

Evergreen Vines in Deciduous Woods



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Provisional Preface

I have drafted this booklet during the winter of 2012–13 in order to focus attention on special problems posed by evergreen vines in deciduous woods. As invasive species, some of these vines thrust unwelcome urgent issues into the science of conservation. Understanding their general ecology helps to deal with these issues. As the reader will discover, this ecology appears much connected to mammalian herbivory—or lack thereof. I urge a broader approach to conservation of temperate forests in eastern North America, involving more consideration of intense seasonal browsing as a natural force. There has been excessive interest in burning for some habitats. I am particularly interested in stimulating more work on the restoration of eutrophic woodlands in the central Ohio Valley, where so little remains of the original ecosystem. Despite the pitiful condition of our remnants and difficulties in developing a coordinated effort among conservationists, some progress is possible. We do know that Virginian settlers established ‘woodland-pastures’ in more affluent areas, with the help of their slaves. The majestic old oaks and ashes of these sites accelerated growth and developed their large spreading limbs during the early decades of settlement—but during the 18th Century a more shady condition had prevailed over the landscape (Campbell 1989, McEwan & McArthur 2008). In recent years, conservationists have secured land with the best remnant of such woods (Griffith Woods in Harrison Co., Kentucky), but management remains unsettled (see p. 147–156). I aim to develop the framework of this booklet further with more information from these woods, plus extensions eventually into a general work on forest ecology in temperate regions. Much of this current draft will be tightened up after more analysis of plot data, better review of herbivory and secondary chemistry, deeper discussion of morphology—including sexuality (see p. 139–141)—and paleontology. I invite comment and collaboration.

**Evergreen Woody Vines in Deciduous Mid-Temperate Forests: a Review of Habit,
Habitat, Mammalian Herbivory, Secondary Chemistry and Biogeographic Context**
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Abstract. Prompted by local invasion of East Asian *Euonymus fortunei* (‘purple winter creeper’) into eutrophic woodlands of eastern U.S.A, this paper reviews the ecology of all evergreen vines in mid-temperate regions across the Northern Hemisphere (Hardiness Zones 6 and 7). Only about 15–20 species of moderate (3–5 m) to extreme (15–20+ m) height are largely evergreen—in *Bignonia*, *Celastrus*, *Euonymus*, *Hedera*, *Holboellia* and *Lonicera* and *Smilax*. Additional species have ‘semi-evergreen’ leaves that tend to senesce gradually during hard winters. Some of these evergreen-tending vines also cover much ground below deciduous trees. They are concentrated on nutrient-rich soils, in marked contrast to evergreen trees and shrubs—a trend that may be linked with a ‘contrarian’ strategy in these vines: to capture light during winter in deciduous forests. However, these plants are often eaten by mammalian herbivores, especially during winter. None are severely toxic, and woody vining taxa in general tend to have less defensive chemistry than their closest non-vining relatives. Diverse evidence suggests that large native herbivores and livestock reduce vines, as well as other undergrowth. It is suggested that, in eutrophic woodlands of humid mid-temperate regions before human influences, intense seasonal browsing was a major ecological factor and that it remains more important than burning for maintenance of native biological diversity. Biogeography and phylogeny indicate that genera with evergreen-tending vines diverged from their subtropical ancestors during mid- to late-Tertiary eras, long after deciduous genera first appeared in the fossil record. These vines grew into the expanding, drying, cooling deciduous biome—and proliferated locally—but became susceptible to browsing in winter by migrating megafauna.

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Front cover. Woods behind Henry Clay High School (he would be shocked), where *Euonymus fortunei* has taken over several acres; Layton Register to right, 6 feet tall. These woods used to form one of the best little remnants in the city of Lexington, Kentucky, with several uncommon wildflowers, including trout-lilies (*Erythronium* spp.) and Dutchman’s breeches (*Dicentra canadensis*). Much good could be done if youthful energy of the school was directed.

Intersections and back cover. Views of crossvine (*Bignonia capreolata*) in garden and wild. Although gone from many woods around town, it can be easily reestablished. It is probably the “jasmine” that Jessamine County was named after—then well-known and much-loved.

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Introduction

The ‘purple winter creeper’—*Euonymus fortunei* [= *E. hederaceus*, *E. radicans*]—is native to East Asia. But it has become widely promoted by the horticultural industry as an ‘ornamental’ ground-cover across the eastern U.S.A. during 1950-2000, together with closely related segregates or cultivars such as the non-climbing *E. kiautschovicus* (Graves 1940, Zouhar 2009). In warm- and mid-temperate zones on base-rich soils, *fortunei* climbs up trees, cliffs and walls, where it flowers and fruits, leading to widespread invasion of woods and thickets. Escapes initially occurred mostly in or near urban areas of east-central states, from Massachusetts to Missouri. They have now spread across most of the Ohio watershed and elsewhere (Kartesz 2012, SE-EPPC 2012). *E. fortunei* has become one of the most problematic invasive plants in woodland of central Kentucky, often dominating shady ground. But its degree of spread over the ground, versus up trees, varies much among sites (Campbell 2012).

Euonymus fortunei is one of only a few species of evergreen woody vines (lianas) that are native to mid-temperate regions of the Northern Hemisphere. Species of European ivy—*Hedera*—are the most well-known in traditional Anglo-European horticulture. Also, the popular evergreen to semi-evergreen *Lonicera japonica* from East Asia is now an abundant invasive species in much of eastern North America (Nuzzo 1997, Larson et al. 2006). The only widespread native vine with an pronounced evergreen tendency in mid-temperate regions of the eastern U.S.A is *Bignonia capreolata* (cross-vine). Other natives are just semi-evergreen, tending to loose some leaves in most winters: *L. sempervirens* and the greenbriars, *Smilax bona-nox* and *S. glauca*. *Decumaria barbara* (wood-vamp) is also semi-evergreen but extends only into warmer parts of mid-temperate zones (Duncan 1967).

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Such vines are largely restricted to warm- plus mid-temperate zones, as are many other broad-leaved evergreen trees and larger shrubs. These zones correspond to Heinrich Walter’s “moist warm temperate forest” plus warmer parts of his “deciduous temperate forest” (Breckle 2002). In North America (USDA 1990, 2012), they have often been referred to as Hardiness Zones 9 and 8 (warm-temperate) plus 7 and 6 (mid-temperate), with mean annual minimum temperatures of about -5° C to -10° C (warm-temperate) or -15° C to -20° C (mid-temperate).

The physiological basis of adaptation by evergreen plants to winter cold has been the subject of much research (e.g., Öquist & Huner 2003). Seasonal development of hardiness in *Hedera* involves complex changes in sugars, proteins and anthocyanins (Parker 1962, Bauer & Koffler 1987, Oberhuber & Bauer 1991). The protective role of xanthophylls has been shown in *Euonymus* (Adams & Demmig-Adams 1995, Verhoeven et al. 1998). But, despite the distinctive increase of purple coloration in much *E. fortunei* during winter, there is no published research on the possible role of anthocyanins in hardiness of this species. (Purplish winter coloration also occurs often in *B. capreolata*, *L. japonica* and *S. glauca*.) And even general understanding of differences among evergreen plants remains limited—what exactly allows the conifers to extend so much further north than most woody angiosperms? Their somewhat inexplicable supercooling ability must be a critical factor (Margesin et al. 2007, and their citations), with significant implications for secondary chemistry.

Meanwhile, refinements of the Hardiness Zone system are being developed (e.g., DeGaetano & Schulman 1990, McKenney et al. 2007, Daly et al. 2012), as well as more global and functional approaches (Magarey et al. 2008, Harrison et al. 2009). The combined effects of mean annual (overall) temperature, mean annual (extreme) minimum temperature, and precipitation patterns on vegetation still need much more analysis. For example, in East Asia,

relatively evergreen forest extends into zones with cooler mean annual temperatures than in North America and Europe. As noted by Walter and others, this contrast can be attributed to the generally less extreme minimums within temperate zones of East Asia (Wolfe 1987, Widrlechner 1997). In Eastern North America, standard deviation of the annual extreme minimum temperate exhibits a remarkable peak in the Ohio Valley, extending locally into the Appalachians (Daly et al. 2012). A detailed comparative analysis of such patterns in relation to vegetation patterns across all North Temperate regions has yet to be done, building on Grellier's work (1989, 2003).

Evergreen vines are potentially vigorous, aggressive competitors in forests of mid-temperate regions, given suitable soils—and they are often concentrated on moist fertile soils. Some can also survive on the ground in cool-temperate regions (Zone 5 and even 4), although they do not generally climb high, flower and fruit there except near coasts with ameliorated winters. These vines are able to keep photosynthesizing during the winter in mid-temperate zones, where most trees are deciduous on moist fertile soils. However, the evergreen habit does make them susceptible to browsing by mammalian herbivores, especially during the winter when other green leafy forage is often scarce on richer soils.

There has already been considerable interest in the generally high degree of herbivory on vines, and whether aliens are favored more than natives in North America (e.g., Schierenbeck et al 1994, Schweitzer & Larson 1999, Ashton & Lerdau 2007, Knapp et al 2008, Lieurance et al. 2012). Secondary chemistry in plants is generally known to influence mammalian herbivory in varied ways, through deterrence, toxicity or more complex interactions (Harborne 1991a). An attempt is made below to summarize relevant information from the vast and complex literature on chemical patterns and their potential effects. This effort is based on natural history rather

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than theory, although a moderate course for understanding is suggested (following Grubb, 1992) between simple starting points (as offered by Feeney, 1976) and the quagmire of potential complexity (as reviewed by Stamp, 2003).

This paper is in three parts. The first summarizes distributions of evergreen-tending woody vines along habitat gradients in east-central U.S.A. The second part presents an ecological summary of all evergreen vines in mid-temperate regions of the Northern Hemisphere, together with reports of mammalian herbivory, patterns in secondary chemistry and comparisons with deciduous vines. Notes are added on some deciduous woody vines and on some herbaceous or 'subshrubby' vines that are relatively robust or allied with the woody vines. In several taxa, it is not possible to classify them clearly as evergreen, deciduous or herbaceous. The third part provides a general discussion of evolutionary trends and ecological factors, with a focus on potential relationships to mammalian herbivory.

The species of central interest are evergreen-tending vines with the following features.

- (1) All climb onto trees and shrubs, or occasionally cliffs and walls, up to a height of at least 3 m. Only a few species can generally climb up limbless boles of larger trees using adventitious roots or similar means—perhaps just *Decumaria*, *Euonymus fortunei*, *Hedera* and, in a special way, *Bignonia*. Some can also spread much on the ground.
- (2) All are truly evergreen, with leaves usually persisting through winter, or semi-evergreen, with leaves persisting until damaged by cold, rather than by physiologically-timed abscission.
- (3) They all extend significantly into mid-temperate zones (Hardiness Zones 7 or 6).

These characteristics are well-known for native evergreen-tending vines in North America (Duncan 1967) and in Europe (with only *Hedera*). There are diverse evergreen-tending species in East Asia, but published information on them in English is much sparser. It is hoped that this

initial review will lead to more thorough assessment of East Asian species, deeper functional understanding across the temperate world, and improved management of their invasive tendencies. Eventually, the framework used here could be extended to functional comparisons of whole floras. Campbell (1982) presented an initial foray along those lines for trees.

Genera with strictly herbaceous vines in temperate regions are not compared here in detail. In Kentucky, these include Apocynaceae (*Cynanchum*, *Gonolobus*, *Matelea*), Asteraceae (*Mikania*), Convolvulaceae (*Calystegia*, *Convolvulus*, *Cuscuta*, *Ipomaea*), Cucurbitaceae (*Cayaponia*, *Cucurbita*, *Echinocystis*, *Melothria*, *Sicyos*), Dioscoreaceae (*Dioscorea*), Euphorbiaceae (some *Tragia*), Fabaceae (*Amphicarpaea*, *Apios*, *Clitoria*, *Galactia*, *Lathyrus*, *Phaseolus*, *Vicia*), Lygodiaceae (*Lygodium*), Passifloraceae (*Passiflora*), Polygonaceae (*Fallopia*), Rubiaceae (some *Galium*) and Sapindaceae (*Cardiospermum*). Trends in several features that are outlined below (e.g., Tables 7–9) could be extended with inclusion of herbaceous vines, but not trends in dioecy—which is remarkably rare amongst them (only in *Dioscorea* and perhaps some cucurbits). An expanded treatment will eventually embrace these genera as well.

Taxonomic nomenclature generally follows Weakley (2011) and Flora of China (1996–2011). See also Campbell & Medley (2012) for notes on species in Kentucky. Authors of taxa can be checked in those sources, and are not listed here. Suggested common names for genera are listed in Table 7 below, but these are not widely accepted in some cases.



Fig. 1a. *Bignonia capreolata* (purplish) and *Smilax bona-nox* on 14 Dec 2012, 3-5 m high.



Fig. 1b. *Smilax bona-nox*: climbing shoots in Dec (above); juvenile shoots in Jan (below).



Fig. 1c. Unusually pronounced camouflouage in *Smilax bona-nox*; Feb, Grayson Co., KY.



Fig. 1d. *Smilax glauca* on 6 Jan 2013: climbing shoots (above); juvenile shoots (below).

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Fig. 1e. January mix of *Euonymus fortunei* and *Hedera helix* in front-yard of Lexington, Kentucky, a common sight in urban areas of east-central U.S.A.; note striped venation.

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Fig. 1f. *Euonymys fortunei* in February. Purplish color of lower leaf surfaces increases through the winter. F.N. Meyer selected more colored plants as “forma *colorata*” in 1914.

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Fig. 1g. Shoots of alien *Lonicera japonica* (upper) and native *L. sempervirens* (lower), on 18 Jan 2013, Lexington, Kentucky; note flower buds at lower left, and yellowing leaves.

