake

Empidid and dolichopodid enthusiasts are fortunate in having a large and very detailed literature covering British species. They are rather less fortunate in that amassing a 'complete' collection of these works is expensive, some of the works are out of date or difficult to obtain, some keys are overly simplistic or needlessly complicated and additional species have been added as British since publication of the major works. A complete revision and updating of the pantheon of literature is a daunting prospect. A more piecemeal preparation of keys to difficult or otherwise inaccessible groups seems more likely to succeed so here we provide a first offering ~ a revised and updated key to the large hybotid genus *Platypalpus.* We hope to be able to provide keys to other difficult groups in the not too distant future.

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A Key to British Species of *Platypalpus* Macq. (Hybotidae)

Adrian Plant

Platypalpus Macq. is a very speciose genus of small predatory hybotids commonly encountered running on leaf and other surfaces. Collin's British Flies (Collin 1961) provided a useful key and detailed descriptions of the 75 species of *Platypalpus* known from Britain at the time. Since Collin's seminal work, the number of species confirmed as British has risen to at least 88, thanks largely to excellent keys to the entire European fauna produced by Chvála (1975, 1989) and Grootaert & Chvála (1992). Whilst all British species are keyed in Grootaert & Chvála (1992), this large work is not readily available to many. The key provided here is based unashamedly on their key, but with considerable modification in the light of the wider literature and my own notes. It includes all species currently known as British as well as some which might turn up in future. Unfortunately, lack of time and potential copyright issues have prevented inclusion of illustrations to the key and the user may still have to look for these in the primary references.

Platypalpus is not an easy group to identify correctly.- but it is not difficult either and with a little practice and persistence most species may readily be identified using this key alone. Male genitalia illustrations are useful and will be needed to confirm some identifications. However, males are rare in some species (e.g. *P. candicans* & *P. cursitans*) over large parts of their range while for others (e.g. *P. major*) males are almost unknown and determinations must rely on characters of parthenogenetic females. Dwarf forms occur occasionally (e.g. form *minor* of *P major*) and may be the product of repeated parthenogenetic reproduction.

Common pitfalls encountered in identification include confusion over the number of vertical setae, presence / absence of an anterior notopleural, colour of the basal antennal segments (very variable in some species) and the extent of dusting on the thoracic dorsum; ~ you have been warned! I have made extensive use the *length to width* ratio of the postpedicel (3rd antennal segment) and of the stylus (arista); these can be difficult to estimate accurately and it is often better to use the ratio between the *length of the stylus compared to the postpedicel*. Colour is always *ground colour* (visible in both wet and dry-mounted specimens) and any reference to colour of dusting is specifically mentioned. Some species are keyed both ways but if you get into difficulties then try the other option in a couplet (as with any key, don't try to 'force' an option).The key should be regarded as provisional and

it is preferable to confirm determinations by reference to original works.

Primary Literature

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Chvála, M. (1989). Monograph of the northern and central European species of *Playpalpus* (Diptera, Hybotidae), with data on the occurrence in Czechoslovakia. *Acta Universitatis Carolinae – Biologica* 32: 209-376.

Grootaert, P. & Chvála, M. (1992). Monograph of the genus *Platypalpus* (Diptera: Empidoidea, Hybotidae) of the Mediterranean region and the Canary Islands. *Acta Universitatis Carolinae – Biologica* 36: 3-226.

Platypalpus laticinctus ©Adrian Plant

Platypalpus major ©Adrian Plant

Key to species groups

(Note that species groups are formed for convenience of constructing a key and may not necessarily reflect systematic relationships between the species included)

1 Thorax with predominantly yellow ground colour; if darkened dorsally then pleura always distinctly yellowish. ..... **GROUP A**
- Thorax with predominantly black ground colour although often densely grey or yellowish-grey dusted. **2**

2 *vt* setae not clearly differentiated from numerous evenly long fine setae of upper occiput and vertex (care needed as occasionally [e.g. *P. pallipes*] 1 pair *vt* may be weakly developed in some individuals but in doubtful cases the scutum has fine hairs rather evenly distributed over its surface with line of *acr* and *dc* hardly differentiated); T_2 with apical spur never strongly developed. ..... **GROUP B**
- 1 or 2 pairs distinct *vt* setae which even if small are clearly differentiated from other fine setae of upper occiput and vertex. **3**

3 1 pair distinct *vt* setae. **4**
- 2 pairs distinct *vt* setae. **8**

4 Scutum more or less polished black or subshining, at least along median line; any dusting usually rather thin (care and comparative experience are useful but all species lacking *h* and with whitish stylus belong here). ..... **GROUP C**
- Scutum lightly to densely greyish dusted. **5**

5 Scape and/or pedicel blackish (occasionally reddish-brown or yellowish brown). ..... **GROUP E**
- Scape and/or pedicel yellowish or yellowish brown. **6**

6 Pleura entirely grey dusted, including *katepisternum*. ..... **GROUP D**
- Pleura with *katepisternum* partly polished. ... **7**

7 Posthumeral setae (anterior notopleural) present; T_1 and T_3 with short dark setae dorsally. ..... **GROUP F**
- Posthumeral setae (anterior notopleural) not developed; T_1 and T_3 without short dark setae dorsally (except in *P. stabilis* and *P. major*). ..... **GROUP G**

8 Scutum polished black, at least partially on posterior third; T_2 lacking of with only short apical spur, never longer than limb is deep. ..... **GROUP H**
- Scutum entirely dusted grey or greyish yellow; T_2 with or without strong apical spur. ..... **GROUP I**

KEY A. Species with predominantly yellow thorax

1 *vt* setae absent or indistinguishable in length or strength from surrounding hairs of upper occiput; *h* absent; scutum mostly polished; T_2 covered with minute pale pile distally or apically; T_2 with apical spur absent or very weakly developed. **2**

- 1–2 pairs *vt* clearly distinct from other hairs; *h* present or absent; T_2 without minute pile distally and apical spur always present, if sometimes little more than a very small projection at tip. **4**

2 *acr* and *dc* small and numerous, rather evenly distributed over scutum, the two series hardly distinguishable from each other; T_2 with short blunt spur; [a short section of C beyond junction with R_1 often, but not always, deep black contrasting strongly with otherwise yellowish veins]. ..... ***pectoralis* (Fallén)**

- *acr* and *dc* distinctly separated by bare areas; T_2 lacking apical spur. **3**

3 *acr* 4–6 serial, minute; antenna black; T_2 yellow apically. .... ***inexpectatus* Smith & Chvála**

- *acr* 2–4 serial, fine, diverging; scape and pedicel yellow, postpedicel black; T_2 velvety-brown on distal 0.5. ..... ***mikii* (Becker)**

4 2 pairs of distinct *vt* setae. .... ***aurantiacus* (Collin)**

- 1 pair of distinct *vt* setae. **5**

5 Head entirely yellow; apical tarsal segments of front leg dilated. ..... ***luteus* (Meigen)**

- Head with occiput darkened; apical tarsal segments of front leg not dilated. **6**

6 F_2 without *pv* setae behind double row of dark spine-like setae (look carefully). .... ***exilis* (Meigen)**

- F_2 with distinct pale *pv* setae behind double row of dark spine-like setae. **7**

7 Postpedicel entirely whitish-yellow; tarsi yellow; katepisternum polished. **8**

- Postpedicel blackish, at least at tip; tarsi weakly annulated brownish; katepisternum dusted. ..... ***leucocephalus* (Von Roser)**

8 T_2 with strong black spur longer than limb is deep; postpedicel yellowish, 1.5X long as wide, stylus black (but sometimes distinctly yellowish at base); male genitalia smaller with left lamella yellow. ..... ***luteolus* (Collin)**

- T_2 with short blunt black spur, shorter than limb is deep; postpedicel whitish, 2–2.5X long as wide, stylus whitish at base; male genitalia large with left lamella black. .... ***luteoloides* Grootaert** [Not British]

KEY B. Species with black thorax and lacking vertical setae

1 F_2 lacking distinct setae behind ventral row of double spines; *acr* and *dc* rows at least narrowly separated from each other. **2**

- F_2 with more or less distinct setae behind ventral row of double spines; *acr* and *dc* rather evenly and densely distributed on scutum, not obviously separated from each other. **6**

2 Basal antennal segments yellow. .... ***unguiculatus* (Zett.)** [Not British]

- Basal antennal segments blackish. **3**

3 *acr* 2–4 serial, numerous, numerous fine setulae between line of *dc* and lateral margin of scutum; pleura dusted except for katepisternum. **4**

- Scutum almost bare with *acr* and *dc* minute, only a few fine setulae between line of *dc* and lateral margin of scutum; pleura largely polished. **5**

4 T_2 with dense covering of pale pile along entire length or at least distal 0.8; frons subshining black (care with wet-preserved specimens); male genitalia small. .... ***parvicauda* (Collin)**

- T_2 with dense covering of pale pile on distal 0.5 only; frons densely grey dusted; male genitalia globular, greatly enlarged. .... ***ciliaris* (Fallén)**

5 Frons and vertex dusted grey; T_2 with covering of pale pile apically; antennal stylus ≥2 X length of postpedicel; male abdomen sparsely setulose. ..... ***confinis* (Zett.)**

- Frons and vertex shining black; T_2 without covering of pile apically; antennal stylus shorter (♂) or slightly longer (♀) than postpedicel. ..... ***stigmatellus* (Zett.)**

6 Antenna with postpedicel 3.5–4X long as wide, stylus 0.5X as long; F_2 only slightly more inflated than F_1; *acr* irregularly 6-serial, narrowly separated from multiserial dc; male apical tarsal segments on front leg narrow and elongate. ..... ***longimanus* (Corti)**

- Antenna with postpedicel ≤3 X long as wide, stylus longer; F_2 obviously more inflated and longer than F_1; lines of *acr* and *dc* not obviously separated. **7**

- Frons shining black; antenna with postpedicel slightly longer than wide; F_2 and F_3 more or less darkened subapically; F_2 with fine yellow *pv* setae. ..... ***macula* (Zett.)**
- Frons dusted grey; antenna with postpedicel ≥ long as deep; F_2 and F_3 yellow; F_2 with strong dark *pv* setae. ..... ***pallipes* (Fallén)**

KEY C. Species with blackish thorax, one pair of vertical setae and scutum at least partly shining or subshining.

1 T_2 without or with only very small apical spur; scutum more or less polished with at most only lateral margins slightly dusted; [if scutum completely but very thinly dusted and antennal stylus white, the species will be found in this section; otherwise species with very thinly dusted scutum and stylus dark which fail to be resolved in this section should be keyed in Group E] **2**

- T_2 with large sharply pointed apical spur, if only as long or slightly longer than limb is deep then F_2 with distinct *pv* setae behind double row of ventral points. **11**

2 F_2 without distinct *pv* setae behind double row of ventral points. .... ***unguiculatus* (Zett.)** [Not British]

- F_2 with distinct *pv* setae behind double row of ventral points. **3**

3 Anepisternum partly or completely shining; humeri shining; F_1 with at least 1–2 long black median setae in posterior ventral row longer than limb is deep. **4**

- Anepisternum completely dusted; at least posterior part of humeri dusted; F_1 without long setae in posterior ventral row. **8**

4 Antenna with postpedicel 3X long as wide, stylus much longer; frons narrow, no wider than front ocellus. **5**

- Antenna with postpedicel 4–5X long as wide, stylus similar or slightly longer than postpedicel; frons dorsally wider than front ocellus. **7**

5 *ocl* and *vt* setae of equal length (♂ L. cercus very large, apically bent). .... ***smirnovi* (Kovalev)** [Not British]

- *ocl* setae 0.6–0.7X length of *vt* (♂ L. cercus broadened or pointed, but not greatly enlarged). **6**

6 Antenna with postpedicel >3X long as wide, stylus ≤2X as long (♂ L. cercus apically pointed). ..... ***albiseta* (Panzer)**

- Antenna with postpedicel about 3X long as wide, stylus almost 3X as long (♂ L. cercus apically broadened). ..... ***pygialis* Chvála**

7 Legs extensively blackish; frons grey. ..... ***albocapillatus* (Fallén)**

- Legs with coxae, femora and T_3 yellowish brown; F_1 black dorsally and on apical 0.3; F_2, F_3 and T_3 darkened apically. .... ***niveocapillatus* Chvála** [Not British]

8 *acr* 4-serial long and distinct. **9**

- *acr* 2-serial; scutum almost bare with only minute setulae. **10**

9 Larger thoracic setae whitish; antenna with postpedicel 2.5X long as wide, stylus 2X as long. ..... ***pallidiseta* Kovalev**

- Larger thoracic setae black; antenna with postpedicel ≥4X long as wide, stylus almost as long. ..... ***obscurus* (Von Roser)** [Not British]

10 Antenna with postpedicel 3X long as wide, stylus 2X as long; ♂ R. periandrial lamella with tooth-like process on left margin, L. lamella distally narrow with black spines apically. ..... ***caroli* Grootaert**

- Antenna with postpedicel 4X long as wide, stylus as long or slightly longer; ♂ R. periandrial lamella without tooth-like process on left margin, L. lamella not so narrow distally and lacking distinct black spines apically. ..... ***niveiseta* (Zett.)**

11 Scutum shining, polished black at least in prescutellar area or extensively anteriorly between humeri; often mostly polished with dusting confined only to a patch behind humeri, notopleural depression and postalar calli. **12**

- Scutum very thinly dusted (can appear subshining or with metallic tinge) sometimes (*minutus* / *australominutus*) with narrow median shining stripe. **16**
[this couplet can cause problems, especially with rubbed dry-mounted material; *P. niger* is keyed both ways as intensity of dusting varies between populations. *P. politus* may cause problems and its dusting pattern is sexually dimorphic. Problematic specimens should be keyed both ways and never 'forced']

12 Legs black with only base of all femora yellow. ..... ***niger*** **(Mg.)**
[*niger* varies somewhat throughout its European range and the form keying out here is typical of British and European Atlantic seaboard populations; specimens of other European populations will resolve at couplet 18 which is included 'just in case'. Note that *P. ater* (Wahlberg) is a boreal species that might just occur in the north of Scotland, it resembles *niger* but all larger thoracic setae are pale and the femora are more uniformly dark]
- Legs mostly yellow. **13**

13 F_2 conspicuously inflated, pv bristles weak or absent except for 2-3 apically. ..... ***pseudociliaris*** **(Strobl)**
- F_2 less strongly inflated, only slightly deeper than F_1 (in doubtful cases thoracic pleura extensively polished). **14**

14 *h* present, distinct; *acr* and *dc* distinct, [tarsi with brown annulations becoming darker apically, apical segment entirely dark]. ..... ***ingenuus*** **(Collin)**
- *h* minute; *acr* and *dc* minute [apical 2–3 tarsal segments darkened]. **15**

15 Antenna yellow including postpedicel; scutum polished on prescutellar area (♂) or prescutellar area and median stripe (♀). ..... ***ruficornis*** **(Von Roser)**
- Antenna rather darker yellow, postpedicel rather narrower and apically darker; scutum polished at extreme front (♂) or more or less entirely polished (♀). ..... ***politus*** **(Collin)**

16 Legs yellow with darkened tarsi; occiput polished black; antenna with scape and pedicel yellow, postpedicel small and black, stylus long and whitish; *h* absent; thoracic pleura and humeri shining. ..... ***leucothrix*** **(Strobl)**
- Legs extensively darkened; occiput dusted; *h* present even if sometime very fine; antenna dark including basal segments and stylus. **17**

17 *h* distinct; large thoracic setae yellow; scutum metallic black with very thin dust; frons very narrow; vein CuA_2 closing cell *cup* (ie. vein closing anal cell) distinctly S-shaped and recurrent. ..... ***aeneus*** **(Macq.)**
- *h* small and fine; large thoracic setae dark; vein CuA_2 closing cell *cup* less strongly recurrent. **18**

18 Legs black with only base of all femora yellow; pleura extensively polished; T_1 without small rim-like apicoventral process; scutum variably thinly dusted. ..... ***niger*** **(Mg.)**
- Femora with different pattern (take care as *minutus* / *australominutus* can have base of all femora yellow but they also always have yellowish markings apically); pleura silvery grey dusted except for shining katepisternum. **19**

19 T_1 without small rim-like apicoventral process; *acr* 2-serial, rows widely separated; legs black, tarsi uniformly dark without annulations, at most F_1, (rarely F_2 and F_3 very narrowly) yellowish apically. .... ***albifacies*** **(Collin)**
- T_1 with small rim-like apicoventral process; *acr* 2-serial, rows narrowly separated; legs black, femora apically yellowish. **20**

20 ♂ genitalia with R. periandrial lamella apically bifid, tips of bifurcation rounded and separated by a narrow slit; cercus with 2 apical spines. ..... ***minutus*** **(Mg.)**
- ♂ genitalia with R. periandrial lamella apically bifid, tips of bifurcation pointed and separated by a V-shaped gap; cercus with 3 apical spines. ... ***australominutus*** **(Grootaert)**
[females of *minutus* and *australominutus* are inseperable].

KEY D. Species with black thorax, one pair of vertical setae, scutum dusted greyish, basal antennal segments yellowish and katepisternum entirely dusted

1 Veins M and R_{4+5} almost parallel, M at most gently bowed. **2**
- Veins M and R_{4+5} obviously not parallel, M strongly bowed away from R_{4+5}. **3**

2 *dc* setae large and bristle-like; abdomen shining black; antenna with postpedicel yellow (darkened only at tip) in male or dark (yellowish only at base) in female. .... ***verralli*** **(Collin)**
- *dc* setae small (except 1-2 pairs in prescutellar area larger); abdomen yellow; antenna with postpedicel entirely yellow in both sexes. ..... ***dessarti*** **Grootaert** [Not British]

3 Vein M strongly and evenly bowed, even apically; tarsi pale or indistinctly annulated; frons very narrow (no wider than front ocellus). ..... ***candicans*** **(Fallén)**
- Vein M not so strongly and evenly bowed, curving upwards rather abruptly apically where subparallel with R_{4+5}; tarsi with brownish annulations; frons broader. .. .. ***cursitans*** **(Fabr.)**

KEY E. Species with black thorax, one pair of vertical setae, scutum distinctly dusted and basal antennal segments dark

1 F_2 without *pv* bristles behind the double row of small black ventral spines; *acr* and *dc* minute; tarsi yellow or with only apical tarsomere dark. **2**
- F_2 with distinct *pv* bristles behind the double row of small black ventral spines [look carefully, *pv* bristles can be very fine and difficult to see]; *acr* and *dc* moderately long; tarsi distinctly annulated or with apical 1-2 tarsomeres darkened. **3**

2 T_2 with long sharply pointed apical spur about as long a limb is deep; postpedicel 3X long as wide, stylus thickened; tarsi completely yellow. ..... ***aristatus*** **(Collin)**
- T_2 with blunt apical spur about as long a limb is deep; postpedicel 2-2.5X long as deep, stylus slender; apical tarsomere black. .... ***tonsus*** **(Collin)** [note that in *tonsus* the double row of black ventral spines becomes longer and yellowish basally and could be mistaken for basal *pv* bristles]

3 T_2 with apical spur shorter than tibia is deep or if about as long, then blunt tipped. **4**
- T_2 with apical spur large, sharply pointed, longer than limb is deep [species with blunt tipped spur about as long as limb is deep are keyed both ways]. **5**

4 Wing membrane distinctly yellowish; apical 1-2 tarsomeres of all legs dark; T_2 with apical spur about as long as limb is wide, blunt, ♂ with tiny spine at tip. ***cothurnatus*** **Macquart**
- Wing membrane clear or faintly brownish; tarsi distinctly annulated; T_2 with apical spur very small. ..... ***cryptospina*** **(Frey)**

5 T_2 with apical spur only about as long as limb is wide, blunt, ♂with tiny spine at tip Wing membrane distinctly yellowish; apical 1-2 tarsomeres of all legs dark. .... ***cothurnatus*** **Macquart**
- T_2 with apical spur long, sharply pointed (other characters various). **6**

6 Larger thoracic bristles black. **7**
- Larger thoracic bristles yellowish to brownish. **8**

7 Legs extensively darkened, coxae black; T_2 with apical spur sharply pointed (*acr* clearly 4-serial at front). ..... ***melancholicus*** **(Collin)**
- Legs extensively yellowish, coxae yellow; T_2 with apical spur blunter tipped with a minute spine and even smaller hair apically (*acr* usually 4-serial but sometimes 2 or 3 serial in part). ..... ***optivus*** **(Collin)**

8 Scutum with *acr* 4-serial, at least in front. legs usually extensively darkened; F_1 not much narrower than F_2. ..... **9**
- Scutum with *acr* 2-serial [care! *P. notatus* can have a few extra *acr* and appear 4-serial in part]; legs yellowish or extensively darkened; F_1 sometimes obviously narrower than F_2. **10**

9 Large thoracic bristles yellowish; *acr* rather long, usually 4-serial throughout; legs usually paler with C_2, C_3, C_1 at base, rather broad rings on all femora and tip of T_1 and T_3 darkened; hind trochanter usually yellowish; tarsi very strongly dark annulated; wing membrane vaguely darkened, veins brown; smaller species (2.3-3.3 mm). .... ***annulatus*** **(Fallén)**