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PART ONE: Distributions of Species along Ecological Gradients in East-central U.S.A.

Typical Positions of Species along Hydrological Gradients

Figure 2 presents a simplified summary of two major ecological gradients among tree species in more natural habitats of east-central U.S.A.—from mesic to xeric conditions, and from mesic to hydric conditions. The topology of this diagram was initially developed to allow two-dimensional displays of all natural vegetation types recognized by NatureServe (2012) across this region. Each species is located at its approximate modal position, as derived from much review of the literature and much general experience in the woods. These gradients are interpreted to reflect the somewhat independent stresses from xeric and hydric conditions, based on a general analysis of compositional data from across east-central U.S.A. (Campbell 1987) and on much local detail in environmental pattern (e.g., Braun 1950, Jones et al. 1984, Campbell & Grubbs 1992, Campbell 2004a, Kupfer et al. 2010, Campbell & Seymour 2011). Sites with ‘xerohydric’ tendency (e.g., ‘post oak glades’) experience relatively large ranges of dry versus wet conditions through the seasons. Unfortunately, there has been little definitive research into the independent nature of xeric and hydric stresses on forests in North America, and a one-dimensional concept of the ‘moisture gradient’ persists in most environmental analysis (e.g., Schaetzl et al. 2009). However, a rigorous approach has been recently initiated in Germany (Schwärzel et al. 2009).

Of course, there are also complicating relationships with past and present disturbance regime. In particular, burning and browsing over several millenia have probably been positively correlated with the gradient from deeper woods at the mesic extreme (often on more rugged terrain) to more open grassy conditions at the ‘xerohydric’ extreme (usually on more gentle

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terrain). And the special effects of forceful flooding are largely confined to the ‘rheic’ (lower left) sector of the diagram. But more short-term or small-scale patterns of disturbance have relatively little projection along these gradients. For example, abundance of *Liriodendron* after disturbance in *Fagus*-dominated woods involves only a small shift away from the mesic extreme. And the extensive artificial edges in modern landscapes are not sampled here.

The most commonly associated tree species for each vine are gleaned from descriptions of vegetation types in NatureServe (2012) and from other sources; see caption to Figure 2. These tree species are abbreviated in the overlays for each vine species. The eight species of evergreen-tending vine in mid-temperate regions have broad overlapping distributions along these gradients of tree composition (Figure 2c-j). There is a sequence from relatively hydric or riparian habitats to relatively xeric or disturbed uplands, as follows: *Decumaria barbara*, *Hedera helix*, *Euonymus fortunei*, *Bignonia capreolata*, *Lonicera* species (abundant *japonica*, local *sempervirens*) and the *Smilax* species (*bona-nox*, *glauca*). The more distinctly evergreen species of *Smilax* that occur in warmer zones are not included here (Weakley 2011): *laurifolia* (esp. swampy sites), *smallii* (esp. drier bottoms) and *auriculata* (esp. submesic to xeric sands).

Except for the wetter habitats of some *Decumaria*, there is a general absence of association between these vines and trees typical of more hydric sites (e.g., *Salix nigra*, *Fraxinus pennsylvanica*, *Taxodium distichum*, *Quercus lyrata*, *Q. phellos*). *Hedera* and *Euonymus* are aliens with only local abundance, but they are centered in habitats transitional from *Decumaria* to *Bignonia*. And the alien *Lonicera* has now filled in much of the potential habitat between *Bignonia* and *Smilax*.

Figure 2a–k. Simplified summary of hydrological gradients in mid-temperate forests of eastern states, with overlays of evergreen vines. Mesic conditions (M) are at center-left; more open, stressed or disturbed conditions increase to upper right (xeric extreme/X), to lower right (hydric extreme/H), or to lower left (riparian/rheophytic with active scouring/R).

- (a) [Next page] General concept of gradients in hydrology and vegetation.
 (b) Six-letter codes indicating approximate modal positions of species, using first three letters of genus and species (see Appendix 1); if more than one mode, the average is shown. Red letters = codes for species associated with strongly acid soils; green = medium acid; blue = circumneutral. Dark grey fill = generally continuous forest; medium grey = more open woods, often mixed with shrubland or grassland; pale grey = zones usually dominated by shrubs or graminoids. Parentheses indicate species with relatively northern ranges, which are replaced by other species to the south within this region. ** Ring-porous wood; * semi-ring-porous.
 (c–j) Distribution of evergreen-tending vines along the gradients, based on general review of common woody associates (including forest types of NatureServe 2012, VegBank of Peet et al. 2012) and personal experience. Shadings indicate concentrations.
 (c) *Decumaria barbara*; see also on Jones+ (1984), Bledsoe+ (2000), Boyle+ (2009) etc.
 (d) *Hedera helix*; based largely on Waggy (2010).
 (e) *Euonymus fortunei*; based largely on Zouhar (2009).
 (f) *Bignonia capreolata*; see also Goebel et al. (2001).
 (g) *Lonicera japonica*; see also Ladwig & Meiners (2010).
 (h) *Lonicera sempervirens*; see also Waters+ (1974), Martin+ (2002), Campbell+ (2011).
 (i) *Smilax bona-nox*; see also Goebel et al. (2001).
 (j) *Smilax glauca*; see also Bazzaz (1968), Clinton et al. (1993), Goebel et al. (2001).
 (k) Sequence of classes for vertical dimension in Figure 3.

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GENERAL CONCEPT OF GRADIENTS IN HYDROLOGY AND VEGETATION Hydric conditions increase to lower right. Xeric conditions increase to upper right. Mesic sites have minimal xeric, hydric & rheic (flood-scoured) influences.		xeric pine/cedar-oak (shrubs abundant)
	subxeric oak woods	varied pine/oak woods or grassland
mesic woods (minimal stress)	submesic woods (some oak)	xerohydric oak woods or grassland
riparian woods (rheic influence)	subhydric woods (some oak)	hydroxeric woods (minor extent)
open rheic woods (shrubs abundant)	open hydric woods (shrubs abundant)	open stagnant water (minor extent)

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(b) Hydrological gradients among trees of east-central states.

		CLIFFS	CLIFFS	CLIFFS	PINVIR JUNVIR ULMSER**	PINRIG QUEPRI** ULMALA**	QUELAE** QUEILI** CELTEN**	XERIC EXTREME
MESIC SLOPES (below)	CLIFFS	QUECOC** QUEVEL** QUESCH**	QUEMON** CARGLA** FRAQUA**	VACARB QUEFAL** (ULMTHO)**	PINECH (POPGRA) FRABIL**	QUEMAR** MALCOR** GLETRI**	QUEMRG** MALANG** (RHUTYP)**	GRASS- LAND
TSUCAR HALTET CLAKEN	MAGMAC QUERUB** TILAME	CASDEN** CAROVA** ULMRUB**	OXYARB QUEALB** QUEMUE**	CARPAL** CARTOM** CARCAR**	PINTAE SASALB* PRUSER*	PINPAL QUESTE** GYMDIO**	QUEINC** (QUEIMB)** (QUEMAC)**	GRASS- LAND
TSUCAN FAGGRA ACESAC	BETLEN LIRTUL AESFLA	MAGACU ACERUB CARCOR*	ILEOPA NYSSYL FRAAME**	(PINSTR) (JUGCIN)* JUGNIG*	QUENIG** ROBPSE** MORRUB*	QUEHEM** DIOVIR* MACPOM**	PINSER QUEPHE** QUESIN**	GRASS- LAND
PLAOCC	MAGTRI BETNIG ACENEG	QUEMIC** ULMAME**	QUESHU** CARLAC**	QUEPAG** CELSPP**	QUELAU** ILEDEC FRAPEN**	(QUEPAL)** CHATHY (QUEBIC)**	PINELL TAXASC QUELYR**	BOG or MARSH or FEN
POPDEL	ACESNM CATSPE**	ACETRI CARILL*	LIQSTY FRASUB**	NYSBIF PLAAQU	MAGVIR FRACAR** FORACU	PERPAL TAXDIS POPHET	CYRRAC NYSQU GLEAQU**	STAGNANT WATER
RHEIC EXTREME	SALSER SALCAR SALINT	ALNSER SALNIG	CORSTR	CEPOCC	DECVER	STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(c) Common associates of *Decumaria barbara* (in more southern region).

								X
				QUEFAL?				
M	MAGMAC QUERUB?		OXYARB QUEALB		PINTAE			
TSUCAN FAGGRA ACEFLO	MAGGRA LIRTUL	ACERUB	ILEOPA NYSSYL FRAAME	PINGLA?	QUENIG	DIOVIR		
		QUEMIC ULMAME	QUESHU?	QUEPAG?	QUELAU	ARUTEC		
		ACETRI	LIQSTY FRAPEN	NYSBIF	MAGVIR FRACAR	NYSOGE TAXDIS		
R		ALNSER						H

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(d) Common associates of *Hedera helix* (English Ivy).

					PINVIR?			X
		QUESCH	FRAQUA					
M	QUERUB	CAROVA	QUEALB QUEMUH		PINTAE			
FAGGRA ACESAC	LIRTUL AESFLA	ACERUB CARCOR	(LIQSTY) FRAAME	JUGNIG	QUENIG			
PLAOCC	BETNIG ACENEG	QUEMIC ULMAME	QESHU CARLAC	QUEPAG CELOCC				
			LIQSTY			TAXDIS?		
R								H

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(e) Common associates of *Euonymus fortunei* (Winter-creeper).

					JUNVIR?			X
		QUESCH	FRAQUA					
M	QUERUB TILAME	CAROVA ULMRUB	QUEALB QUEMUH	CARCAR	PRUSER			
FAGGRA ACESAC	LIRTUL AESFLA	ACERUB CARCOR	FRAAME	JUGNIG				
PLAOCC	ACENEG	ULMAME	QESHU CARLAC	CELOCC	ARUGIG			
	ACESNM							
R								H

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(f) Common associates of *Bignonia capreolata* (Crossvine).

								X
M		QUEVEL QUESCH	CARGLA	QUEFAL				
CLAKEN	QUERUB	CAROVA	OXYARB QUEALB QUEMUH	CARCAR		QUESTE?		
FAGGRA ACESAC	LIRTUL	ACERUB	ILEOPA NYSSYL FRAAME			DIOVIR?		
PLAOCC	BETNIG ACENEG	ULMAME	QESHU					
		ACETRI	LIQSTY					
R								H

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(g) Common associates of *Lonicera japonica* (Japanese Honeysuckle).

					PINVIR JUNVIR	ULMALA	SHRUBBY TRANS- ITIONS	X
		QUEVEL QUESCH	QUEMON CARGLA	QUEFAL	PINECH	GLETRI?	SHRUBBY TRANS- ITIONS	
M	QUERUB	ULMRUB	OXYARB QUEALB QUEMUH	CARTOM	PINTAE SASALB PRUSER	QUESTE	QUEIMB	
FAGGRA	LIRTUL AESFLA	ACERUB CARCOR	NYSSYL	JUGNIG	ROBPSE?	DIOVIR		
		ULMAME	QESHU	CELOCC				
		ACETRI						
R								H

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(h) Common associates of *Lonicera sempervirens* (Trumpet Honeysuckle).

					JUNVIR			X
		QUEVEL QUESCH	CARGLA FRAQUA	(ULMSER)	(POPGRA) FRABIL	GLETRI		
M	QUERUB	CAROVA ULMRUB	QUEALB QUEMUH	CARTOM CARCAR	PINTAE SASALB PRUSER			
ACESAC	LIRTUL	ACERUB	NYSSYL FRAAME	JUGNIG	QUENIG	DIOVIR MACPOM		
		QUEMIC ULMAME	QESHU	CELSPP	QUELAU			
			LIQSTY FRAPEN					
R								H

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(i) Common associates of *Smilax bona-nox* (Saw-brier).

					PINVIR JUNVIR	ULMALA	CELTEN	X
		QUECOC QUEVEL QUESCH	QUEMON CARGLA FRAQUA	VACARB QUEFAL	PINECH FRABIL		RHUGLA	
M	QUERUB	CAROVA ULMRUB	QUEALB QUEMUH	CARTOM CARCAR	PINTAE SASALB PRUSER	QUESTE	SHRUBBY TRANS- ITIONS	
FAGGRA ACESAC	LIRTUL	ACERUB CARCOR	NYSSYL FRAAME	JUGNIG	ROBPSE	DIOVIR		
	ACENEG	QUEMIC ULMAME	QESHU CARLAC	QUEPAG CELLAE	ILEDEC ARUGIG			
			LIQSTY					
R								H

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(j) Common associates of *Smilax glauca* (Cat-brier).

					PINVIR JUNVIR	PINPUN	SHRUBBY TRANS- ITIONS	X
M	RHOMAX	QUECOC QUEVEL	QUEMON CARGLA	QUEFAL	PINECH	KALLAT	RHUCOP	
TSUCAR	QUERUB	CASDEN CAROVA	OXYARB QUEALB	CARTOM	PINTAE SASALB PRUSER	QUESTE	SHRUBBY TRANS- ITIONS	
TSUCAN FAGGRA ACESAC	LIRTUL	ACERUB	NYSSYL FRAAME	PINSTR	QUENIG?	DIOVIR		
	BETNIG			QUEPAG				
		ACETRI	LIQSTY					
R								H

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(k) Typical species for the nine divisions in vertical gradient of Figure 3.