- Large thoracic bristles brownish to black; *acr* shorter, usually 2-serial about middle and posteriorly; legs more extensively darkened with all coxae and all femora (except at tip) strongly darkened; hind trochanter darkened; tarsi less strongly annulated (tarsomeres with dark apical part less abruptly divided from paler basal part); wing membrane distinctly brownish, veins blackish brown; larger species (2.9-3.8 mm). ..... ***melancholicus*** **(Collin)**

10 Antenna with postpedicel at least 2.5X long as deep, stylus about as long or slightly longer than postpedicel. **11**
- Antenna with postpedicel shorter , no more than 2X long as deep, stylus obviously longer than postpedicel. **16**

11 Abdomen polished black, sometimes with small patches of grey dusting laterally on tergites 1 and 2. ..... **12**
- Abdomen with distinct patches of grey dusting on all tergites basally. **14**

12 Vt setae closer together (hardly 1.5X width of frons by anterior ocellus); F_2 much stouter than F_1; tergites 1 and 2 with small lateral patches of dusting; tarsi faintly annulated but apical tarsomere black, legs otherwise yellow with conspicuous black 'knees'. .... ***infectus*** **(Collin)**
- Vt setae wider apart (about 2X width of frons by anterior ocellus); F_2 not much stouter than F_1; abdomen entirely shining black; legs yellowish or extensively darkened but always with distinct annulated tarsi. **13**

13 Legs extensively darkened; at least C_2, C_3, C_1 at base, F_2 and F_3 apically dark [paler individuals occur, their coxae are dark at least about base, F_2 and F_3 have at least a dark dorsal patch or median ring]; face narrower than frons anteriorly; *vt* setae pale. .... ***notatus*** **(Mg.)**
- Legs extensively yellowish [pale yellow to orange]; C_2, C_3 at most dark basally, F_2 and F_3 sometimes faintly dark ring or apex; frons broader, similar width as face; *vt* setae brownish. ..... ***strigifrons*** **(Zett.)**
[very pale examples of the *notatus* can be confused with dark specimens of *strigifrons* and determination should be confirmed by genitalia examination; *notatus* is common and widespread whereas *strigifrons* is confined to sand dunes]

14 Legs yellow (including coxae); basal antennal segments sometimes dark reddish brown [some individuals recall dark examples of *P. pallidiventris* but in that species the anterior notopleural is developed]. .... ***praecinctus*** **(Collin)**
- Legs obviously darkened on coxae and femora. **15**

15 Antenna entirely dark; acr irregularly 2-3 serial; stylus of equal length as postpedicel (♂) or slightly longer (female). .... ***carteri*** **(Collin)**
- Antenna with basal segments reddish yellow; acr regularly 2-serial; stylus 1.5X length of postpedicel. ..... ***latemi*** **Grootaert**
[specimens conforming with *latemi* have been found in Britain. It has not been admitted formally to the British list and since first describing the species, Grootaert has expressed doubts (pers. com.) that it is a valid species. It may be a dark form of *P. praecinctus*].

16 Smaller [1.5-2.6 mm]; palpi smaller, greyish yellow [can be quite dark]; clypeus polished black; antenna with postpedicel only slightly longer than wide; legs yellow (including coxae and femora), tarsi annulated; *vt* setae wider apart; dusting on scutum tinged golden yellow. ..... ***clarandus*** **(Collin)**
[*P. luteipes* Zusková occurs on near continental Europe; it resembles *clarandus* but is somewhat larger (3.0 mm) with yellow palpi, anterior and mid tarsi only faintly annulated and F_2 conspicuously larger than F_1].
- Larger [2.4-3.6 mm]; palpi quite large, brownish; clypeus dusted silvery-grey; antenna with postpedicel 1.5X long as wide; legs usually with dark markings on femora; *vt* setae closer together; dusting on scutum tinged brownish grey. **17**

17 Palpi clearly longer than broad; *acr* less numerous, the 2-serial rows conspicuously wide apart; ♂T_1 not spindle-shaped, with short ventral hair. ..... ***interstinctus*** **(Collin)**
- Palpi broadly ovate, hardly longer than broad; *acr* more numerous, the 2-serial rows closer together; ♂T_1 spindle-shaped, with longer pv bristles and a few bristly hairs dorsally. .... ***pseudofulvipes*** **(Frey)**
[leg colour of *interstinctus* and *pseudofulvipes* varies from predominantly yellow to extensively darkened. *P interstinctus* usually has at least a dark subapical ring on F_2 and *pseudofulvipes* often has

F_1 and F_2 dark but both species (especially *pseudofulvipes*) can have almost entirely dark legs and both have strongly annulated tarsi. Palpus shape is the best character to distinguish the two].

KEY F. Species with black thorax, one pair of vertical setae, scutum distinctly dusted, basal antennal segments yellow (at least dark reddish yellow), katepisternum partly polished, posthumeral setae (anterior notopleural) present and with T_1 and T_3 bearing short dark setae dorsally.

1 Smaller (2-3 mm); *acr* 2-serial [acr are usually not numerous in prescutellar depression]. **2**
- Larger (3-4 mm); *acr* 3 or 4-serial, clearly more 2-serial but series sometimes rather irregular [*acr* are usually numerous in prescutellar depression]. **5**

2 *Acr* with the 2 rows clearly separated (if narrowly), hairs not diverging; anterior notopleural of similar size to posterior pair. **3**
- *Acr* with the 2 rows hardly separated, appearing almost 1-serial but diverging strongly from each other. **4**

3 Antenna with postpedicel narrowly yellowish about base; ♂T_1 slightly spindle-shaped and front tarsi yellow with contrastingly blackish apical segment; ♀with all tarsi annulated but apical segment sometimes obviously somewhat darker than preceding segments. ..... ***longiseta* (Zet.)**
- Antenna with postpedicel not paler basally; male T_1 not inflated; all tarsi strongly annulated in both sexes. ..... ***pallidiventris* (Mg.)**

4 Coxae and trochanters yellow (sometime slightly darkened); palpus yellow in male, darkened in ♀. ..... ***kirtlingensis* Grootaert**
- Coxae and trochanters black (C_1 pale apically in female); palpus dark in both sexes. ..... ***pictitarsis* (Becker)**

5 Katepisternum with large polished patch anteriorly; *acr* irregularly 3-serial (at least behind); antenna with postpedicel 2.5X long as deep, stylus ≤ 1.5X as long; tarsi annulated, but ♂ narrowly brownish annulated on 3rd and 4th segment with 5th blackish (as in *longiseta*), ♀ with all tarsi rather faintly brownish annulated but 5th segment darker. ..... ***biapicalis* Wéber**
- Katepisternum indistinctly polished at middle; *acr* irregularly 3-4 serial (usually 4-serial) throughout; antenna with postpedicel ≤ 2X long as deep, stylus ≥ 2X as long; tarsi distinctly blackish-annulated but metatarsi brownish. ..... ***analis* (Mg.)**

KEY G. Species with black thorax, one pair of vertical setae, scutum distinctly dusted, basal antennal segments yellow (at least dark reddish yellow), katepisternum partly polished, posthumeral setae absent and with T_1 and T_3 not usually bearing short dark setae dorsally.

1 Antenna yellow; at most tip of postpedicel and stylus darkened. **2**
- Antenna with postpedicel darkened, at most narrowly paler / yellowish basally. **7**
[this character can be problematical in some examples; if in doubt key both ways.- go to couplet 2 if the palpi are large and couplet 7 if palpi are small. *P. stabilis* in particular can have the postpedicel yellowish basally and should be considered. Additionally, in *P. divisus* ♀ the postpedicel may appear brownish].

2 Vein M very obviously bowed; F_2 strongly inflated, 2X thick as F_1; usually large species but dwarf forms occur. (2.8-5.5 mm). ..... ***major* (Zet.)**
- Vein M almost parallel with R4+5; usually smaller (≤3 mm). **3**

3 Palpi long, narrow, silvery white; Antenna with postpedicel pale (♂) or brownish (♀), long (2.5X long as deep with stylus slightly longer); tarsi yellow with 5th segment blackish and basal three segments of front tarsus subannulate ventrally (i.e. with a small dark spot apicoventrally); wings distinctly yellowis..... ***divisus* Walker**
- Palpi broadly ovate; Antenna with postpedicel ≤ 2Xlong as deep; wings clear or faintly yellowish. **4**

4 Very small species (1.4-1.8 mm); antenna deep yellow; T_2 with apical spur short, trowel-like; tarsi yellow, only 5th segment darkened. ..... ***ochrocerus* (Collin)**

- Larger species (usually c2.5 mm); antenna paler yellow; T_2 with strong sharply pointed apical spur (if blunt then spur is longer than T_2 is deep or [*pallidicornis* ♂] tarsi dark annulated). **5**

5 Tarsi yellow with only 5th segment darkened. ..... ***albicornis* (Zet)**
- Tarsi with dark annulations. **6**

6 Frons broader; face silvery grey; abdomen distinctly dusted grey laterally; T_2 with apical spur sharply pointed in both sexes; tarsi strongly annulated black. .... ***flavicornis* (Mg.)**
- Frons narrow; face yellowish grey; abdomen very indistinctly dusted grey laterally; T_2 with apical spur sharply pointed (♀) or blunt (♂); tarsi less strongly annulated black. ..... ***pallidicornis* (Collin)**

7 T_2 with apical spur blunt & shorter than limb is deep. **8**
- T_2 with apical spur large and sharply pointed (if blunt tipped [*annulipes* & *subtilis* ♀] then at least as long as limb is deep). **10**
[all species following couplet 8 are small (≤ 2 mm) whereas those following couplet 10 are larger (excepting *calceatus* & subtilis which are c1.3-2.6 mm)].

8 Antenna with postpedicel ≥ 3X long as deep, stylus shorter; stigma present at apex of R_1. ..... ***stigma* (Collin)**
- Antenna with postpedicel c1.5X long as deep, stylus longer; stigma absent. **9**

9 C_2, C_3 and palpi yellow; ♂ with all tarsal segments of front leg sharply annulated black (less darkened in ♀). ..... ***articulatoides* Frey**
- C_2, C_3 and palpi black; ♂front leg with tarsal segments 1 & 2 annulated black, 3 & 4 yellow with only very faint annulations, 5 darkened (less distinct in ♀). ..... ***articulatus*** Macquart
[*P. maculimanus* (Zet.) may yet be found in Britain; the tip and outer margin of the left periandial lamella have only short hairs whereas in *articulatus* there are c4 strong apical bristles. ♀♀ indistinguishable].

10 Large species (3-5 mm). T_2 with large sharply pointed spur. **11**
- Smaller species (≤ 3 mm, if c3 mm the tibial spur blunt [*annulipes*] or antennal postpedicel yellowish basally [*stabilis*]. **13**
[*P. latemi* (see Key E) is also c3 mm; its abdomen is strongly dusted grey, coxae and front legs darkened and basal antennal segments reddish yellow].