					8	8	9	9
	5	6	6	7	7	8	8	9
4	4,5	5	6	7	7	8	8	9
3,4	3,4	4,5	6	7	8	8	8	9
2,3	2,3	3	3	7	8			
	2	1	1					
	2	1						

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The pH-related Gradient: General Floristic Patterns in Kentucky

The third major gradient among tree species in east-central states is indicated by pH (Table 1, Figure 2). This gradient has been revealed by several analyses (e.g., Campbell 1987, Ulrey 2002, Peet et al. 2003, Simon et al. 2005, Bledsoe & Shear 2000), but it is undoubtedly caused by a complex of trends in soil chemistry and associated environmental factors, rather than just pH. Parallel gradients in Europe have been well-documented by Ellenberg (1988, and previous German editions), leading to many applications of botanical indices for pH and nutrient levels (e.g., Wittig & Neite 1986, Meerts 1997, Thompson et al. 1997, Ertsen et al. 1998, Wamelink et al. 2002, Chytrý et al. 2003, Ewald 2003, Thimonier et al. 2006, Verheyen et al. 2012). However, physiological mechanisms remain poorly understood for most non-crop plants, even in Europe (Fisher & Binkley 2000, Mengel & Kirkby 2001, Brady & Weil 2002, Sparks 2003). Soils with pH about 5.5–6.5 tend to have the highest natural levels of available nitrogen and phosphorous, and pH itself need have little or no simple direct relationship to the gradient. But the prevalent form of nitrogen changes from ammonium at lower pH to nitrate at higher pH (e.g., Aciego-Petri & Brooke 2008). Also, aluminum and manganese can become more soluble and toxic to some plants in strongly acid soils (e.g., Vanguelova et al. 2007).

Table 1 summaries trends in some ecomorphological features of the woody flora in Kentucky as a whole. Each of the 279 native species has been assigned to its most typical (modal) position along the pH-related gradient. (Although perhaps absent from Kentucky in 1492, *Maclura pomifera* is included here since it has become widely naturalized from the southwest, and it has instructive ecology.) The following five classes are used to represent the pH-gradient: A to E (Campbell & Medley 2012).

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- A. Association with strongly acid soils (ca. pH 4–5) and low overall fertility, usually with a sandy component or on Devonian black shale. *Tsuga canadensis* is potentially dominant on mesic sites; common upland oaks include *Quercus montana* and *Q. coccinea*; *Castanea dentata* was formerly frequent; Ericaceae are common. Typical species are concentrated in the Knobs and Appalachian regions, coupled with virtual absence in calcareous regions.
- B. Transitional or uncertain assignment between A and B; usually on soils derived from sandstone or acid shales.
- C. Association with medium acid soils (ca. pH 5–6) and medium overall fertility; soils often have mixed origin with some sand. *Fagus grandifolia* is potentially dominant on mesic sites; common upland oaks include *Q. alba*, *Q. velutina*, *Q. stellata* and *Q. falcata*; Ericaceae are uncommon to absent. Typical species are scattered over most of the state, including hills of the Bluegrass, other calcareous regions, Knobs and Appalachian hills.
- D. Transitional or uncertain assignment between C and E; soils often have mixed origin, sometimes with much sand. Species that are common on farmland or alluvial soils with relatively high fertility (especially in N and P) are generally assigned here.
- E. Association with weakly acid to neutral soils (ca. pH 6–7) and medium to high overall fertility, especially in bases (Ca, Mg, K); soils generally lack much sand. *Acer saccharum* (sensu lato) is potentially dominant on mesic sites; common upland oaks include *Q. muhlenbergii*, *Q. shumardii*, *Q. macrocarpa* and locally *Q. imbricaria*. Typical species have higher frequency in calcareous regions, especially the Bluegrass, compared to the Knobs and Appalachian hills (where they are absent or restricted to unusually fertile valleys and local base-rich shales).

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The majority of native woody species (60%) are assigned to classes C or D, but the subset of woody vines have a much stronger concentration in these two classes (88%). The two vines with most evergreen tendency (*Bignonia capreolata* and *Smilax bona-nox*) are both typical of class D, together with the evergreen mistletoe (*Phoradendron leucarpum*). *S. glauca* is centered in class B, but it has less persistent leaves, often falling during colder winters. In contrast, the evergreen habit among trees and shrubs is strongly concentrated among species of class A, on more acid and infertile soils. Among large tree species, those with entire simple leaves are concentrated in class B, while those with compound leaves are concentrated in classes D and E. And, among trees, other features have increased occurrence from A to D or E: dioecious flowering (large trees only); potential for clonal spread with lateral roots; unusually large edible fruits; and thorns (large trees only).

Table 2 shows patterns in some functional groups among the herbaceous flora of Kentucky. Again, distinctly evergreen species—ferns, subshrubs and herbs (excluding graminoids)—are concentrated on strongly acid soils (class A). Mycotrophic and parasitic species also tend to be associated with more acid soils. Herbaceous vines, including annuals, are most frequent among species typical of moderate position along the gradient (class C). Other annuals, biennials or monocarpic perennials tend to be concentrated at higher pH, especially winter-annuals.

The pH-related Gradient: Vine Frequencies in Plots from Central Kentucky & VegBank

Figure 3 is designed to combine the pH-related gradient with the general gradient from mesic or riparian woods to more open xeric or disturbed woods on uplands (as expressed in Figure 2k). Distinctly hydric vegetation is generally excluded, although riparian woods and

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subhydric transitions to thin swampy woodland are included in the lowest rows. Figure 3b indicates approximate modal positions for each common tree species, and the approximate correspondence of vegetation classes depicted here to the associations of NatureServe (2012).

Figure 3d–j presents overlaid data from 746 plots surveyed in central Kentucky. These are plots of 15 m radius (706.5 m²) for trees over 9.5 cm dbh, with subplots of 10 m radius (314 m²) for smaller stems and ground vegetation. They include 628 plots surveyed on Daniel Boone National Forest during 1993–1995 (with M. Hines & D. Taylor); plus 118 surveyed in the central Bluegrass region during 1977–79 (Campbell 1980). Plots were located regularly along subjectively selected, diverse topographic transects in order to represent the full range of species composition as equitably as possible. They do not systematically cover the whole landscape. They were selected to include some canopy trees of at least 25 cm dbh, and they exclude obvious edges or transitions to recent old fields. But in most cases there has been some cutting of trees, and in many cases the woodland is successional, especially from old pastured areas. These plot data are supplemented with other general information on occurrence of each vine species based on selected published sources and VegBank (Appendix 2). Data from VegBank (Peet et al. 2012) were checked on 6 Dec 2012, and refer only to the 1485 plots from southeastern states (AL, MS, NC, SC, TN, VA, WV). These plots mostly range from 100 to 1500 m² in area, with miscellaneous sampling methods.

Evergreen trees—plus minor amounts of evergreen shrubs—are most abundant, as a proportion of basal area, in vegetation typical of strongly acid soils, and at the xeric or disturbed extreme (Figure 3c). In contrast, six of the seven evergreen-tending vines are concentrated in vegetation typical of medium acid to base-rich soils, and most of them are concentrated in mesic to subxeric woods (Figure 3d–j). Each vine has relatively little

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occurrence in vegetation typical of mesic to subhydric alluvial sites on base-rich soils. However, there is a curious tendency for extension into subhydric vegetation on medium acid soils, especially by *Lonicera japonica*. Could rarity on damp base-rich soils be caused by the history of herbivory?

Other Distributional Data for Species in East-central U.S.A.

Geographic distributions of these vines across the U.S.A. are fairly well-documented by BONAP (Kartesz 2012) and others (e.g., Tables 3 and 4), but there are inconsistencies and gaps, especially in ranges of the less widespread aliens, *Hedera helix* and *Euonymus fortunei* (Figure 4). Although many thousands of plots have been surveyed for flora and vegetation in eastern states during recent decades, there has, unfortunately, been little synthesis of these data until the recent efforts of VegBank (Peet et al. 2012). A much larger database, begun in 1930, is the continuing Forest Inventory Analysis (FIA) of US Forest Service, providing a systematic-random sample of forested area that is “at least 37 m wide; 0.4-ha in size; covered, or formerly covered, by trees; capable of tree-growth; and not developed for nonforest uses” (Rudis et al. 2005). The FIA has been largely limited to better known woody species, but more details of other plants are now being collected (e.g., Miller et al. 2008; see Table 3).

Lonicera japonica. The map of this species produced by recent FIA for southeastern states (Figure 5) shows its frequency by county in the subplots of 168 m² (groups of four subplots are dispersed within ca. 6000 m²). This alien vine is most abundant, with frequencies of 50-90%, in the Piedmont (from Virginia to Georgia), in the Upper East Gulf Coastal Plain, and in other low hills where much forest has been fragmented and farmland has been abandoned. It is infrequent

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(< 10%) to absent in some counties of the Mississippi Alluvial Plain, Nashville Basin, Kentucky Bluegrass, and Ridge-and-Valley regions.

With these plot data from southeastern states, Wang et al. (2012) showed a positive relationship of *L. japonica* to site productivity (site index), and negative to recent occurrence of fire (within past 5 years). Across northeastern states, Schulz & Gray (2012) found relatively high frequency in whole FIA plots (subplots combined) that were classified as mixed upland forest (17.1%); less in white oak (13.9%), oak-hickory (13.6%), and hard maple (11.4%) types. They found little difference comparing plots of intact forest to plots at edges (9.0% versus 11.7% overall), but the difference was much greater for the subset of plots classified as white oak (9.1% to 21.4%). Possible relationships of these data to patterns in deer populations do not appear to have been studied. Some recent statistical models have focussed on the likely effects of forest fragmentation and global warming, and on the extent to which *L. japonica* currently saturates its potential habitat or has room for further expansion (Lemke et al. 2012, Wang et al. 2012). The species is clearly concentrated at edges of forests, especially roadsides, where is often the most abundant alien plant. It appears to have spread relatively fast into most suitable habitat across southeastern states within the past century or so (Merriam 2003).

Smilax. A large amount of plot data exists for these species in the literature, but there has been virtually no general analysis in relation to habitat gradients. Data in a recent thesis by Ulrey (2002) does allow some confirmation of the trends reported here for *S. glauca* (Table 4)—much more synthesis of such information is warranted for this important genus.

Table 1. Some pH-related trends in ecomorphological features of Kentucky's native woody plants, as indicated by typical positions for each species along the pH-related gradient. See text for explanation of gradient; see Appendix 1 for species abbreviations; see Campbell & Medley (2012) for listings and distributions of large trees, small trees, large shrubs, small shrubs and vines. Varieties are counted in total taxa. Features are as follows.

Percentage of all trees and shrubs with evergreen leaves. All trees are listed.

Percentage of trees with entire simple leaves. These do not include needle-leaved gymnosperms. Trends are weak to absent among shrubs and vines.

Percentage of total that are vines (or parasitic epiphytes). All species with evergreen leaves are noted plus the parasitic mistletoe, *Phoradendron leucarpum*; those with semi-evergreen to deciduous species are in parentheses.

Percentage of large trees with compound leaves. These do not include needle-leaved gymnosperms. Trends are absent among smaller trees, shrubs and vines.

Percentage of all trees with clonal spread. These have frequent lateral spread by root-suckering or layering. Large trees are listed. Clear trends are absent among shrubs.

Percentage of large trees that are dioecious. Included here are 'polygamo-dioecious' species: with sexes concentrated on separate trees but with some bisexual flowers in some cases. Clear trends are absent among smaller trees, shrubs and vines.

Trees with unusually large fruit. These are large or small trees with the largest fruits (in Kentucky) within their genus if more than one species, or the largest within their family if only one species per genus, or the largest within their order if one species per family. Excluded are species with normal types of bird-dispersed or wind-dispersed seeds.

Large trees with thorns. Trends are absent among smaller trees and shrubs. *Ilex opaca*, with prickles on leaves, is classified here as a small tree. *Juniperus* might also be added.

Ecomorphological Features	Percentages of species (and noted species) at their typical positions along the pH-related gradient				
	A: strongly acid	B: intermediate	C: medium acid	D: intermediate	E: base-rich
% of all trees and shrubs with evergreen leaves; trees listed below	29% Pinrig Tsucan (Rhomax)	9.5% Pinvir Pinech Pinstr (Ileopa)	0	6% Junvir Thuocc	0
% of all trees with entire simple leaves	30%	21%	14%	10%	11%
% of total that are vines; evergreen species listed below	0	3% (Smigla)	17% (Lonsem)	21% Bigcap Pholeu (Smibon)	4%
% of large trees that are strictly dioecious (*) or partially so	0	0 [smaller tree: Ileopa*]	23% Acerub Divir* Nysaqu* Nyssyl* Popgra* Sasalb*	38% Acescn Diopub* Fra(5) Gleaqu Junvir* Morrub Pop(2)* Salnig*	26% Aceneg* Gletri Gymdio* Frapro Macpom*
% of all trees with compound leaves	0	14%	7%	25%	25%
% of all trees with clonal spread (lateral roots or layers); large trees listed	10%	14%	12% Diovir Sasalb	19% Robpse Salnig (Thuocc) Ulmrub	30% Gymdio Gletri ?Ulmitho Macpom
Trees with unusually large edible fruits (versus related taxa)		Casden	Diovir Malcor	Carlac Aesfla Asitri	Quemac Jugnig Gymdio Prumio Macpom Cramol
Large trees with thorns				Robpse (Gleaqu)	Gletri Macpom
Total large trees	7	9	25	35	19
Total small trees	3	5	18	17	9
Total large shrubs	9	19	16	6	9
Total small shrubs	5	6	10	11	7
Total woody vines	0	2	14	18	2
Combined Totals	24	39	83	87	46

Table 2. pH-related trends in some functional groups among Kentucky's native herbaceous vascular plants, as indicated by the distribution of typical positions for each species along the pH-related gradient. See text for explanation of the pH-related gradient; see database of Campbell & Medley (2012) for listings and distributions of functional groups. Groups without clear trends are not shown, but they are included in the totals (determinate spring woodland perennials, rosette perennials, shorter spring-summer perennials, summer woodland perennials with simple broad leaves, shorter graminoids of more open land, and aquatic plants). Numbers of species within each group are provided in parentheses. Percentages are based on the total numbers of all woody plus herbaceous native species at each position along the gradient (A to E); these totals are provided in the bottom row. Shadings indicate concentrations. Details of functional groups are as follows.