11 Abdomen strongly dusted grey dorsally with only narrow median triangles polished black; *acr* and *dc* dark brown, *dc* strong. ..... ***fasciatus* (Mg.)**
[F_2 and F_3 usually darkened near tip; tarsi annulated.- brownish basally, broader & darker on apical segments]
- Abdomen more extensively polished with grey lateral patches narrower; *acr* and *dc* pale and small. **12**

12 Abdomen with lateral dust patches confined to anterior two segments; segments 3-5 with narrowly dusted anterior margins; antenna with postpedicel usually yellowish basally. ..... ***laticinctus* Walker**
[very variable in size and colour; postpedicel yellowish or dark basally; legs usually yellowish but F_2, F_3 and sometimes even F_1 darkened; front tarsi generally yellowish, mid and hind leg indistinctly annulated].
- Abdomen with broad lateral dust patches on all segments; antenna with postpedicel entirely blackish. ..... ***cruralis* (Collin)**[Not British]

13 Antenna with postpedicel broadly yellowish on basal 0.5 (look from below ~ occasionally only narrowly yellowish in ♀); T_1 of ♂ thickened; tarsi faintly annulated brownish, darker on apical 2 segments [size very variable, usually 2.6-3.3 mm]. ..... ***stabilis* (Collin)**
- Antenna with postpedicel completely blackish; (narrowly yellow at base in *ecalceatus* and rarely so in *annulipes* which has strongly annulated tarsi). **14**

14 *Acr* irregularly 3-4 serial (care! can appear 2-serial in rubbed specimens); tarsi with distinct black annulations (very strongly defined on front leg); larger, 2.5-3.5 mm. ..... ***annulipes* (Mg.)**
[this common species is confusingly variable; the basal segments of the antenna are sometimes almost black and the postpedicel may be yellowish basally and varies considerably in length. Leg colour varies from entirely yellow to extensively blackish but the tarsi are always distinctly annulated.; most specimens with very distinctly annulated front tarsi turn out to be *annulipes*. A putative undescribed but related species is present

in Britain; it has C_1 and F_1 (except at tip) deep black and left periandrial lamella narrower].

- Acr regularly 2-serial; tarsi entirely yellowish, with apical 2 segments darkened or annulated; generally smaller, 1.5-3.2 mm. **15**

15 Fore tarsi distinctly annulated. .... ***subtilis*** **(Collin)** [only 1 notopleural, tibial spur blunt in ♂, sharply pointed in ♀; looks like small pale version of *annulipes* but in *subtilis* wing veins are all pale whereas in *annulipes* they are only yellowish on the costal half only].

- Fore tarsi yellow or with apical segments darkened. **16**

16 Fore tarsi entirely yellow. .... ***ecalceatus*** **(Zet.)**

- Fore tarsi with apical 2 segments darkened. ..... ***calceatus*** **(Mg.)**

KEY H. Species with black thorax, two pairs of vertical setae, scutum polished black (at least posteriorly)

1 Antenna with postpedicel short ≤ 3X long as deep, stylus as long or longer than postpedicel. **2**

- Antenna with postpedicel long, ≥ 4X long as deep, stylus ≤ 0.6X length of postpedicel. **7**

2 Antenna entirely black; F_2 with 4-5 long dark *pv* setae; very small species (1.0-1.5 mm). ..... ***sylvicola*** **(Collin)**

- Antenna yellowish, at least on vassal segments (basal segments sometimes dark yellowish brown in *commutatus*); F_2 without *pv* setae behind the double row of ventral black spine-like bristles; rather larger species 1.8-2.5 mm. **3**

3 Antenna entirely yellow; pleura polished; *humeral* and *acr* setae absent. ***alter*** **(Collin)**

- Antenna with postpedicel dark; pleura dusted but katepisternum with polished patch at middle; humeral and acr setae present. **4**

4 Antenna with postpedicel long, ≥ 2X long as deep, stylus about as long. **5**

- Antenna with postpedicel shorter, only slightly longer than deep, stylus c 2X as long [not British]. **6**

5 Legs yellow (including 5th tarsal segment); antenna with postpedicel slightly more than 2X long as deep and stylus hardly as long; *acr* 3-4 serial, small. ..... ***unicus*** **(Collin)**

- Legs yellow but distal tarsal segments somewhat brownish, 5th segment almost black; antenna with postpedicel 2.5-3X long as deep, stylus of similar length, basal segments sometimes darkened; acr 2-serial, distinct, hair-like. ..... ***commutatus*** **(Strobl)**

6 At least 5th tarsal segment dark; F_2 long, slender, as deep as F_1; *acr* regularly 2-serial. ..... ***boreoalpinus*** **Frey** [Not British]

- Tarsi entirely yellow; F_2 short, stout, deeper than F_1; *acr* irregularly 2-4 serial. ..... ***alpinus*** **Chvála** [Not British]

7 F_2 lacking *pv* setae behind double ventral row of short spine-like bristles. **8**

- F_2 with dark *pv* setae. **9**

8 Scutum polished black (at most very thinly dusted); antenna with basal segments pale. ..... ***longicornis*** **(Mg.)**

- Scutum dark grey dusted; antenna with basal segments blackish. ..... ***brachystylus*** **(Bezzi)** ..... [not yet confirmed as British].

9 C_1 yellow, only base slightly darkened; T_3 in ♂ simple; F_2 in ♀ at most dark dorsally on distal half. ..... ***nigritarsis*** **(Fallén)**

- C_1 blackish, somewhat paler apically; T_3 in ♂ conspicuously curved medially with ventral excision at middle and swollen near base; F_2 in ♀ dark dorsally along entire length. ..... ***excisus*** **(Becker)**

KEY I. Species with black thorax, two pairs of vertical setae, scutum entirely dusted (if only thinly)

1 T_2 without apical spur (or spur very small); antenna with basal segments often yellowish. **2**

- T_2 with long sharply pointed apical spur; antenna with basal segments blackish. **8**

2 *Acr* 2 serial; F_2 ≤ width of F_1. ..... **3**

- *Acr* 3-4 serial or more; F_2 > width of F_1. ..... **6**

3 Antenna with postpedicel very short, hardly longer than deep; small species 1.2-1.8 mm. ..... ***nanus*** **(Oldenberg)** [Not British]

- Antenna with postpedicel longer ≥ 3X long as wide. **4**

4 Antenna very long, postpedicel ≥ 5X long as deep, stylus c0.25X as long; legs dark yellow, tibiae brownish, tarsi almost black. ..... ***brachystylus* (Bezzi)**
[probably British as specimens collected by Cole determined as this sp. by Chvála]

- Antenna shorter; postpedicel ≤ 4X long as deep; legs yellow, tarsi darker apically. **5**

5 Antenna with postpedicel c4X long as deep, stylus c0.5X as long. ..... ***luteicornis* (Mg.)**

- Antenna with postpedicel c3X long as deep, stylus about as long. ..... ***tuomikoskii* (Chvála)**

6 Legs yellow but tarsal segments 4 and 5 faintly brownish; T_2 with small apical spur. ..... ***pulicarius* (Mg.)**

- At least C_2 and C_3 darkened at base; tibial spur hardly present on T_2. ..... **7**

7 C_2 and C_3 strongly darkened; antenna with stylus 2X as long as postpedicel. ..... ***incertus* (Collin)**

- C_2 and C_3 less strongly darkened; antenna with stylus 1.5X as long as postpedicel. ..... ***vegrandis* Frey** [Not British]

8 *Acr* 4-6 serial (at least anteriorly), on broad neduan stripe. **9**

- *Acr* 2-serial throughout. **12**

9 Larger thoracic setae yellow; legs extensively yellow including mid and hind coxae. ..... ***maculipes* (Mg.)**

- Larger thoracic setae black; at least C_2, C_3, F_2 & C_3 extensively darkened. **10**

10 T_2 with apical spur rather short (≤ 1X long as limb is deep); legs darker with paler parts brownish rather than yellow, F1 black on at least basal 0.5; small species (1.5-1.8 mm). ..... ***celer* (Mg.)** [Not British]

- T_2 with apical spur sharply pointed, long (obviously longer than limb is deep); legs with contrasting yellow and black markings; larger (1.7-2.6 mm). **11**

11 *Acr* 4-serial; C_1 yellow; C_2, C_3, most of F_2 and F_3 on distal 0.5 black. ..... ***rapidus* (Mg.)**

- *Acr* irregularly 6-serial; legs more extensively black, F_1 with only tip yellow and F_3 almost entirely black. ..... ***rapidoides* Chvála**

12 Large thoracic setae brownish. ....***bilobatus* Wéber** (♀)

- Large thoracic bristles black. **13**

13 Thoracic pleura subshining, katepisternum and anepimeron polished black; small (2 mm). ..... ***pygmaeus* (Mg.)**

- Thoracic pleura dusted except polished katepisternum. **14**

14 Postpronotal lobe [humerus] more or less shining on outer face. Legs yellow (♂) or extensively darkened (♀); antenna with postpedicel 1.5X long as deep, stylus 1.5X as long; larger (2.4-3.6 mm). ..... ***agilis* (Mg.)**

- Postpronotal lobe [humerus] dusted; relative proportions of postpedicel and stylus otherwise; smaller (2-3 mm). **15**

15 Antenna with postpedicel 2.5X long as deep, stylus about same length; *acr* pale; larger thoracic setae blackish (♂) or brownish(♀); femora extensively darkened with paler tip (♂) but F_2 and F_3 blackish only apically in ♀. ..... ***bilobatus* Wéber**

- Antenna with postpedicel 1.5X long as deep, stylus c2X as long or more; *acr* black; larger thoracic setae always black; legs yellow in both sexes with dark dorsal strip on F_2 and distal 0.5 of F_3 black. .... ***pseudorapidus* Kovalev** [Not British]

Platypalpus luteolus ©Adrian Plant

Plate 16,5 from A Dipterists Handbook. *Platypalpus major* Photo: Adrian Plant

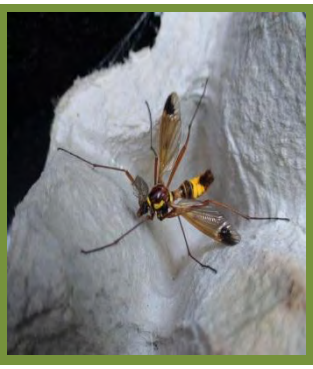
Cranefly News

Dipterists Forum Cranefly Recording Scheme

For Superfamily Tipuloidea& Families Ptychopteridae & Trichoceridae

Newsletter No 23 **Spring 2012**

Editor: John Kramer Layout: John Dobson



Ctenophora ornata (Keith Godfrey)

Field Work 2011(See also Issue 22)

A good year for Ctenophorines

2011 was certainly a good year for the Ctenophorines. Perhaps the unusual weather over the Winter and Spring played a part, or perhaps not (more phenology research is needed; anyone?). Judy Webb recorded four Ctenophorines: *Ctenophora pectinicornis*, *C. flaveolata*, *Tanyptera atrata and T. nigricornis* from a beech wood near Ipsden in the Chilterns in May. Six *C. flaveolata* were seen flying there on the same day. Reports also came in from other recorders. If you also had a record, please make sure that you send it in.

Cranefly news from Shropshire.

The highlight of the Spring from VC40 that didn't make it into my Shropshire update in Cranefly News #22 was the discovery of *Molophilus niger* G in G & T, 1920, new to the county from the Borle Brook near Highley, a few miles north of the Wyre Forest. I visited the site in late April to reccy it for a summer Invertebrate Challenge field day and came across this diminutive dark small fly whilst sweeping over log-jams. The site offers much potential and I plan to go back and look for *Lipsothrix* craneflies there next May. On the field trip in July the highlight was probably *Helius flavus* (Walker, 1856) on a day when craneflies were fairly hard to come by.

Elsewhere Nigel Jones and I did some work for the Shropshire Environmental Data Network (SEDN), our virtual local record centre, and searched for flies around the Meres and Mosses of north Shropshire and upland flushes around the Long Mynd. Craneflies of interest around the Meres and Mosses were *Metalimnobia quadrinotata* (Meigen, 1818) from the Marl Allotments by Fenn's, Whixall & Bettisfield Mosses NNR, and *Prionocera turcica* (Fab. 1787) from Clarepool Moss NNR amongst more common species. Again as the summer progressed towards its climax, dry conditions made searching for flies more difficult.

The Long Mynd however was a little more productive with several interesting species recorded including *Molophilus occultus* de Meijere, 1918, *M. flavus* Goetghebuer, 1920 and *Phylidorea squalens* (Zetterstedt [1838]) regularly found at flushes. Elsewhere the only real highlight of the summer was *Diogma glabrata* (Meigen, 1818) from its second Shropshire location at Bury Ditches near Clun.

What started as a really promising year petered out somewhat as the summer season continued.

The 2007 Shropshire cranefly atlas text and maps can be downloaded as a PDF from the resources page at www.invertebrate-challenge.org.uk – it is hoped that an update of new county and uncommon species will be compiled and published on the website during 2012/13.

Pete Boardman
Cranefly recorder for Shropshire VC40

Local Lists

Shropshire

Peter Boardman continues to do very useful work in Shropshire and his list of records for the year also included *Nephrotoma analis*, which is not common, from near the Discovery Centre, at Craven Arms. *Hoplolabis areolata*, a rare fly of sandy river banks, and *Paradelphomyia dalei,* were also very good finds from Downton Gorge NNR. *Ctenophora pectinicornis* had a good year in Shropshire, as in other areas. *Tipula rufina* was also recorded; records for *T. rufina* seem to be falling, perhaps because it emerges late, when many dipterists have ceased collecting for the year. Other species worthy of note, apart from those mentioned by Peter, were *Antocha vitripennis* and *Molophilus curvatus*.

Leicestershire

After some digging in the Archives, I have produced another updated list of the Craneflies of Leicestershire, now standing at 146 species. As with all of these lists, it will form a very useful basis for further work, and perhaps an Atlas for VC 55 is the next step. The list has been published by the Leicestershire and Rutland Entomological Society as an Occasional Publication (LESOP). I have sent out a few pdfs to those who I know might use them. Please let me know if you want one, to compare with your own County Cranefly List. The LES will soon have a website from where all of the publications can be downloaded.

Contents

Pipunculid Parasites of Craneflies

The larvae of most of the species of the family Pipunculidae (Diptera) are parasitoids on various species of bugs (Hemiptera). In 1966 only one species of *Nephrocerus* (Pipunculidae) had been discovered in Britain, and that was *Nephrocerus flavicornis*. In his 1966 RES Key Coe describes the 'considerable speculation' as to its hemipteran host. It was assumed that it was a bug, and since the fly is large, then, they speculated, the bug must also be large. In 1980 Alan Stubbs added *Nephrocerus scutellatus* to the British list, but there were still no clues as to the host.

The relationship was first discovered in 2007 by Koenig & Young who reared infested hosts, not of Hemiptera but of five species of *Tipula* (*Lunatipula*) and *Tipula* (*Yamatotipula*).

Kehlmaier and Floren have recently published an interesting paper about Pipunculidae in the Bialowieza Forest in Poland (1). In that paper they report the occurrence of three larvae of *Nephrocerus flavicornis* (Pipunculidae) in three female adults of *Tipula* (*Beringotipula) unca* (Tipulidae), and also one larva of *N. scutellatus* from a female *T.* (*Lunatipula*) *helvola.*

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John Kramer

Cranefly References

A Message from Pjotr Oosterbroek, Amsterdam

Dear friends and colleagues,

Almost all Alexander's PDFs are now available for easy download from the literature section of the CCW, with thanks to Sigitas Podenas who shared some 90% of them. In total 1042 of Alexander PDF's are available. Missing only are 4 obituaries, the Neotropical catalogue (PDF in preparation by Guilherme Ribeiro) and the Oriental catalogue (PDF in preparation by Herman de Jong's team).

Thanks to Dmitry Gravyushin and Vladimir Lantsov, more than 60 Savchenko PDFs have become available as well, including large ones such as Fauna USSR. You can also search for papers by Oosterbroek (106 papers, 48 with PDF's).

The references in the Citations parts are linked to the reference database. If you click on one of them, the reference is specified, incl. a PDF button if the PDF is available.

Try Tipula helvola; go to the distribution citations for Spain and select Oosterbroek, 2009c. From the page with this reference you can download the PDF immediately.

Have fun.
All the best,
Pjotr Oosterbroek

Illustrated Catalogue of the Craneflies of the World (CCW). Updated 30th November 2011.
Over 15,300 species incl. distributions, citations, illustrations.
PDF's. Online at: http://nlbif.eti.uva.nl/ccw/

This is very good news as many papers, especially those by E. N. Savchenko have good drawings of the male genitalia. It is an excellent resource, so thanks to all involved.

The 2012 Season

We have a good spread of cranefly recorders over the country, so hopefully we will get a good spread of records from everyone at the end of the 2012 season. We can use the records to see if there are changes nationally in the area occupied by a species, and may get some idea of the changes in numbers. Some of you may be part of the Butterfly Conservation teams that do the walked transects to study changes in abundance of your local butterflies. It is a good idea, and if there is a nature reserve near you, you may be able to monitor changes in numbers of craneflies. Some families are good flyers and Malaise traps are useful. My hunch is the craneflies fly much more at night, in the absence of drying sunlight, so any traps which can catch them at night may reveal some new species at a site, as well as give a more accurate idea of numbers flying.

Members of the Northants Group enjoy an Autumn workshop organised by John Showers; examining some of their annual catch of craneflies at Holcote Lodge, the Anglian Water Interpretative Centre, at Pitsford Reservoir.

This is a good way to identify your samples, so if you want to organise something similar for your local group, in your local centre, I can probably organise a time to visit and help. An 'Introduction to Craneflies' course is also available. If you are

organising your programme for 2012, a Cranefly Workshop might be an interesting possibility.

John Kramer

Population Explosion of Autumn Daddy long-legs - *Tipula paludosa*

In March this year I had a mail from Roger Payne, who works at the Southend-on-Sea Museum.

"*A few days ago, (Mar 14th 2011) I received about 50 cranefly larvae which I identified as probably T. paludosa from the groundsman at Belfairs High School in Southend. These had become trapped on hard concrete standing between the school and the playing field. This is not an unusual phenomenon and I have had this enquiry before. Larvae are usually washed out of the soil after rain and become trapped on the hard surface unable to burrow.*

The groundsman informed me that last year (Autumn 2010) they experienced the largest numbers of craneflies that he had seen in 7 years of working at the school. He said the school walls were covered in craneflies and had photographs (see below).

In view of Alan Stubbs article, 'The dog that did not bark' in Dipterists Forum Bulletin, Spring 2011, where he discusses the apparent absence of this species in large numbers last year, I thought you may find this interesting and am enclosing the picture of a large mass of craneflies."

Tipula Records Needed

Are flies in the genus *Tipula* (family Tipulidae, 'daddy long-legs') getting scarcer?

There are 61 species on the current British check-list and, despite the previous item, 29 of these have shown a decline in the number of hectads from where they are recorded, over the past 30 years. It is probable that this has more to do with changes in sampling effort, or a temporary fluctuation, than with a long-term trend in species numbers. If we can increase the recording effort and the numbers still show a decline, it will support the idea that something is happening.

The larvae of all of these species prefer moist conditions. Some live beneath mosses, while others live in peaty or marshy ground. Reasons for a possible decline may be linked with this.

Some species, such as *Tipula unca*, *T. pagana*, and *T. scripta*, are ones that you will find locally. Others are species of woodland (*T. signata*, *T. staegeri*), upland (*T. montana, T. subnodicornis*) or moorland species. Please have a look in your 'Provisional Atlas of the long-palped craneflies' by Alan Stubbs and explore. If you are going to the mountains, collect along the borders of streams and in any sheltered area. If you know someone who is using a light to record nocturnal moths, see if you can beg the by-catch of craneflies.

As usual, let me know if you need any help; an Atlas, or the complete list of declining craneflies, although all records are useful. Specimens can be sent to me for identification.

John Kramer

NB. The next copy deadline for Cranefly News will be July 15th. Why not focus on your local nature reserve and send in a report?

All correspondence through john.kramer@btinternet.com please.

Flowers for flies

J A Webb

I'm principally a botanist, but I'm very keen on flies and I have been looking into how to encourage them in a garden, considering that many flowers they depend on are declining in the general countryside these days. Many flies need to stock up on nectar for energy and of course some, especially hoverflies, need to feed up on protein-rich pollen to produce eggs. Thus flowers for flies do not always need to be ones which produce lots of nectar, often big pollen-producers like some wind-pollinated plants, are favoured by pollen-gathering hoverflies (they even visit the anthers of grass and mugwort flowers plus catkins of wind pollinated trees like hazel, oak).