Evergreen ferns, herbs and subshrubs. These are species with tough evergreen leaves usually evident above the litter, excluding the many species with only basal tufts of leaves that are protected near ground level. Nevertheless, some assignments are tentative, and species of rock outcrops deserve may deserve distinction; sandstone species are mostly assigned to A, limestone species to E. Graminoids are excluded. The exceptional plants under D and E are mostly ferns, also *Hepatica acutiloba* and *Paxistima canbyi*.

Mycotrophic plants. Partially or (in a few cases) completely dependent on fungi for nutrition; most species are in Orchidaceae. Further refinement of concepts is needed.

Parasitic plants. Partially or completely dependent on host plants for nutrition; most species are in Orobanchaceae.

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Larger summer-fall graminoids. These are all C4 species in Cyperaceae and Poaceae that are typical of open sunny sites. Excluded are those with short stature, ca. 0.1–0.2(–0.4) m tall, which are mostly typical of wetlands, shorelines and trampled areas; also excluded are “woodland” species such as *Muhlenbergia* spp.

Later woodland graminoids. These are C3 panic-grasses (*Dichanthelium* spp.) plus a few C4 muhly-grasses (*Muhlenbergia* spp.).

Herbaceous Vines. These include annual vines. A similar trend (with peak in C–D) occurs among woody vines (Table 1).

Earlier woodland graminoids. These all have C3 photosynthesis.

Distichous woodland lilioid perennials. This small group is affiliated with some woodland graminoids or herbs.

Tall summer-fall perennials. Plants reach ca. 1–3 m. Trends are similar for running and non-running subgroups.

Tall annuals. These plants reach ca. 1–3 m and are self-supporting; vines are excluded. Note also tall annual crops on fertile soils: okra, hemp, tobacco, corn (maize).

Summer woodland perennials with large divided leaves. Trends are similar for early and late flowering subgroups.

Biennials and monocarpic perennials. Trends are similar for early and late flowering subgroups.

Winter-annuals. Trends are similar for subgroups of woodlands, old fields and rocky glades.

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Functional Groups of Native Herbaceous Species (total numbers in parentheses)	Percentages of species in divisions of the pH-related gradient				
	A: strongly acid	B: intermediate	C: medium acid	D: intermediate	E: base-rich
Evergreen ferns, herbs and subshrubs (58)	16%	2.4%	1.4%	1.0%	3.5%
Mycotrophic plants (51)	8.9%	3.4%	3.4%	0.7%	0.5%
Parasitic plants (27)	2.4%	2.0%	2.0%	1.2%	2.0%
Larger summer- to fall-flowering graminoids of grassland (49)	1.6%	5.1%	2.4%	1.8%	1.5%
Later woodland graminoids (28)	1.6%	2.4%	1.6%	1.0%	0.5%
Herbaceous vines (43)	0.8%	0.3%	3.3%	2.1%	2.0%
Earlier woodland graminoids (28)	0	0	0.8%	2.7%	2.0%
Distichous woodland lilioids (21)	0	1.0%	1.0%	1.3%	1.0%
Tall summer-fall perennials (66)	0	1.0%	3.0%	5.2%	3.5%
Tall annuals (15)	0	0	0.3%	1.6%	1.0%
Summer woodland perennials with large divided leaves (37)	0	1.0%	1.7%	1.9%	4.5%
Biennials and monocarpic perennials (51)	0.8%	0.7%	1.0%	4.1%	6.4%
Winter-annuals (34)	0%	0%	1.1%	2.4%	5.9%
Total Species (herb. + woody)	123	293	693	660	200

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Figure 3a–j. The pH-related gradient and dryness/disturbance gradient among trees of east-central U.S.A., with trends in evergreen-tending woody plants as overlays.

Figure 3a,b. Diagrams combining the pH-related and dryness/disturbance gradients. See text for explanation of the pH-related gradient (left to right). The vertical gradient with nine divisions, as outlined below, expresses the broadly defined trend from lowlands to uplands, with increasing dryness and associated disturbance (Figure 2k). Purely hydric vegetation is excluded. Six-letter codes for species (as in Figure 2b) show approximate positions of their maximum frequency (with alternatives in parentheses for a few bimodal or broadly ranging species); some smaller species are excluded due to lack of space, especially Rosaceae. Four-number codes indicate approximate positions of most native vegetation types (CEGLs) that have been described by NatureServe (2012) for non-hydric sites in Kentucky plus adjacent states.

9. Grassy openings due to xeric conditions, burning or browsing.
8. Xeric to subxeric woods plus open shrubby or grassy savannas.
7. Subxeric/seral woods, plus thin or shrubby transitions/savannas.
6. Transitions from subxeric/seral to more mesic woods.
5. Transition from mesic to subxeric/seral woods, often ‘submesic’.
4. Mesic colluvial woods, especially toeslopes and NE-faces.
3. Mesic alluvial woods, often transitional to riparian woods.
2. Riparian woods, with forceful but temporary flooding.
1. Subhydric woods, but often transitional to submesic woods.

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Figure 3c. Percentage of evergreen species in basal area of forest, based on plots from central Kentucky. See text for sources of data; plots are 15 m radius for stems >9.5 cm dbh, with subplots of 10 m radius for stems 1–9 cm dbh and ground vegetation. The evergreen species are *Juniperus virginiana*, *Ilex opaca*, *Pinus* spp. (*echinata*, *rigida*, *strobus*, *virginiana*) and *Tsuga canadensis*, plus local *Kalmia latifolia* and *Rhododendron maximum* in the shrub layer.

Figures 3d–j. Distribution of evergreen-tending vines. Shadings indicate concentrations based on the following data.

In parentheses: (numbers/total plots) indicate occurrences of the species in plots of 10 m radius (0.314 ha); see text for sources of data.

To left: + indicates added observations of the author from central Kentucky;
 ++ indicates observed concentrations observed by the author.

To right: + indicates typical presence in CEGF descriptions of NatureServe (2012); or concentrations reported by Ulrey (2002), Biggerstaff & Beck (2007), Waggy (2010), Zouhar (2009), Campbell & Seymour (2011), etc.

indicates plot numbers with the species in VegBank, based on searches in Dec 2012; see Appendix 2 for lists of individual plots.

(a) Summary diagram, with common names of predominant trees.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
xeric or seral (to subxeric)	PINES, HEATHS and transitions with sassafras, persimmon		Varied mixes esp. post oak blackjack o.	RED CEDAR, LOCUSTS CHERRIES, PLUMS diverse hawthorns, briars	
subxeric or seral (to submesic)	OAK-CHESTNUT (former)	OAK-HICKORY: esp. white oak, black oak, s. red oak; pignut, mockernut, shagbark; local red maple, blackgum			OAK-ASH+ chink. oak bur oak+
mesic (to submesic or subxeric)	HEMLOCK BIRCHES +	BEECH, SUGAR MAPLE TULIP, BUCKEYES, BASSWOODS and drier transitions with n. red oak			BLACK MAPLE bitternut+
riparian (to mesic)	absent or rare	RIVER BIRCH, SYCAMORE shrubby willows		BOX ELDER, SILVER MAPLE, SYCAMORE local willow, cottonwood	
subhydric	absent or rare	SWEETGUM, SWAMP RED MAPLE, ALDER		GREEN ASH, WHITE ELM taller willows	

(b) Details of pH-related and dry/open gradients among trees of east-central U.S.A.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	Danser 4061 7805	Andter Danspi 2417 4756 7707	Schsco Andger Andvir Pananc 4044 4686 7705	Setgen Rhuspp Rubspp 2024 4624 4738	Spocom Trifla 4078
8 xeric-sx.+	Pinrig Oxyarb 3617	Pinvir Quemar 3765 6327 7500	Queste Diovir Chivir Popgra 4217 5018 7121	Junvir Queimb Robpse Arugig 3836 4732 7279	Quemac Gletri Macpom Fracar 3835 4436 4544
7 subxeric+	Quecoc Steova 4412 6271 8431	Pinech Carpal 2591 7247 7493	Quefal Cartom Sasalb Corflo 7244 7746 7795	Pruser Cercan Vibruf Ulmala 3836 4133 7699	Jugnig Celocc Ulmtho/ser 4693 7180 7879
6 sx.-mesic	Quemon Magmac 4425 7267	Pintae Casden 7196 7268 8521	Quealb Cargla Quevel Nyssyl 2067 7219 8428	Carova (Ulmame) Morrub Asitri 2070 6445 7240	Quemuh Queshu Carlac Claken 3876 4697 4741
5 submesic+	Betlen Rhomax 6923 7102 7565	Pinstr lleopa 6192 7286 7300	Querub Lirtul Acerub (Liqsty) 7220 7218 7881	(Carcor) Ulmrub Fraame Ostvir 4793 7698 7233	Aesgla Gyndio Fraqua 4437 6237 8442
4 mesic coll.	Tsucan Betall 7136	Magtri 5043 8407	Faggra Magacu 6055 7200 7201	Acesac Aesfla Tilhet Carcar 2411 5222 7695	Acenig Tilame (Car cor) 4411 6471 8412
3 mesic all.		transitions/mixes 7143 7565	transitions/mixes (Quemic) 2099 7281 7340	transitions/mixes (Ulmame) 7184 7334 8429	transitions/mixes Cellae (Queshu) 4697 7339
2 riparian		Salser 3895 7314	Betnig Salcar 7312 8471	Plaocc Acescn 2431 2586 4626	Aceneg Popdel 2018 4690 5033
1 subhydic		Alnser 3737 3894 7443	Acetri Quepal (Liqsty Quepag) 2438	Frapen Salnig (Quemic) 6548 7703	Salint 8562

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(c) Percentage of evergreen trees in basal area of Kentucky plots

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	66	no plots	no plots	no plots	63
8 xeric-sx.+	51	56	61	no plots	29
7 subxeric+	7.9	9.4	8.5	12	27
6 sx.-mesic	34	17	14	3.3	4.4
5 submesic+	50	37	4.2	0.4	2.3
4 mesic coll.	65	36	8.6	0	0
3 mesic all.	no plots	41	3.1	0	0
2 riparian	no plots	(1 plot)	0.2	0.1	0
1 subhydic	no plots	33	3.3	0.5	0

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(d) *Hedera helix*: in only 2 of 746 Kentucky plots and 6 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)				(0/2)
8 xeric-sx.+	(0/22)	(0/25)	(0/16)		(0/9)
7 subxeric+	(0/43)	(0/72)	(0/27)	(0/5) +	(0/10)
6 sx.-mesic	(0/12)	(0/47)	(0/56) +	(0/45) +	+ (0/41)
5 submesic+	(0/7)	(0/19)	(0/24) + #3,4,6	+ (0/25) +	+ (1/26)
4 mesic coll.	(0/11)	(0/14)	(0/22) #1,5	+ (0/26)	+ (1/8)
3 mesic all.		(0/8)	(0/13) +	+ (0/3) #2	+ (0/2)
2 riparian		(0/1)	(0/9)	(0/9)	(0/4)
1 subhydic		(0/7)	(0/48)	(0/4)	(0/5)

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(e) *Euonymus fortunei*: in only 8 of 746 Kentucky plots and 1 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)				(0/2)
8 xeric-sx.+	(0/22)	(0/25)	(0/16)		+ (1/9)
7 subxeric+	(0/43)	(0/72)	(0/27) +	+ (1/5)	+ (1/10) +
6 sx.-mesic	(0/12)	(0/47)	(0/56) +	+ (1/45)	++ (2/41) +
5 submesic+	(0/7)	(0/19)	(0/24) +	+ (0/25) +	++ (0/26)
4 mesic coll.	(0/11)	(0/14)	(0/22) #1	+ (1/26) +	++ (0/8)
3 mesic all.		(0/8)	(0/13)	+ (1/3) +	+ (0/2)
2 riparian		(0/1)	(0/9)	+ (0/9) +	+ (0/4)
1 subhydic		(0/7)	(0/48)	(0/4)	(0/5)

50

(f) *Bignonia capreolata*: in 126 of 746 Kentucky plots but only 5 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)				(0/2)
8 xeric-sx.+	(0/22)	(1/25)	(0/16)		(2/9)
7 subxeric+	(0/43)	(1/72)	(2/27)	(2/5) +	(1/10)
6 sx.-mesic	(2/12)	(2/47)	(11/56)	(19/45)	(15/41)
5 submesic+	(0/7)	(2/19) +	(5/24) #4	(5/25)	(11/26) +
4 mesic coll.	(0/11)	(1/14)	(10/22) +	(6/26)	(4/8)
3 mesic all.		(0/8)	(5/13)	(1/3) #?	(0/2)
2 riparian		(0/1)	(5/9)	(1/9)	(0/4) #3
1 subhydic		(0/7)	(11/48)	(0/4) #1,2,5,?	(1/5)

51

(g) *Lonicera japonica*: in 66 of 746 Kentucky plots and 17 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)				(0/2)
8 xeric-sx.+	(0/22)	(0/25)	(0/16)	(0/0) + #13	(2/9)
7 subxeric+	(0/43)	(1/72) + #1	(1/27) + #17	(0/5) + #5,15,16	(2/10) +
6 sx.-mesic	(0/12)	(0/47)	(5/56)	(2/45) #9?	(7/41)
5 submesic+	(0/7)	(0/19) +	(2/24) +	(3/25) + #2,7,8,10,11	(3/26) + #4
4 mesic coll.	(0/11)	(1/14)	(2/22) +	(1/26)	(0/8)
3 mesic all.		(0/8)	(2/13) #6	(0/3) #3	(0/2)
2 riparian		(0/1)	(1/9)	(0/9) #12	(0/4)
1 subhydic		(0/7)	(7/48)	(4/4) #14	(0/5)

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(h) *Lonicera sempervirens*: in zero Kentucky plots and zero VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)				(0/2)
8 xeric-sx.+	(0/22)	(0/25)	(0/16) +	(0/0) +	(0/9) +
7 subxeric+	(0/43)	(0/72)	(0/27) +	(0/5) +	(0/10) +
6 sx.-mesic	(0/12)	(0/47) +	(0/56) +	(0/45) +	(0/41) +
5 submesic+	(0/7)	(0/19)	(0/24) +	(0/25) +	(1/26) +
4 mesic coll.	(0/11)	(0/14)	(0/22) +	(0/25) +	(1/8) +
3 mesic all.		(0/8)	(0/13)	(0/3) +	(0/2) +
2 riparian		(0/1)	(0/9)	(0/9)	(0/4)
1 subhydic		(0/7)	(0/48)	(0/4)	(0/5)

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(i) *Smilax bona-nox*: in 17 of 746 Kentucky plots and 9 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(0/19)			(0/0) #6	(1/2)
8 xeric-sx.+	(0/22)	(0/25)	(0/16)	+ (0/0)	+ (1/9)
7 subxeric+	(0/43)	(0/72) #2,4	(0/27) + #1	+ (0/5)	+ (1/10)
6 sx.-mesic	(0/12)	(0/47)	(0/56)	(4/45)	(5/41)
5 submesic+	(0/7)	(0/19)	(0/24)	(1/25)	+ (2/26)
4 mesic coll.	(0/11)	(0/14)	(0/22)	(0/25)	(2/8)
3 mesic all.		(0/8)	(0/13) #5	(0/3) #7,8	(0/2)
2 riparian		(0/1)	(0/9)	(0/9)	(0/4)
1 subhydic		(0/7)	(0/48)	(0/4) #9	(0/5)

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(j) *Smilax glauca*: in 254 of 746 Kentucky plots and 7 of 1485 VegBank plots.

GRADIENT IN DRYNESS AND DISTURBANCE	pH-RELATED GRADIENT (A: strongly acid to E: circumneutral)				
	A	B	C	D	E
9 openings	(6/19)				(0/2)
8 xeric-sx.+	(17/22) + #3,9	(18/25) + #4,8	(10/16) +		(0/9)
7 subxeric+	(28/43) ++ #5,6	(47/72) ++ #7	(20/27) +	(0/5)	(0/10)
6 sx.-mesic	(5/12) +	(24/47) +	(25/56) +	(12/45)	(0/41)
5 submesic+	(2/7) +	(6/19) +	(9/24) +	(7/25)	(1/26)
4 mesic coll.	(1/11)	(3/14) +	(2/22) +	(0/25)	(0/8)
3 mesic all.		(2/8)	(2/13)	(0/3)	(0/2)
2 riparian		(0/1)	(2/9)	(0/9)	(0/4)
1 subhydric		(3/7)	(14/48)	(1/4)	(0/5)

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Table 3. Estimated “cover” percentages of alien evergreen vines within the “forested” area of southeastern states, based on Forest Inventory and Analysis by US Forest Service. Data come from Miller et al. (2008); see text. Euofor = *Euonymus fortunei*; Hedhel = *Hedera helix*; Lonjap = *Lonicera japonica*. In Texas, only eastern forested regions are included.

SPECIES	ESTIMATED PERCENT COVER IN EACH STATE											
	AL	AR	FL	GA	KY	LA	MS	NC	SC	TN	TX	VA
Euofor	0	0.0004	0	0	0.06	0	0	0.0009	0	0.03	0	0.00004
Hedhel	0.007	0.00004	0	0.00008	0	0	0.02	0.008	0.02	0.007	0	0.02
Lonjap	12.9	2.1	0.02	3.0	4.8	0.7	9.0	3.0	6.3	9.7	2.1	5.8

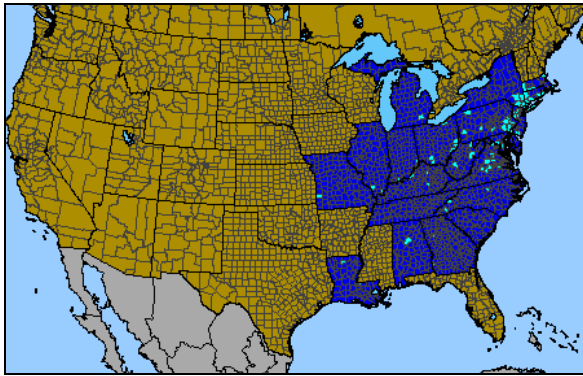
Table 4. Distribution of *Smilax glauca* among forest types < 4500 ft elevation in the southern Appalachian region, based on information assembled by Ulrey (2002: Table 4). Each cell indicates the forest type name and the percentage of occurrence in 0.1 ha plots.

HABITAT DESCRIPTORS	STRONGLY ACID approx. = A of Fig. 3	INTERMEDIATE approx. = B of Fig. 3	MEDIUM ACID approx. = C of Fig. 3
7. XERIC-SX.	Table Mt. Pine 78	Shortleaf Pine 74	(no plots)
6. SUBXERIC	Chestnut Oak 78	Oak-hickory 94	Calcareous oak +
5. MESIC-SX.	Carolina Hemlock 86	White Pine 65	Red oak 52
4. MESIC	Canada Hemlock 24	Acid cove forest 50	Rich cove forest 75

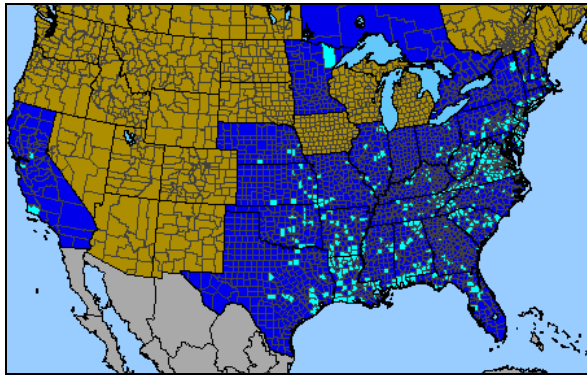
56

Figure 4. Maps of documented counties with evergreen or semi-evergreen vines of mid-temperate regions in east-central states, from BONAP (Kartesz 2012).

Pale green on dark green indicates native species; pale blue/pink on dark blue indicates alien species, with pink indicating official noxious status. The current Hardiness Zones occupied by these species are indicated after each name (USDA 2012). *Clematis terniflora* has little or no woody growth above ground in Zones 6–7; see notes in text under Ranunculid Orders.

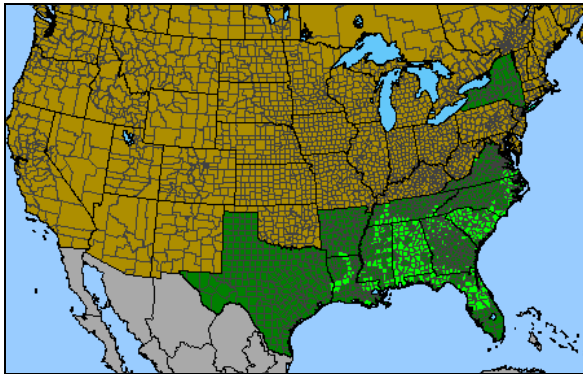


Akebia quinata: Zones 6a to 7b (?)

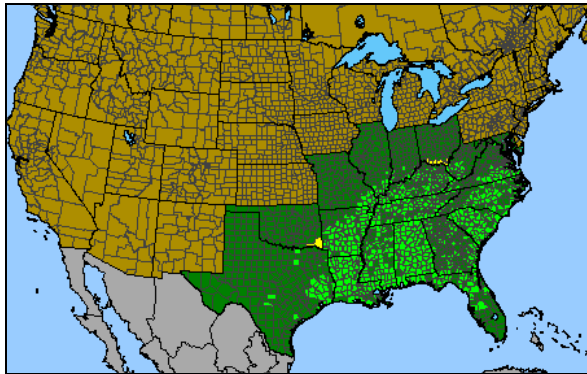


Clematis terniflora: Zones 6a to 9a

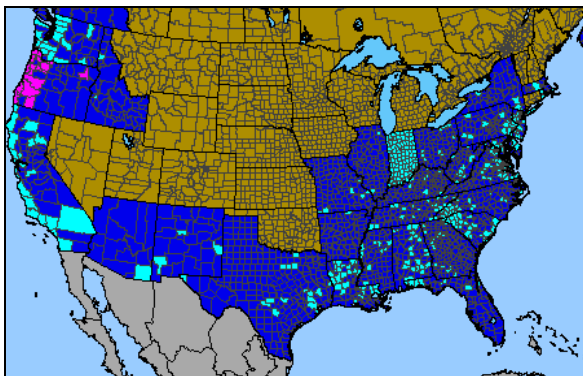
57



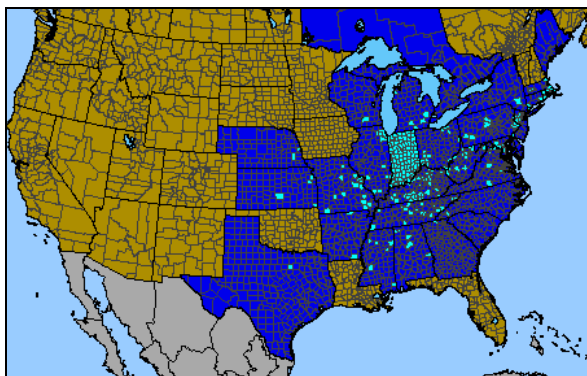
Decumaria barbara: Zones 7b to 9b



Bignonia capreolata: Zones 6b to 9b

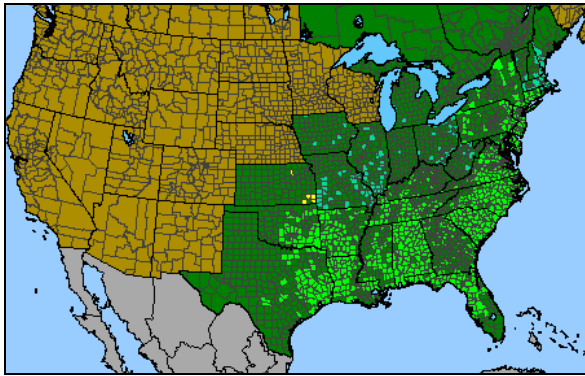


Hedera helix: Zones 6a to 9a (?)

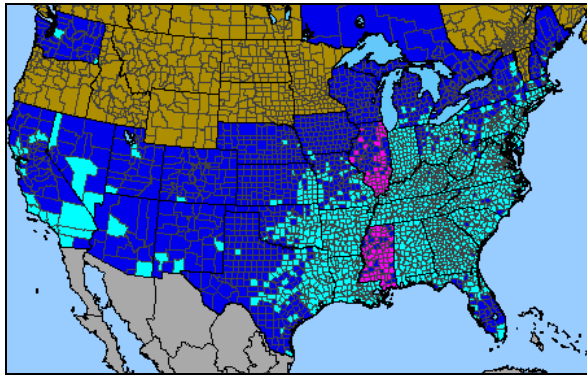


Euonymus fortunei: Zones 6a to 8a (?)