Eristalis tenax on ivy. Photo: Steve Woodward

Generally flowers for flies need to be:

- Open at the right time of year when the fly is on the wing (garden full of winter flowering shrubs is no good). The main garden hoverfly peaks tend to be in May and late July/early August, into September if the weather is favourable.
- Open and flat or with many flowers in a flat or globular head to provide a good landing platform (only bee flies with their long tongues can hover and feed at the same time).
- Have nectar exposed or down only a short flower corolla-tube. Flies do not, with exception of a few like bee flies (*Bombylius*) hoverflies (*Eristalis. Rhingia, Volucella*) tachinids (*Siphona*) conopids and some empids, have the long probosces necessary for drinking deeply hidden nectar in longer corolla-tube flowers (these are usually specialized for bees or moths)

So what can you do to feed flies in your own garden by selective planting? The Royal Horticultural Society has produced a 'Perfect for Pollinators' list (see ref.) which will give you lots of ideas, but my impression is that it is dictated by the needs of bees and butterflies. For flies, my researches and conversations with other Dipterists indicate that if space and aesthetics are no object, the simple answer would be hogweed, hogweed and yet more hogweed, but of course life is not that simple. In a garden, one needs acceptably attractive plants and of course small gardens cannot host such big tall 'thugs' in a border. It is also nice to have a succession of tasty things coming into flower throughout the year, rather than only in one month. One also needs to consider the type of soil in your garden, whether acid or alkaline, dry and free draining, or heavy and prone to water-logging and buy plants appropriately. Flowers for flies need to be in warm sunny positions to be useable, as they will not go to those in cool shade – especially important in spring. However if you do have some semi-shade and want a fly food plant there, a carpet of enchanter's nightshade (*Circaea lutetiana*) will be just great for small hoverflies of shady situations like *Neoascia* and *Syritta.*

Phasia hemiptera (Tachinidae) enjoying hogweed (*Heracleum sphondylium*) Photo: Steve Woodward

Border Perennials

Fly-friendly small spring flowers for a border? Well it has got to be primroses, cowslips and lungwort (*Pulmonaria*) in full sun for the bee flies *Bombylius* and *Rhingia* hoverflies. These can also make use of spring bulb flowers such as grape hyacinth (*Muscari*) and bluebells. Anemones provide only pollen, but this may attract spring hoverflies. *Rhingia* can also use bugle (*Ajuga*) flowers and this can come in pretty variegated leaf ground cover varieties.

Rhingia campestris on red campion *Silene dioica*. Photo: Steve Woodward

Small perennials for the front of the border? Anything with open flowers in a cluster for easy landing and short corolla tubes with abundant nectar – sedums (stonecrops), marjoram, mints and thymes are good. Saxifrages of all sorts have small open starry flowers with exposed nectar that would enjoy such a position. For medium height in a hot sunny position, what about garden varieties of the common yarrow (*Achillea millefolium*). These come in every shade from the normal white flowers through sugar pink to yellow and are a good mid-summer nectar source used by flies.

Taller perennials for middle to back of a border? I suggest for later summer flowering hemp agrimony (*Eupatorium*) is a must (good for butterflies as well). Ornamental members of the Apiaceae (Umbelliferae) are a big draw. This family of plants is extremely important for flies in the wild (see 'Hoverflies of Surrey' by Roger Morris and see how often they are mentioned as being visited). If you don't fancy hogweed, what about tall, elegant, ferny-leaved fennel (*Foeniculum*) with its yellow flower clusters (and you can eat the leaves) also garden angelica is very pretty but ever so tall (wild angelica is almost as pretty and much shorter). If you want to go for the really enormous, try *Ferula communis*, you will need binoculars to actually see if there are any flies on the flowers up high. Umbellifer (Apiaceae) family herbs like lovage, coriander, parsley and chervil, will all produce those flat plates of tiny white nectar-rich flowers that flies love to land on (if you don't eat all the leaves and let them actually flower). Alexanders (*Smyrnium olusatrum)* is a herb introduced by the Romans that is the first umbellifer to flower in spring for very early flies. Ivan Perry finds the most amazing variety of flies, including rare tachinid flies, on his bush of perennial 'shrubby hare's-ear' (*Bupleurum fruticosum***) in his garden. This is an umbellifer with large flower heads of tiny yellow-green flowers which rival hogweed for nectar production. Of course Ivan lives next to some wonderful habitats in Cambridgeshire that produce the exciting tachinids as they provide the plants that feed their Lepidoptera hosts, but you never know what special flies might be just round the corner in your neighbourhood looking for a good dinner.

Continuing thinking about the Apiaceae, if you have a largish garden, how about a patch of common cow parsley (*Anthriscus sylvestris)* for an early spring bite for the flies. If you can't have such a wild area why not look into a plant I'm very interested in trialling - a pretty pink garden variety of the greater burnet saxifrage, which is normally white-flowered and is used by flies in damp calcareous grasslands, but has become rare as a wild plant in my area. If you put its garden variety name '*Pimpinella major* Rosea' into any search engine, you will come up with photos and lots of suppliers of potted plants. Pretty blue sea holly (*Eryngium)* of all sorts seem very useful as well, later in summer. Here, rather surprisingly, I have a good word for that 'gardener's bane' plant known as ground elder (*Aegopodium*). It is a creeping perennial thug with rhizomes that are the very devil to eradicate from a border once it has a hold. However, as an umbellifer, it has heads of tiny white flowers very attractive to flies out in May. In my local fen in Oxford city, there are often no flies to be found at this time by sweeping the fen vegetation which is mostly rushes and sedges. All the flies breeding in the fen are to be found on the nectar-rich large patch of ground elder flowers on an adjacent drier bank (where they have spread out from previous dumping of waste garden rubbish). I read that one can buy a prettier variegated-leaved garden variety of ground elder which is less invasive. Worth a try in a confined area like a large tub, perhaps sunken in the border in a good sunny spot?

Chrysogaster solstitialis hoverflies enjoy ground elder (*Aegopodium podagraria*). Photo: Judy Webb

Flies in the wild go for knapweeds and thistles (*Centaurea* and *Cirsium* spp.). These are perfectly attractive plants with purple daisy-type flowers for growing in the garden. Don't forget that if you do that you may attract lovely tephritids like *Urophora* spp. to actually breed in the flower-heads. Big- flowered thistles like musk thistle or woolly thistle are attractive enough for a garden. Spear thistle is a biennial, so might be tolerated for one year on an allotment in a way that the perennial pest creeping thistle would not be. Sow thistles (*Sonchus* spp.) are also useful 'weeds'. Daisy-type flowers that are similar to the common wild oxeye daisy (*Leucanthemum*) are good (but why not have a mini meadow full

of buttercups and wild oxeye daisies in a sunny corner?). Plants in the Scabious family are also useful with attractive lilac/purple flowers (*Knautia* and *Scabiosa* spp.). Devil's-bit Scabious (*Succisa*) is a useful late summer flowerer, when the only other thing out in late summer to early autumn are michaelmas daisies (*Aster* spp.). These last are useful for flies, -but I cannot bring myself to plant them, as I spend such a lot of time pulling them out of my local nature area where they are thugs, having escaped from gardens and are romping away in a monoculture, excluding native, useful, earlier-flowering plants. Good for a garden where they can be controlled more easily. The yellow-green flowered spurges (*Euphorbia* spp.) are sometimes used as ground cover. There are short and tall flowered versions, but all have open flowers with exposed nectar and are used by hoverflies. Mallows, Lavateras and hollyhocks (*Alcea*) produce such an abundance of pollen in open flowers they must surely be useful to pollen-consuming hoverflies, but I have no observations on this and would welcome input from other people with views on these plants.

In wild habitats there is a 'nectar gap' in late July/August which is admirably filled for flies by that often reviled plant ragwort (several *Senecio* sp). Nothing to stop you having plenty of ragwort in your garden in your mini-meadow if you want! The daisy bush from New Zealand (*Olearia haastii)* flowers exactly in that August gap and I wonder if it is popular with flies? Personally I quite like the alien Oxford Ragwort (*Senecio squalidus)* as a sunny border plant as it has bigger flowers and starts flowering earlier than other ragworts, here in Oxon as early as April and certainly by May.

Syritta pipiens on ragwort (*Senecio* sp.) Photo: Steve Woodward.

Garden Ponds

Even if your pond is very small, how about some water plantain (*Alisma plantago-aquatica*) as a tall emergent and some floating frogbit (*Hydrocharis morsus-ranae*). Their open small 3-petalled white flowers are both visited by hoverflies and ephydrids. What about my personal favourite pretty emergents – the arrowhead (*Sagittaria*) with large white open flowers and the flowering rush (*Butomus*) with clusters of open 3-petalled pink flowers. Marsh Marigold (*Caltha*) with its large buttercup-style flowers would also be good for the pond edge along with the water forget-me-not (*Myosotis scorpioides*). Creeping jenny (*Lysimachia nummularia*) perhaps in the garden golden-leaved version, could creep over the damp paving around such a pond and if in sun, will produce abundant open yellow flowers used by flies. Common valerian (*Valeriana officinalis*) or marsh valerian (*V. dioica)* will attract flies in any marginal marsh/bog garden, but the all-time winners for such a positions have to be wild angelica (as good as hogweed) and water mint (*Mentha aquatica*) with lilac flower heads late in the year - loved by all sorts of flies, perhaps combined with the cheerful yellow button-shaped daisy-type flower heads of fleabane (*Pulicaria*). Meadowsweet (*Filipendula*) will supply abundant pollen but no nectar. Fool's water-cress (*Apium nodiflorum*) is an umbellifer much used by flies in wild ditches and ponds, so nothing to stop you putting it in your garden pond along with white flowered water-cress from the brassica family. All the dolichopodids and ephydrids from your pond will love those.

Every pond should have water mint (*Mentha aquatica*) [top] and fleabane (*Pulicaria dysenterica*) with the Marmalade fly *Episyrphus balteatus*. Photos: Judy Webb

Allotments or Vegetable Gardens

On the allotment, why not allow some of your un-harvested carrots, celery and parsnips to grow up and flower ?– these umbelliferous plants have flowers that are some of the most favoured by flies in wild habitats. As these are biennials, if you want more instant results in one year, why not buy some carrots with green tops in a shop and actually plant them out in a border – they will then grow flowers that year. Un-harvested cabbage, broccoli or cauliflower should be left to go to seed where the yellow four-petalled flowers can be used by flies. A clump of chives produces abundant spherical purple flower heads that are much used and what about a patch of wild ramsons with their white globular flowerheads for flies and delicate garlic-scented leaves for salads? Do you have a bindweed problem on the allotment? Perhaps don't eradicate it all, but leave some in a hot sunny portion where it will flower abundantly with those pretty pink trumpets and attract hoverflies. How about leaving the poppies that pop up as weeds? These produce only pollen (no nectar) but the pollen is abundant as a reward to pollinators, so they will be good for hoverflies which can be attracted to lay eggs on your crop plants and then their larvae can usefully consume lots of greenfly and blackfly. The best plant sold as a hoverfly-attractant for allotments or for organic farming to control aphids is the scorpion weed, *Phacelia tanacetifolia*. This is a tall annual, flowering continuously from July to September in with curled racemes of pretty blue-lilac flowers. Once the hoverflies are attracted to this plant they are very likely to lay eggs on the aphid-infested crop plants nearby and their larvae will consume the pests. I'm sure it will be good for other flies as well as hovers, but I would welcome some feed-back on this from gardeners. If you have room for soft fruit, blackberries have flowers much used by flies in mid-summer and red currants and black currants have small green flowers useful for early flies.