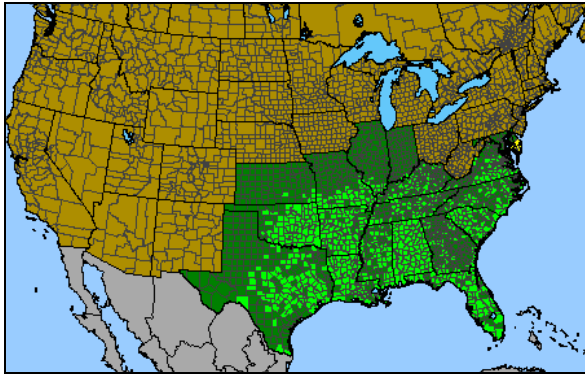
58



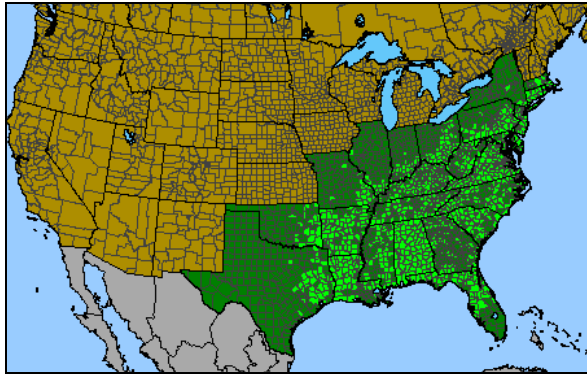
Lonicera sempervirens: Zones 5b to 9b



Lonicera japonica: Zones 5b to 9b



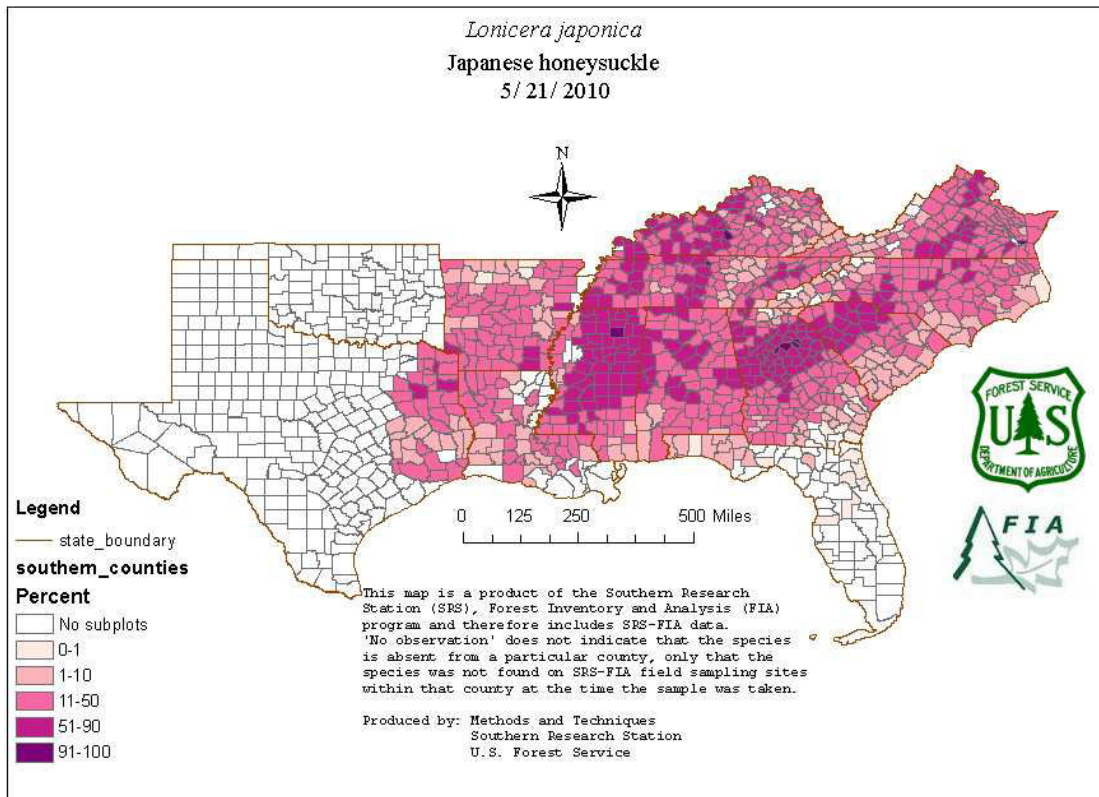
Smilax bona-nox: Zones 6b to 10a



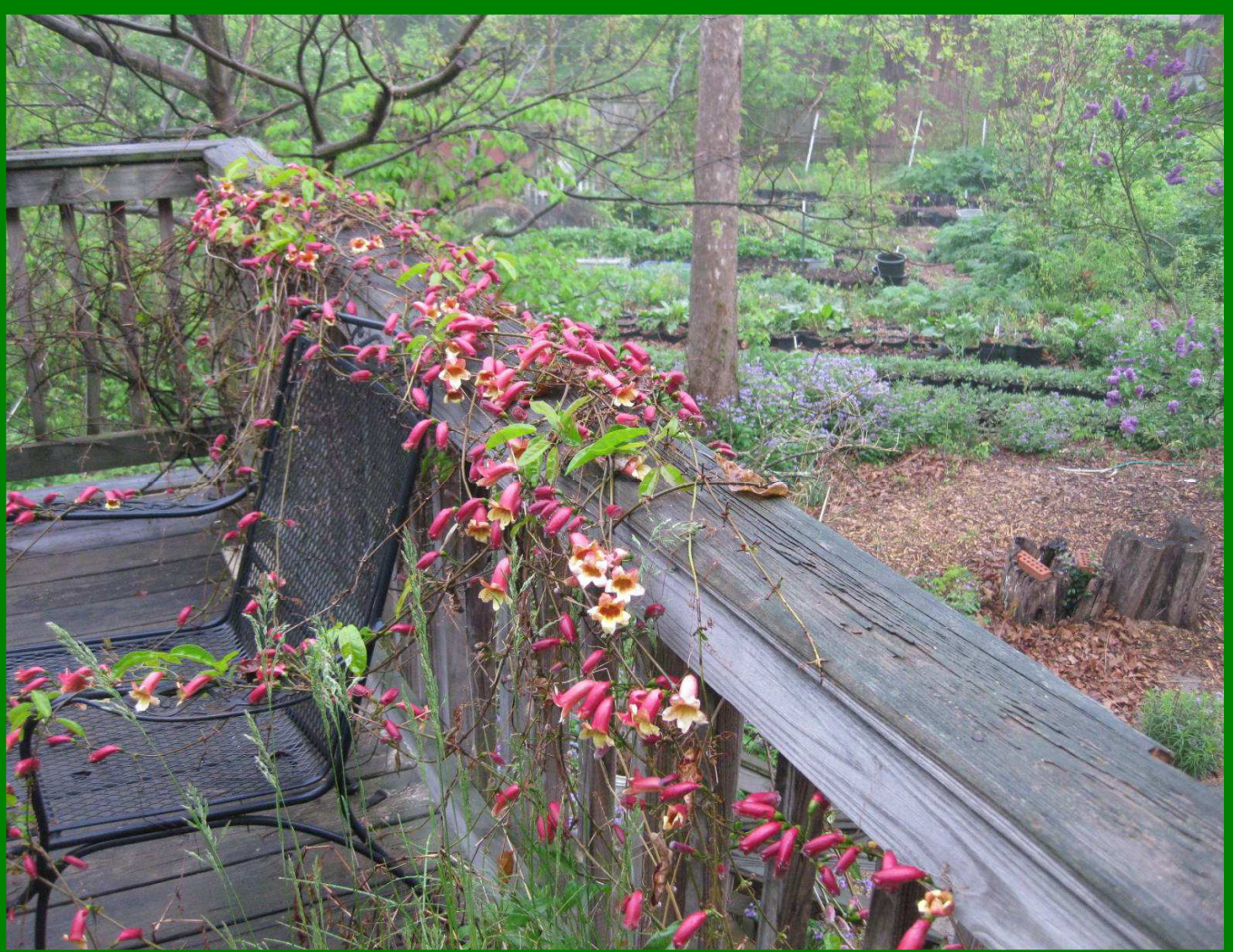
Smilax glauca: Zones 6a to 9b

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Figure 5. Forest Inventory Analysis Map of *Lonicera japonica* in southeastern states (from USDA 2010, http://srsfia2.fs.fed.us/images/srsfia/nni/a09_3101.pdf).



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PART TWO: Evergreen-tending Vines in Mid-temperate Regions

Notes on Individual Taxa

The notes below cover all evergreen or semi-evergreen vines that extend into mid-temperate regions of the Northern Hemisphere, plus brief consideration of excluded taxa. The taxonomic sequence follows current concepts of angiosperm phylogeny and its usual linear projection (Stevens 2012), except that Monocots are placed after Eudicots since they are generally more distinct in biological and ecological features, compared to the presumed common ancestors in early Magnoliid plants. The only Monocots included here are species of *Smilax*, which are the only mid-temperate woody vines without cambium; they are almost the only ones with thorns; they may have the most diverse non-alkaloidal chemistry; they have the most consistent consumption by mammalian herbivores; they are the only ones that are strictly dioecious; and they have almost the largest number of species per genus (Table 7).

Table 5 summarizes general information on the ecology of each species, including provisional information on mammalian herbivory (with listed references), and indicators of secondary chemistry. General information on ranges, zones, habits and habitats comes largely from the following sources: Flora Europaea (1964–80), Flora of North America (1993–2010), Flora of China (1996–2011), Ohwi (1965), Duncan (1967), Raulston (1992), Yuan et al. (2009), Fang et al. (2011), Weakley (2011), JSPS (2012), and Kartesz (2012). Additional references are listed under each taxon. Data on chemistry or toxicity are widely scattered in voluminous literature, and a comprehensive listing of all sources is not provided here. Much older information is summarized in Muenscher (1961), Darnley-Gibbs (1974), Levin & York (1978), Glasby (1992), Beckstrom-Sternberg & Duke (1994), Harborne et al. (1999), Burrows

& Tyr1 (2001), Hegnauer & Hegnauer (2002) plus associated databases. There has been no access to the more complete databases that exist in some institutions (e.g., Ehrman et al. 2007).

MAGNOLIID ORDERS: excluded. There are a few vines in these orders that extend into warm-temperate zones—often as evergreens, or into mid-temperate zones but there strictly deciduous or just herbaceous: Schisandraceae (*Kadsura*, *Schisandra*); Menispermaceae (*Cocculus*, *Calycocarpum*, *Menispermum*); Aristolochiaceae (*Aristolochia*, *Isotrema*). Among species of eastern North America, *Cocculus carolinus* is semi-evergreen in Zones 8–9a. It does extend into Zones 6b–7 but just as a largely deciduous plant with little or no woody stem.

RANUNCULID ORDERS

Lardizabalaceae. *Akebia* contains five species of evergreen to deciduous vines in warm- and mid-temperate zones in China, Korea and Japan. *A. trifoliata* and *A. quinata* (Figure 4) extend into Zone 7 as wild plants, and can remain semi-evergreen there except in colder winters (Raulston 1992). In Japan, taller shoots of *A. trifoliata* lose most leaves during winter, but shoots in the understory tend to retain a greater proportion of leaves, especially those formed late in the summer (Koyama & Kikuzawa 2008). *A. quinata* is widely cultivated and has escaped locally in the eastern U.S.A., tangling up the shrub layer, covering ground and suppressing native vegetation. It is able to grow through regularly mowed grass and spreads into adjacent woods (J. McCandless, Louisville, pers. comm.). In Louisville, Kentucky (currently transitional from Zone 6 to 7), *quinata* retains some leaves during most winters in gardens and parks, at least on the ground where leaf litter protects it from the coldest winds (J. Wysor, pers. comm.). Flowering is occasionally observed here but fruiting in this self-incompatible species is rare.

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Holboellia is a closely related, larger genus of evergreen vines, centered in warm- and mid-temperate zones of the Sino-Himalayan region. It is weakly separated from the more tropical genus, *Stauntonia*, in which it has been merged by Christenhusz (2012). Some species extend into montane cloud forest zones or mid-temperate zones and remain evergreen: *angustifolia* (= *fargesii*), *grandifolia*, *latifolia* and *coriacea* (Shi & Zhu 2009, Chettri et al. 2010). The only deciduous species of *Holboellia* may be *medogensis* of southeast Tibet.

There is only sparse information about mammalian browsing on *Akebia* or *Holboellia*. Some use of *A. quinata* by Sika deer has been indicated in Japan (Takatsuki & Hirabuki 1997), but this plant is reportedly avoided by white-tailed deer in North America (Jull 2001). Squirrels can feed intensively on its buds and fresh shoots during April in Japan (Setoguchi 1990). *H. angustifolia* is used to feed cattle and other livestock during the winter in Nepal (Samant 1998), and some feeding on its shoots by monkeys has been reported (Grueter et al. 2009).

Ranunculaceae: excluded. Several species of *Clematis* are evergreen-tending woody vines of warm temperate regions, but these do not generally extend into mid-temperate zones. Such species include *armandii*, *uncinata* and allies of East Asia, and *cirrhusa* of southern Europe. The species that is most persistent above ground in mid-temperate zones may be the East Asian *terniflora* (= *maximowicziana*, *dioscoriaefolia* or *paniculata* or some authors). In Zones 8–9a, it can climb to 9 m high with semi-evergreen foliage and semi-woody stems; in Zones 6a–7 it tends to be tardily deciduous, and stems usually die above ground by end of winter (Figure 4).

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ROSID ORDERS

Celastraceae. The widespread genus *Celastrus* comprises many scandent to twining woody species, but these are mostly deciduous in temperate regions (Hou 1955). *C. hindsii* is an evergreen twining shrub of mountains in Southeast Asia, native but becoming a “noxious weed” in some areas of southern China, as well as *Pueraria lobata* (Peng et al. 2009). Several other species of the genus in China and Japan are evergreen, especially section *Sempervirentes*, but most are restricted to warmer zones. The common invasive East Asian species, *orbiculatus*, is consistently deciduous, as is the North American native, *scandens*. Schaller et al. (1986) reported eating of *hindsii* by takin (a goat-antelope) in China. The genus in general appears to be relatively palatable, with potential for reduction under intense browsing (Asnani et al. 2006, Rossell et al. 2007, Ashton & Lerdau 2008, Averill 2012; but see Burroughs & Dudek, 2008).

Euonymus fortunei is a persistently evergreen vine that can climb trees and cover ground in woods (Brothers & Springarn 1992, Zouhar 2009)—and it can also spread into mowed lawns from wooded edges, becoming invasive in North America (Figure 4). Unfortunately, there is little published information in English that describes the ecology of this species in its native range of southern China, Korea and Japan. There are several related evergreen species of vine or “ascending subshrub” in *Euonymus* section *Ilicifolia*, and their taxonomy has been confused. Some reports of *E. vagans*, a “shrub or ascending subshrub to 3 m” (according to the Flora of China) may refer to *fortunei*, sensu lato (e.g., Shi & Zhu 2009). The common cultivar of eastern U.S.A. named “Manhattan” is often placed in *kiautschovicus*, which is a poorly defined species that appears intermediate between *fortunei* and *japonicus*—a shrub centered in warm temperate to subtropical zones. Cuttings from ‘adult’ climbing stems tend to remain upright and retain branch mutations in leaf shape or coloration (Dirr 1998; see also, US Patents).

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In East Asia, species of the *E. fortunei* group are locally common from montane forests to urban lowlands, especially on base-rich soils. They are often planted for revegetation of steep slopes or for ornamental uses (e.g., Wang et al. 2009, Xia et al. 2007). However, the author’s experiences in some wilder parts of Sichuan and Yunnan suggest that *Euonymus* climbers or scramblers there are largely restricted to two types of site: (1) steeper slopes and cliffs where browsing animals have less easy access—often in generally undisturbed forest (Yuan et al. 2009); and (2) fenced or walled areas near buildings. In central Japan, Sakai et al. (2002) reported it as the second-most common vine (after *Wisteria*) in a deciduous oak-dominated woods that had been recovering for three decades from burning and grazing.

From ornamental plantings of *E. fortunei* in North America, there is much anecdotal evidence that this species is often preferred for browse by deer, especially during the winter (Table 5). In Kentucky, there can be locally intense browsing by deer, beaver, cattle, horses and sheep, based on personal observations or conversations with land managers. From East Asia, there are few direct indications of browsing in English literature, but the observations of Sika deer by Takatsuki & Hirabuki (1997) are suggestive. In a caging experiment at Brookhaven, New York, Ashton & Lerdau (2008) found that *fortunei* actually grew more with exposure to deer and rabbits. But, due to competition or other complicating factors in the experiment, its growth there was generally weak—even negative in some individuals, unlike compared species; the data did show more damage from herbivory among the uncaged plants (I. Ashton, pers. comm.). The species does proliferate with mowing under some conditions, especially in partial shade, but it does not seem to invade unmowed grassland.

Outside section *Ilicifolia*, there are several other shrubby species of *Euonymus* that are described as “scandent” but these do not appear to be truly robust tall vines exceeding 3 m.

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They occur mostly in section *Echinococcus* (Flora of China Vol. 11), with both evergreen and deciduous species in mid-temperate zones, and they can be browsed (e.g., Samant et al. 2007). In eastern North America, *americanus* and *obovatus* are semi-evergreen to deciduous plants of warm/mid-temperate and mid/cool-temperate zones, respectively—and stems stay distinctly green. Both spread on or near the ground with rooting stems. *E. americanus* also grows up to become a deciduous shrub if it can escape from deer. Several reports indicate strong preference for browsing on these American species by deer and other mammalian herbivores, especially during winter: for *americanus*, Lay (1967), Blair et al. (1980), Moreland (2003), Segelquist & Pennington (1968), Thrift (2007) and Webster et al. (2005); for *obovatus*, Asnani et al. (2006). The larger deciduous shrub of North America, *E. atropurpureus*, is also among the more sensitive woody plants to mammalian browsing in eutrophic woodland, based on personal observation, Dennis (1997) and others. The invasive tardily deciduous shrub, *E. alatus*, can also become intensively browsed, despite the corky wings along its winter-green twigs.

Other Rosid families excluded. There are many vines in these families that occur in warm-temperate zones (often as evergreens), and extend into mid-temperate zones but there largely deciduous or just herbaceous: Vitaceae (*Ampelopsis*, *Parthenocissus*, *Vitis*); Anacardiaceae (*Toxicodendron*); Rosaceae (*Rosa*, *Rubus*); Rhamnaceae (*Berchemia*); Cannabaceae (*Humulus*); Fabaceae (*Lackeya*, *Pueraria*, *Wisteria*). Some roses have more persistent leaves.

A note is inserted here on the temperate mistletoes: *Phoradendron* and *Viscum* (Viscaceae) in Santalales—which phylogenetic models have rooted above the Rosid clades and below the base of Caryophyllid and Asterid clades (Stevens 2012). These epiphytic evergreen parasites on trees have some ecological similarities to the evergreen vines. They have abrupt

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northern limits that are close to the vines. Their hosts are usually deciduous trees (often late-leaving species in Kentucky), usually in thin woods on soils with moderate to high fertility. Mistletoes in general have high mineral contents and digestible carbohydrates, and perhaps highly variable protein (from 6–10% to 24% in Littlefield et al.); they are often preferred browse by larger mammalian herbivores, when access is possible (Atwood 1941, Troels-Smith 1960, Watson 2001, Dillard et al. 2005, Mathiasen et al. 2008, Umcailar et al. 2007, Littlefield et al. 2011). Their leaves have little direct structural or chemical defense, with simple phenolics, flavonols or tannins predominant. But some species have unusual proteins (thionins, lectins) that can have varied medicinal or cytotoxic effects. Mammalian toxicity is not documented among livestock or wild animals, although excessive consumption of *Viscum* by misguided humans can cause digestive problems or death in extreme cases (Kienle et al. 2011, Evens & Stellpflug 2012).

ASTERID ORDERS

Hydrangeaceae. In the small genus *Decumaria*, *sinensis* is an evergreen climbing shrub of central China, and *barbara* is a semi-evergreen vine of the southeastern U.S.A. (Figure 4). The allied genus *Pileostegia* comprises a few evergreen vines in warm-temperate zones of East Asia. The large complex genus *Hydrangea* also contains a few tall vines, but they are strictly deciduous. In all three genera, the climbing plants use adventitious roots. There is little information on herbivory of *Decumaria*, except for a few indications of moderate use by deer in North America (Moreland 2003, ?Thrift 2007). There is more information on shrubs in *Hydrangea*: *arborescens* and *quercifolia* are reported to have moderate use by deer in North America (e.g., Nixon et al. 1970, Fargione 1991, Knox 2007); and *paniculata*, much use by

Sika deer in Japan (Takatsuki 1986, Yokoyama et al. 2000). However, the popular ornamental, *macrophylla*, is reportedly toxic in Japan despite high nutritional content (Ogura 2011).

Actinidiaceae. *Actinidia henryi* [not = *A. callosa* var. *henryi*] is reportedly semi-evergreen, as are several other Chinese species of this genus, at least in warm temperate zones (Table 5, Flora of China Vol. 12). The evergreen-tendency appears to have evolved at least four different times, in different sections of the genus (Chat et al. 2004). But Japanese species are reportedly all deciduous. The allied monotypic genus, *Clematoclethra*, is a strictly deciduous vine that occurs only in mid-temperate to subalpine zones of China. Schaller et al. (1986) reported eating of *A. henryi* by takin in China, and there are several reports of the deciduous species being eaten by various mammals (Table 5).

Bignoniaceae. *Bignonia* is considered by some authors to contain just one species, *capreolata* (Weakley 2011), but some uncertainty remains (Lohmann 2006, Olmstead et al. 2009). It reaches an abrupt northern limit in southern Indiana and southern Ohio—with a similar range to *Smilax bona-nox* (Figure 4) and other notable evergreens such as the mistletoe, *Phoradendron leucarpum*, and the bamboo, *Arundinaria gigantea*. Yet at the edge of its interior range in southern Illinois, Indiana and Ohio, the species is locally proliferating on south-facing bluffs and terraces (D. Nickrent & D. Boone, pers. comm.). It climbs with tendrils that develop into pads (as reviewed by Seidelmann et al., 2012), and adventitious roots are formed on lower stem sections after 2–3 years (pers. obs.). It can also spread widely on the ground, but does not form dense mats of leaves like *Euonymus* or *Hedera*. The plant can be preferred browse for some mammals in the winter, a phenomenon that is particularly well-documented in the case of swamp rabbits (Table 5). There is little information on use by deer—the few reports indicate

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only moderate usage. But the plant has among the best-tasting leaves of any evergreen vine in this author's experience (rivaling *Smilax* but much more fibrous).

Araliaceae. *Hedera* is an evergreen vine with adventitious roots, flowering in late summer to fall. Its species are concentrated in warm- to mid-temperate zones, with a center of diversity in the Mediterranean region (Ackerfield & Wen 2003, Green et al. 2011). The widespread European *H. helix* (sensu stricto) is most common in western regions with relatively mild winters (Metcalfe 2005). Towards the east, it is more restricted to lowland riparian habitats, where cold winters appear to limit its height. In North America, *helix* (a tetraploid) has become locally abundant at scattered sites across southeastern states, with much local vegetative spread (Figure 4, Okerman 2000, Waggy 2010). The closely related species, *hibernica* (an octoploid), is concentrated in regions with more oceanic climate, and has become the prevalent taxon in western North America (Clarke et al. 2006). In more interior regions of North America, *Hedera* has not been able to tolerate winters as a high vine, although it may persist as a sterile ground-cover. In Kentucky, there has been virtually no evidence of establishment from seed until recent years—seedlings have begun to appear in gardens of the Lexington area during 2000–2012 (as observed by this author and Richard Weber, Springhurst Garden).

Other species of *Hedera* occur in warm- to mid-temperate regions further east: *colchica* and *pastuchowii* in northern Turkey and the Caucasus region; *nepalensis* in the Sino-Himalayan region; *rhombica* in Taiwan and Japan. *H. nepalensis* is becoming widely used in Chinese horticulture. It may have less tolerance of shade and drought than *Euonymus fortunei* (Jiang et al. 2007, Chen & Wang 2008), but it is reportedly more tolerant of SO₂ pollution than most ornamental plants in Chinese cities (Chen et al. 2007).

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Although *Hedera* is quite bitter, there is ample evidence that it can provide relatively nutritious browse for ruminants, especially during winter (Table 5). Its general concentration within hedgerows, hawthorn scrub and younger woods across Britain (Rackham 2003) can be partly interpreted in terms of escape from herbivory. During medieval times, it was common practice in Europe for *H. helix* to be cut down for feeding to livestock during the winter (see also Troels-Smith, 1960). Cattle can eliminate it from the forest floor after several years of concentrated influence (Uytvanck & Hoffmann 2009), and goats have been used recently to reduce the invasion of *Hedera* in Oregon (Ingham 2008). But there is much variation in usage. Boulanger et al. (2009) found the plant to be generally avoided by deer, with an index based largely on summer browsing. And outside deer-exlosures, Rossell et al. (2007) found more reduction in *Lonicera japonica*, and much more in *Celastrus orbiculatus*. In East Asia, *H. nepalensis* is also a traditional source of fodder for livestock in the Sino-Himalayan region (Bajracharya et al. 1978, 1985). It is probably a significant winter-forage for wild ungulates (e.g., Shah et al. 2009), but rhesus monkeys appear to avoid it (Goldstein & Richards 1989).

The general ecology of *Hedera helix* is well-known within its native Europe (Metcalf 2005). Schnitzler & Heuzé (2006) have documented its association with nutrient-rich soils on floodplains of the Rhine, as well as its sensitivity to waterlogging. The plant maintains a relatively constant, conservative flow of water to leaves, in marked contrast to associated trees (Leuzinger et al. 2011). Heuzé et al. (2009) have explored how growth responds to annual variation in temperature, indicating that warmer summers have most effect on uplands, while increases in Feb–Mar temperature have most effect on lowlands. There has been considerable research on the photosynthesis of *H. helix*. Oberhuber & Bauer (1971) initially found that “adult ivy leaves possess an enormous capacity to repair light-induced damage to the photosynthetic apparatus in winter” (see also Bauer & Kofler, 1987); this allows growth to

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resume in early spring before trees leaf out. The maximum photosynthetic rate of *H. helix* is relatively low compared to other vines, including *L. japonica* (Carter & Teramura 1988). However, ‘adult’ leaves on climbing shoots have distinct morphology and physiology, with higher photosynthetic rates per leaf area than ‘juvenile’ leaves on lower shoots, largely due to increased leaf thickness (Bauer & Bauer 1980). But ‘adult’ leaves are less able to adjust physiologically to short-term changes in light (Bauer & Thoni 1988). Zotz et al. (2006, plus cited studies) found that *H. helix* responds much to increases in CO₂—more so than associated shrubs and trees. They measured more stimulation of photosynthesis by CO₂ in the forest understory (where sugar content remained low), as compared to plants high on trees (where sugar content was already higher and increased with more CO₂).

Caprifoliaceae. *Lonicera japonica* is largely evergreen, as well as allies that include *acuminata*, *etrusca*, *ferruginea* and the *macrantha* complex. These plants are mostly twiners of Southeast Asia that extend into mid-temperate zones, where they are locally common in mixed successional deciduous forest (e.g., Lee et al. 1990). The only native species elsewhere is *biflora*, which occurs in scattered parts of the Mediterranean region but is reportedly rather short (< 3 m) and deciduous; it is not included in Table 5. *L. japonica* is widely cultivated, and it has become an abundant invasive alien in North America—while its native congeners tend to decline in some areas (Stransky 1984, Hardt 1986, Nuzzo 1997, Clemants & Moore 2005). *L. acuminata* sensu lato (including *henryi*) has a more western range than *japonica*, largely Sino-Himalayan. It has been cultivated under the name *henryi* in Europe, where it has recently become invasive at several sites (Weber 2005, Verloover 2006).

Mammalian toxicity from *L. japonica* is unknown, and standard tests on rats have revealed no pathological effects after high doses of leaf extract (Thanabhorn et al. 2006). In North

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America, the plant is well known to be eaten much by deer and cattle during hard winters, for which it has been promoted (e.g., Handley 1945, Noland & Morrison 1954, Segelquist & Rogers 1975, Stransky 1984, Hardt 1986, Dyess et al. 1993, Frederick & Kennedy 1995). Even in summer, there is evidence that it can be used more than other vines (Nixon et al. 1970, Ashton & Lerda 2003). Dense deer populations appear to reduce the plant's abundance (e.g., Beaver 2011). However, *japonica* is able to regrow rapidly after damage (Schierenbeck et al. 1994), and it is relatively plastic in its overall growth rate and form (Schweitzer & Larson 1999). Sasek & Strain (1991) found that its growth-response to CO₂ enrichment was greater than that of the native *sempervirens*.

The native *Lonicera* vines of North America and Eurasia—in subgenus *Caprifolium*—are generally rather short and semi-evergreen to deciduous in mid-temperate regions. *L. sempervirens* is largely evergreen in warmer zones of the southeastern U.S.A., but it usually reaches only 3–4 m. It seems to have spread north into mid-western states during recent decades (Kartesz 2012, see also state floras), but it may be limited by intensive deer-browsing in northeastern states (Table 5; Clemants & Moore 2005). In two caging experiments, *sempervirens* was more damaged than *japonica* by deer or rabbits (Schierenbeck et al. 1994, Ashton & Lerda 2008). *L. albifolia* of southwestern U.S.A. and Mexico (Nelle 1996), and the European species (*implexa*, *etrusca*, *caprifolium*, *periclymenum*) are also favorites of deer and livestock (Appendix 4).