Scorpion weed (*Phacelia tanacetifolia*) a favourite with hoverflies. Photo: Judy Webb

Annuals for the edge of a sunny border or corner of an allotment? All sorts of weeds of the scented and scentless mayweed type of open daisy flower (*Tripleurospermum* & *Matricaria* spp.) should be left. Feverfew (*Tanacetum parthenium*) is also very good, along with annual candytuft (*Iberis*). Forget-me-nots (*Myosotis* spp.) are pretty and attract hoverflies like *Platycheirus* and *Syritta*. In full sun the proper perennial chamomile (*Chamaemelum nobile*) is useful as it has attractive grey-green scented leaves and white daisy flowers which can be used for herbal tea (or preferably leave them for flies).

Lawns

What about a fly-friendly lawn? Well, I have been trying that for some years. A mix of native species with the mowing relaxed a little at flowering time is working quite well. Yes, encourage celandines and buttercups of all sorts *(Ranunculus* sp for e.g. *Cheilosia* hoverflies) and common daisies and dandelions, but what I have also added are mouse ear hawkweeds (*Pilosella officinarum)*. This spreads by runners, loves mowing and produces a flush of lemon yellow dandelion type flowers in early summer. It is very drought tolerant so good for a hot dry lawn. Also good in this situation are the other dandelion look-alikes of cat's ear and various hawkbits (*Hypochoeris* and *Leontodon*) especially the late summer autumn hawkbit, *Leontodon autumnale.* Plantains are thought of unattractive flowers that are wind pollinated, but the hoary plantain (*Plantago media)* is scented and has flower spikes with attractive lavender-coloured filaments to the anthers, so is designed for insects. I have seen hoverflies feeding on the pollen. Some speedwells like lawns and flowers of the germander speedwell (*Veronica chamaedrys*) is liked by hoverflies e.g. *Baccha* and *Melanostoma* spp. I'm hoping to be able to introduce the meadow saxifrage (*Saxifraga granulata*) to the lawn in future years and see what likes that. All these lawn things will need the mowing relaxed for a bit around June to flower abundantly, but will survive regular mowing at all other times.

Shrubs

Shrubs I have found to have flowers very attractive to flies include dogwood (*Cornus* spp.) and hawthorn (*Crataegus* spp. but not the ornamental double-flowered, go for the wild type). Privet and elder flowers are used but don't seem so popular and my impression is that there are a lot of common garden shrubs that are not useful, for instance – oleaster (*Elaeagnus,* flowers in winter) japonica, kerrya, snowberry, forsythia, berberis, hydrangea, griselinia, hebe, aucuba, escallonia, fuchsia, buddleia …there are loads. It is not that these are completely unused, maybe some people have seen the odd fly on e.g. buddleia (especially big hoverflies) it is more that they are bulky and take up a lot of space, which in a small garden would be better used for a really good fly-friendly shrub (like the shrubby hare's-ear mentioned previously). Rhododendron and azalea flowers can be visited by flies, but are not hugely attractive. Think about these that follow. Why not find room for a couple of small native spindle (*Euonymus europaeus*)? This has a profusion of small green flowers with exposed nectar for flies and you have the benefit of the attractive pink/orange berries later for birds. Other white and yellow variegated *Euonymus fortuneii* shrubs used for ground cover have similar flowers so I expect them to be useful as well. Small yellow-flowered members of the rose family in the genus *Potentilla*, like tormentil, silverweed and cinquefoil are used by flies in the wild, so shrubby Potentillas (varieties of *Potentilla fruticosa*) are useful in a garden context. Other useful garden

shrubs are species of *Viburnum*, *Cotoneaster* and *Pyracantha* and shrubby yellow-flowered relatives of ragwort *(Brachyglottis).* Single roses can be useful, but fully double roses are of no use to any insect*. Rosemary and lavender seem unattractive as they are definitely bee-plants. Shrubby St John's worts (*Hypericum*) don't seem much good and especially not that ground cover one with the fibre-optic lamp stamens known as Rose of Sharon (*H. calycinum*). If anyone thinks I'm I'm wrong, please let me know. Also let me know if you have a really good shrub for flies.

Volucella pellucens on guelder rose (*Viburnum opulus*). Photo: Steve Woodward

Trees

What trees are best for flies? If you are talking about early to midsummer flowering my answer would be lime, lime and yet more lime – native small-leaved lime or large-leaved lime (*Tilia* spp.). If you could manage one tree of each species, there will be abundant nectar and pollen from mid-June (large-leaved lime flowers first) into July (small-leaved lime). You can extend the dinner for flies if you plant the later-flowering Crimean lime (*Tilia euchlora,* which flowers later in July). Sweeping the accessible regions of three species of lime flowers in my local nature park has produced the most amazing variety of flies, with rare hoverflies like *Criorhina* spp. (must be some good rot holes nearby) and hordes of tiny hybotids in the genus *Platypalpus* (that is if you can fight your way past the numerous bees, wasps and beetles that are also feasting on the flowers). Bee-keepers note that the lime is known as the 'honey-tree'!

Common Lime (*Tilia x europaea*) Photo: Steve Woodward

At other times of the year one can feed flies by planting trees of single-flowered (**not double***) *Prunus* species – blackthorn, plum, cherry and cherry plum for early hoverflies and bee flies. Later, apples (*Malus* spp.) pears (*Pyrus* spp.) and rowans and service trees (*Sorbus* spp.) are all useful to some degree. Maples (*Acers*) like field maple, sycamore and Norway maple produce yellow-green flowers with abundance of nectar in spring to early summer that are highly attractive to flies. Early spring-flowering willows (*Salix* spp.) produce catkins with abundant nectar (and pollen, but **only** on the **male** trees) which can be extremely important for early flies – if you have room only for one, how about a small male pussy willow in a sunlit corner? – can be pruned to keep it small. A later spring-flowerer that is useful to flies is the holly tree (choose male or hermaphrodite versions to make sure pollen available).

Bombylius major enjoys single wild cherry flowers. Photo: Judy Webb

One interesting thing about some trees and other plants is that they have so much attack by sap-sucking aphids (e.g. blackfly and greenfly) that there is usually a good coating of honeydew actually on the leaves. This is especially noticeable with sycamore and lime trees (don't plant a lime tree over where you park your car). This honeydew is dried sugary secretion of the aphids and attracts a lot of flies. I think it may be very important in areas or at times of year where there are not many suitable flowers. I have seen flies attracted to nettle leaves and when I looked closely, the nettles were covered in aphids and the flies were feeding on the honeydew drops which had accumulated on the leaves just below the aphid colonies. Adults of the fungus associated platypezid flies never visit flowers, but spend a lot of time rushing about on leaves and feeding on honeydew there. Perhaps one could help flies by leaving aphid infestations alone to generate honeydew and putting the insecticide spray away!

Climbers

This is simple – the best has got to be ivy, but not in that boring non-flowering evergreen vegetative sheet version that covers shady borders and is a thug that excludes more useful plants, **but** the excellent sort of ivy that is allowed to climb up a wall or fence in the light so it can **flower** – we all know how important ivy flowers are to late summer flies (and butterflies like red admirals) as it has little green flowers with exposed nectar drops. It really does not matter what type of ivy one has – all manner of variegated and interesting leaf shape ornamentals will do, the important thing

being that they are allowed to get up into the light and **flower** (non-flowering ivy can actually be a suffocating thug in a border, see below). For a small garden I would advise against the very large rampant sorts like *Hedera colchica* and go for a more delicate small-leaved ivy that is less work to control each year and get it growing up something, not covering the ground. One must not forget the benefit a fence covered in ivy is in terms of hibernation sites for insects and nest sites for birds. What about other climbers? Honeysuckles and jasmines have the nectar down too long a tube for the generally shorter tongues of flies to get at (these flowers are good for moths). Wisteria is bee adapted and I've never seen any flies using spectacular passion flower (*Passiflora*) but the small unspectacular, greenish, open flowers of Virginia creeper or Boston ivy (*Parthenocissus* sp) and the vine (*Vitis vinifera*) certainly look to be fly-adapted. How about an ornamental grapevine for flowers and fruit? *Clematis* spp. of all sorts have open flowers with abundant pollen, so they may be attractive to hoverflies, but I have only noted these on the flowers of wild clematis.

Myathropa florea on Ivy (*Hedera helix*) flowers. Photo: Steve Woodward

Time for a change?

What border flowers take up space and are not good for flies? Well, anything that is mainly adapted for other pollinators like bees and moths i.e. tobacco (*Nicotiana*) with its really long corolla tubes. Flowers in the Fabaceae (pea/bean/vetch/clover) family have specialized closed flowers that are generally in need of the strong arm tactics which only bees and wasps can apply to prize open the petals to get at the nectar. Most weak-armed, delicate-bodied flies cannot open up these flowers like strong bees can (with the noted exception of a few largish hoverflies). Thus the following garden flowers are pretty useless as fly-attractants: lupins, sweet peas, everlasting sweet pea, broom, gorse, laburnum, snapdragon, toadflax, foxglove, sidalcea, delphinium, monk's hood (*Aconitum*) and others. Flowers in the dead-nettle family (Lamiaceae) are more commonly bee-adapted with long corolla tubes and nectar thus out of reach (except for the likes of *Rhingia* and those already mentioned). The really long-corolla ones like Sage and other Salvias, woundworts (*Stachys*) Jerusalem Sage (*Phlomis*) obviously cannot be used, but short-tubed mint and oregano (marjoram) are great for flies. Also not useful are garden busy lizzies, although other members of the family like balsams (*Impatiens*) can be used by some flies, including the alien plant Himalayan Balsam, which is romping away and changing so many of our river corridors and wetlands. Please don't grow that though, as it may spread from your garden to the wild and cause problems. Begonias of all sorts are useless. Campions seem used only by the likes of *Rhingia* and other pinks and carnations are equally unattractive, along with irises and most of the lily family. Ericaceous things like bell-heathers (*Erica*) and *Pieris* are not much good but wild ling (*Calluna vulgaris*) is enjoyed by flies in summer. Large garden pelargoniums seem useless, but small wild versions of the geranium family like herb robert (*Geranium robertianum*) are used. I've never seen any fly using the big-flowered or open-flowered bell flowers, like Canterbury bells (*Campanulas*). Trees that are not very attractive (except to some early hoverflies) include all the wind-pollinated sorts with no nectar such as ash, hazel, beech, oak, birch, alder.

So what would the worst garden for fly foods be? Full of those easy-maintenance conifers that are prostrate, evergreen mats, or tall cupressus 'Leylandii' hedges casting dense shade, with only winter-flowering heathers, *Ericas* and similar plants. Or with mainly ferns, shaded, non-flowering ivy mats and other evergreen ground cover like rose of Sharon. I see plenty of 'low maintenance' borders like this in municipal plantings and despair for flies and other insects. Beds can be full of shrubs that flower only in winter, when no flies are around like *Viburnum tinus* and oleaster (*Elaeagnus*) or wind pollinated things like sea buckthorn (*Hippophae*). Or one could even have a very pretty flowery garden full of only bee- or moth-adapted flowers and not realize you are starving the flies.

No insects enjoy double cherry flowers! Photo: Judy Webb

Away with all that kind of plant and get in some good fly-food plants instead when you take your spring trip to the garden centre! Also if you find any plant in your garden that has flowers that are a real winner with flies, I would be very pleased if you would let me know. I have just received a packet of *Bupleurum fruticosum* seeds as a birthday present, so I'm looking forward to growing them and conducting the Oxfordshire 'Tachinid attraction trial'! More information on flower visiting by flies is to be found in the latest edition of the Dipterists Handbook in the article by Martin Speight and if you are interested in pollination and pollinators of

all sorts, I recommend the New Naturalist book on this topic by Proctor, Yeo and Lack.

*Double flowers of all sorts are where the extra petals in the middle are modified stamens that no longer produce pollen. Thus they are poor pollen-food for flies, they also do not produce as much nectar as the single flower versions.
** Thompson & Morgan stock the seeds, but plants available in garden centres

References

Morris, R. K. A, (1998) Hoverflies of Surrey, Surrey Wildlife Trust

Proctor, M, Yeo, P. & Lack, A. (1996) The Natural History of Pollination, Collins New Naturalist Series No 83

RHS Perfect for Pollinators list http://besthabitats.com/wp-content/uploads/2011/05/RHS_Polinators_PlantList_V1.pdf

Speight, M D. 'Flower-visiting Flies' in Dipterists Handbook, (2nd Ed.) 2010, Edited by P J Chandler

Stratiomys chameleon on Angelica

Judy Webb
Email: judy.webb@virgin.net

Field Meetings Dipterists Forum

Booking Form - for rates see Bulletin

Meeting location and dates			
Name			
Address			
Telephone number			
Mobile phone number			
email address			
Intended stay (please indicate days and dates)			
Dietary requirements	Omnivore		Please tick relevant box
	Vegetarian		
	Vegan		
Allergies (food)			
Deposit			
Signature			Date

Please Note: We will endeavour to accommodate for part-weeks but this is dependent upon available accommodation and the policy of the host venue

Payment details:
Cheques made payable to Dipterists Forum

Deposits
Deposits will only be returnable if cancellation occurs before the published cut-off date for reduced rates.

Please send your booking form and cheques to:
Roger Morris
7 Vine Street, Stamford, Lincolnshire PE9 1QE
Email: roger.morris@dslpipex.com

Guidelines

Booking your place at events

Dipterists Forum events

In the past, I took personal responsibility for the finances of the meetings and the necessary guarantees of payment. This has caused problems however. For example, I frequently made deposits amounting to up to 20% of the total cost of the meeting and am no longer in a sufficiently strong financial position to underwrite meetings. In addition, I was also liable if anything went wrong (as we had at Swansea when I was threatened with legal action because the college messed up their records of payments!). Moreover, if my bank account was scrutinised for additional income, the deposits and payments might be regarded as income by the Inland Revenue and I might therefore be liable to tax of this money (incidentally participants have only been charged for actual costs and I have borne the administrative costs myself).

There was also a need to simplify the payment system to avoid the complications of past meetings where final costs were not known until the end of the trip when the bill arrived. High numbers of last minute changes made by members (cancellations and changes to duration of stay) have made the process of working out prices very difficult and vague until the last minute and have complicated administration considerably.

Roger Morris

Administration

The Committee have introduced a simplified system for payment. Firstly, the Forum is now responsible for paying deposits and for administering deposits by members. Secondly, a formal booking system is now established, with written records of members' intentions. A form is included within this bulletin and can also be downloaded from the website.

A 10% surcharge will be added to the price for bookings beyond a specified cut-off date. Cancellations before that date will also lead to return of the deposit, but after the date will be non-returnable.

How to book

Please complete the booking form, you can either copy the page later in this Bulletin or use the separate sheet.

Deposits payable to DIPTERISTS FORUM should therefore be sent together with the **booking form** to:

Roger Morris
7 Vine Street, Stamford
Lincolnshire PE9 1QE

Contributing Bulletin items

Text

1. Articles submitted should be in the form of a word-processed file either on disk (3.5", CD or USB Flash) or via E-mail which should have the phrase "DF Bulletin" in the Subject line. Email text alone will not be accepted.
2. Please submit in native format (http://en.wikipedia.org/wiki/Native_and_foreign_ format) and in "text-only" Rich Text Format (.rtf) and additionally send pictures in their original format. An accompanying print-out (or pdf) would also be useful.
3. Please note the width of the borders used in Dipterists Bulletin; for conformity with style would newsletter compilers please match this format.
4. **Do not** use "all capitals", underlining, blank lines between paragraphs, carriage returns in the middle of a sentence or double spaces.
5. **Do not include hyperlinks in your document. Since they serve no purpose in a printed document and the editor has to spend hours taking them out again (as the text is unformattable if it has a hyperlink attached), documents containing hyperlinks will be sent back to you with a request for you to remove them.**
6. Scientific names should be italicised throughout and emboldened only at the start of a paragraph.
7. Place names should have a grid reference.

Illustrations

8. Colour photographs are now used extensively in the Bulletin, they appear coloured only in the pdf or on the covers.
9. Please include all original illustrations with your articles. These **should** be suitably "cleaned up" (e.g. removal of partial boxes around distribution maps, removal of parts of adjacent figures from line illustrations) but please do not reduce their quality by resizing etc. .
10. Please indicate the subject of the picture so that a suitable caption may be included, in some cases it will be possible for the picture file's name to be changed to its caption (e.g. 049.jpg becomes Keepers Pond NN045678 12 Oct 2008.jpg). All group pictures should identify all the individuals portrayed.
11. Powerpoint files may be submitted, they are a useful means of showing your layout and pictures are easily extracted.
12. Pictures contained within Word files are of too low quality and cannot be extracted for use in the Bulletin.
13. Line artworks are also encouraged - especially cartoons
14. Colour pictures and illustrations will be printed in black and white (uncorrected) and so it would be wise to see what a B&W photocopy looks like first, although the print quality from Autumn 2009 onwards gave excellent B&W results.
15. A suitable colour photograph is sought for the front cover (and inside front cover) of every copy of the Bulletin, note that it must be an upright/portrait illustration and not an oblong/landscape one for the front cover.
16. Due to the short time-scales involved in production, the editors will not use any pictures where they consider there to be doubt concerning copyright.

Tables

17. Tables should be submitted in their original spreadsheet format (e.g. Excel)
18. Spreadsheet format is also appropriate for long lists

When to send (deadlines)

Spring bulletin

19. Aims to be on your doorstep before the end of February, the editorial team has very little time available during January and so would appreciate as many contributions as possible by the middle of December; the deadline for **perfect copy is the 31st Dec**, it will be printed then distributed in February in time for the March workshop meeting (which may by that time be fully booked). Please note that the date for contributions is now earlier than for previous Bulletins.

Autumn bulletin

20. Aims to be on your doorstep in mid September, contributions should therefore be made to the editor **by the end of July**. It will be printed then distributed in time for final notification of the Autumn field meeting (although you would be well advised to contact Roger Morris before this time and consult the DF website) and in time to provide details of the Annual Meeting. Please note that the date for contributions is now considerably earlier than for previous Bulletins

Where to send

21. Would Bulletin contributors please ensure that their items are sent to BOTH Darwyn Sumner and Judy Webb

A guide to help contributors with Word, pdf-making etc. is in preparation, at the moment it only contains a few tips but if you want to help with more tips for fellow Newsletter editors then the Bulletin editor would be pleased to hear from you. You can request the pdf now

And now ...

Putting flies in the soup

Our Conservation Officer, Rob Wolton, has suggested that we need a strap-line to say what we are about. Presumably he envisages a bold statement about the magnificence of flies, the worthiness of efforts to conserve them and the other purposes of the Dipterists Forum. The mind whirs as a plethora of thoughts fleetingly pass by.

So how do we say what we do? 'Fly-nets are Us' conveys our out-door image but is unlikely to create a rush to join us. 'A hovering we will go' sounds as if we are the dwarfs that Cinderella rejected.

We need bold statements. 'Two wings good, four wings bad' would make an impressive chant at the BENHS annual exhibition, very satisfying but very politically incorrect. 'The drum beat of halteres' could make a good DF anthem: as yet we need to devise a tune and I for one cannot sing. Indeed we might as well try to become the British entry in the European song contest; lots of buzzing would raise the standards no end, and with the novelty of flapping nets during the performance we would provide a memorable spectacle.

Why not latch onto the topics that are popular on TV? Loft conversions to house fly-collections, titled 'Flies Aloft': As starters for the series we could feature the largest such conversion in the World, at Melksham, Wiltshire. Food programs also gain huge audiences: 'How to make tasty biscuits' or 'The best ten tips in coating leatherjackets in chocolate' would gain DF publicity but that would not lead to more recording.

But Rob wants something more praiseworthy of the study of flies. But why bother? In essence we need something to which the public can relate. There is only one strap-line that qualifies:- 'Putting flies in the soup'.

Alan Stubbs

Criorhina floccosa mating pair Photo Paul Brock

Recording Schemes & Study Groups

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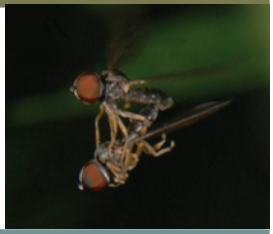

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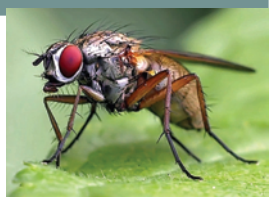
Anthomyiidae



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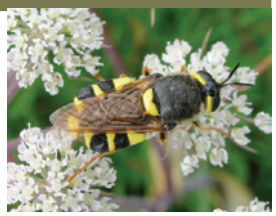

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Please note that Simon has stood down as organiser, we shall be announcing the new organiser shortly

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Whilst all schemes will readily accept records in written form the following symbols are used to indicate some of the known (or surmised) methods by which Scheme Organisers may currently receive records electronically:

Recorder Mapmate Excel Access and other databases & tools NBN Gateway, faded symbol = historic dataset