In South Carolina, Schierenbeck & Marshall (1993) found that both *sempervirens* and *japonica* produce new leaves during January. But *sempervirens* abscised its old leaves during mid-December, while *japonica* retained its old, photosynthetically active leaves through March. Moreover, the new leaves of *japonica* had significantly higher photosynthetic rates than

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sempervirens during January—although for most of the growing season there was no significant difference in sun or in shade. The greater shade tolerance of *japonica* may involve morphological differentiation rather than photosynthetic rates, since plants exhibit a somewhat distinct prostrate growth form on the ground, without twining behavior (Larson 2000). Yet *japonica* is also more responsive to the presence of climbing structures, as measured by morphological changes and increase in overall biomass (Schweitzer & Larson 1999).

Other Asterid families excluded. There are several low vines or subshrubs affiliated with Gentianales (sensu lato) that extend into warm-temperate zones—often as evergreens—but much less into mid-temperate zones—where strictly deciduous or restricted to ground vegetation. Families include Loganiaceae (*Gardneria*), Gelsemiaceae (*Gelsemium*), Apocynaceae (*Trachelospermum*, *Thyrsanthella*, *Vinca*) and Oleaceae (*Jasminum*). Most have bitter iridoids or alkaloidal toxins (with Levin indices of 3 to 6) and may be generally avoided, e.g., *Jasminum* according to Le Houérou (1980). But deer are able to browse some species to a moderate or high degree (e.g., Samant et al. 2007), especially *Gelsemium sempervirens* in the southeastern U.S.A. during winter (e.g., Atwood 1941, Lay 1967, Blair & Burnett 1980, Thill 1984, Thill & Martin 1989, William & Baxley 2008, Wade & Mengak 2010). *Thyrsanthella difforme* is a reasonable North American segregate of *Trachelospermum* (Livshultz et al. 2007). It extends far into Zone 7, and is occasionally browsed (Thill 1984). *Vinca minor* has anomalous growth form, being an evergreen trailing subshrub of mid- to cool-temperate zones in southern Europe, and now widely planted elsewhere. It can spread to dominate the forest floor, with much potential to suppress tree seedlings (Darcy & Burkhart 2002). It has some reputation for toxicity, and there is no more than occasional use by deer (e.g., Sotala & Kirkpatrick 1973, Heinrich & Predl 1993, Jett 1995, Jull 2001). However, cattle can largely eliminate it after several years of grazing and trampling (Uytvanck & Hoffmann 2009).

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MONOCOT ORDERS

Smilacaceae. In the large widespread genus *Smilax*, many species climb and hang with tendrils and thorns, but few have a pronounced evergreen tendency in zones colder than warm-temperate. In eastern North America, the ‘green-briars’ of east-central states are evergreen to deciduous vines: *bona-nox*, *glauca*, *hispida* (= *tamnoides* var. *hispida*), and *rotundifolia*. *S. bona-nox* has the strongest tendency to evergreenness. At the northern edge of its range this species is largely restricted to south-facing calcareous slopes, where it often hangs off the sides of the sheltering evergreen tree, *Juniperus virginiana*. In Kentucky, *glauca* also has some persistent leaves well above the ground during most winters. Leaves of *hispida* leaves stay green only on low shoots (< 1 m high). Leaves of *rotundifolia* leaves are generally all dropped during Oct–Nov, but its stems stay quite green all winter (like several species of *Euonymus*)—more so than its congeners. Species of *Smilax* are well-known to provide frequent browse for deer and other large herbivores in the eastern U.S.A., especially on fresh vigorous growth in spring or after cutting, burning or thinning of the woods (Table 5). However, seasonal and spatial variation in abundance or palatability may often obscure the general preferences (e.g., Halls 1975, Blair & Brunett 1980, Crimmins et al. 2010). Dense deer populations can cause large reductions in cover of *Smilax* (e.g., Beaver 2011). Rabbits can also be a major herbivore on fresh shoots, especially after fires that stimulate much new growth, sometimes leading to declines in cover (e.g., Niering & Dreyer 1989).

In mid-temperate regions elsewhere, there are a few other high-climbing *Smilax* species. (a) *S. californica* of the western U.S.A. is reportedly semi-evergreen but rarely gets more than 3 m tall (Nevin Smith, Suncrest Gardens, pers. comm.).

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(b) *S. aspera* of the Mediterranean region and western Asia is evergreen, but it does not get higher than 4 m or extend into mid-temperate zones. It is often browsed by livestock, and young shoots are traditionally eaten by humans (D’Antuono & Levato 2003, Samant et al. 2007; but see Lev-Yadun 2009).

(c) *S. ferox* and *megalantha* of the Sino-Himalayan region climb to 5 m or more and extend into cool-temperate zones, but it is not clear if they have persistent leaves.

Additional East Asian species may extend into mid-temperate zones, but, again, their degree of evergreenness remains obscure. Samant (1998) reported use of *ferox* and *vaginata* by livestock in the Himalayas. (*S. vaginata* is one of several East Asian species in the genus that are low unarmed shrubs, often resembling deciduous *Vaccinium* species of North America.)

Other Monocot families excluded. Dioscoreaceae (*Dioscorea*) are only herbaceous in mid-temperate zones. Poaceae include some scandent evergreen bamboos but only in warmer temperate zones (*Ampelocalamus*).

Table 5. Features of range, habit, habitat, mammalian herbivory and secondary chemistry for evergreen and semi-evergreen vines that extend into mid-temperate zones.

Abbreviations in each column are as follows.

Range and Habitat: abbreviated regions.

Chi = China; Him = Himalayan region; Jap = Japan; Kor = Korea; Tai = Taiwan.

Plus following data.

HZ: Hardiness Zones where most of the species occurs in the wild.

AL: Altitude above sea level in Sino-Himalayan region and southern China.

Habitats descriptors, based on varied sources, with some added standardization.

Habit Column: following sequence of data

EG = evergreen leaves; SEG = semievergreen; DEC = deciduous; underlined species have more or less coriaceous (leathery) leaves (versus papery or membranaceous).

Typical maximum height in m

ADH = adventitious roots and specialized holdfasts formed on tree trunks.

ADR = adventitious roots usually formed along stems that climb tree trunks.

TEN = climbs with tendrils

THO = thorny, which sometimes enhances climbing ability

TWI = climbs only with twining stems (no roots)

Herbivory Column: references to mammals eating leaves or stems of the plant.

BUT/NOT: indicates that partial/general avoidance is indicated by the research

* Asterisks indicate sources with data on protein contents; see “pr” in last column.

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Chemistry Column: classes of compound reported from the species or its relatives.

? indicates reports only from related species.

Top line indicates predominant types of secondary chemicals as follows:

C = condensed tannins (but not always distinguished from hydrolysable tannins)

N = alkaloids or other nitrogen containing compounds, and their derived glycosides; numbers after / are the alkaloidal toxicity indices of Levin & York (1978).

P = phenolics (including tannic acids, hydrolysable tannins, flavonoids, lignans, stilbenes) and furans, or their derived glycosides.

T = terpenoids (including iridoids, saponins) and their glycosides, lactones, etc.

S = sterols or steroids and their derived glycosides.

TOX: severe (non-digestive) toxicity reported in mammals, with chemical cause if known; excluding reports clearly attributed to fruits or seeds (i.e., perhaps the source of reports in *Celastrus* and *Euonymus*)

TAS: provisional description of human taste, from this author's reaction.

pr = crude protein content of leaves, with references in Herbivory Column indicated by asterisks; where possible, these are from leaves sampled in summer (not fresh spring shoots); crude protein is calculated as N content \times 6.25, but is often an overestimate of the digestible protein (Conklin-Brittain et al. 1999).

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TAXA	RANGE	HABIT	HERBIVORY	CHEMISTRY
RANUNCULID				
Lardizabalaceae <i>Holboellia angustifolia coriacea, grandiflora and latifolia</i> (genus combined with <i>Stauntonia</i> by some authors)	seChi Him HZ 6-8? AL (10)15-27 mesic mountain forest edges, open hillsides, stream-sides	<u>EG</u> 5-10+m? TWI Monoecious	cattle+ (Samant 1998); monkeys (Grueter+ 2009, Fan+ 2009, Huang+ 2010*)	? TP (based partly on <i>brachyanthera, chinensis, hexaphylla</i>) ?phenolics ?triterpenoid-glycosides ?saponins pr = 7-12 (Kusumoto+2012)
Lardizabalaceae <i>Akebia quinata, trifoliata</i> and allies	eChi Jap HZ 7-8? AL 2-21 mesic open forest, scrub	<u>SEG-DEC</u> 5-12m TWI Monoecious	? deer (Takatsuki+ 1997; BUT Jull 2001) squirrel , esp. Apr (Setoguchi 1990) ?monkey (Iwamoto 1982* just fruits/seeds)	T triperpenoids saponins TAS: strongly bitter pr = 10%*
ROSID				
Celastraceae <i>Celastrus hindsii</i> and perhaps allied species (<i>virens, monospermus, ?homaliifolius</i>), but these are mostly in warmer zones	SE Asia HZ 7-9? AL 3-25 forests, thickets (?mesic to sub-xeric)	<u>EG</u> 5-19m TWI ?Dioecious	takin (Schaller+ 1986*) ?monkeys (based on <i>orbiculatus</i> , Nakagawa 1989; and on <i>gemmatus</i> , Fan+ 2009) ?deer (based on <i>orbiculatus</i> , Rossell+ 2007; Averill 2012) BUT note also reported	? TPNS (partly based on <i>angulatus, hypoleucus, orbiculatus, scandens</i>) phenolics (rosamarinic acid) ?flavonoids ?diterpenoides triterpenoids saponins sesquiterpene-

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			toxicity of <i>scandens</i> to horses (Schaffner 1904) and low-moderate preference by deer (Fargione+ 1991, Wade+ 2010)	-esters and -pyridine alkaloids glycoalkaloids (solanine) ?TOX: solanine (glycoside of an indolizidine alkaloid) ?pr = 10%*
Celastraceae <i>Euonymus fortunei</i> and perhaps allies (<i>kiutschovicus, vagans</i> of some authors, <i>bockii, ?pseudovagans, ?kengmaensis, ?hupehensis, ?theacola</i>) but most of these species occur in warmer zones or do not usually exceed 3 m in height)	East Asia HZ 6-8? AL 0-34 mesic to subxeric forests, scrub, urban areas	<u>EG</u> 10-20m ADR	deer , esp. winter (Burroughs+ 2008, Conover+ 1988, 1995, Fargione+ 1991, Masters+ 1991, Masters+ 2004, ?Takatsuki+ 1997; ?NOT Ashton+ 2008); cattle (JC+ pers. obs., ?Pan+ 2005*); beaver (JC+ pers. obs.); horses (JC+ pers. obs.); sheep (JC+ pers. obs.); rabbits (Stafne+ 2005); monkeys (Nakagawa 1989)	TPNS (based partly on <i>alatus, americanus, atropurpurea, europaeus, japonicus</i> , etc.) phenolics, flavonoids lignans; ?furans sesquiterpene-esters -pyridine alkaloids triterpenoids ?sesquiterpenoids ?steroids (cardenolide) ?steroidal glycosides ?TOX: cardenolide TAS: mealy-bitter ?pr = 9-12%*

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ASTERID				
Hydrangeaceae <i>Decumaria sinensis</i>	eChi HZ 7-9 AL 6-13 mesic thickets on slopes, rock crevices	<u>EG</u> 2-5m ADR	?monkeys (based on allied genus <i>Schizo- phragma</i> ; Nakagawa 1989, Enari+ 2010)	?PT phenolics, flavonoids ?saponins ?glucosides ?pr = 9% (Kusumoto+ 2012)
Hydrangeaceae <i>Decumaria barbara</i>	seUSA HZ 7-9 mesic forests	<u>SEG</u> 4-10m ADH	?deer (“moderate” Moreland 2003, ?Thrift 2007)	?PT (see other Hydrangeae) ?flavonoids ?glycosides
Actinidiaceae <i>Actinidia henryi</i> and perhaps <i>rubricaulis</i> (but excluding species of warmer zones: <i>cylindrica</i> , <i>zhejiangensis</i> , <i>rufotricha</i> , <i>fulvicoma</i> , <i>liangguanensis</i>)	seChi HZ 7? AL 14-25 mesic mountain forests, thickets	<u>SEG</u> 15m? ADR/TWI Poly-Dio.	henryi: takin (Schaller+ 1986). Also following deciduous species. arguta: deer (Takah.+ 2001, Takatsuki 2009); monkeys (Enari+ 2010) deliciosa: deer (Palomo 2012) pilosula: monkeys (Grueter+ 2009) polygama: monkeys (Enari+ 2010)	?TPCN (from <i>chinen- sis</i> , <i>polygama</i> , <i>rufa</i>) ?tannins, ?flavonoids (benzopyran) ?saponins ?triterpenoids ?monoterpenoids ?actinidine (from pyridine)/2.0 ?pr = 12% (Kusumoto+ 2012)
Araliaceae <i>Hedera rhombea</i>	Tai Kor Jap HZ 6-8? mesic forests, edges	<u>EG</u> 10+m ADR	deer (Takatsuki 1988, ?Takatsuki+ 1997)	T (see also <i>helix</i>) triterpenoids saponins

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Araliaceae <i>Hedera nepalensis</i>	seChi Him HZ 6-8? AL (0)12-26(35) mesic forests, rocky slopes	<u>EG</u> 10+m? ADR	cattle+ , esp. winter (Bajracharya+ 1985, Samant 1998) ?deer (Shah+ 2009); NOT rhesus monkeys (Goldstein+ 1989)	T (see also <i>helix</i>) saponins monoterpenoids sesquiterpenes
Araliaceae <i>Hedera pastuchowii</i> and <i>H. colchica</i>	Caucasus HZ 6-7? mesic forests, edges	<u>EG</u> 5+m ADR		?T (based on <i>helix</i>)
Araliaceae <i>Hedera helix</i> group, including <i>H. hibernica</i>	WC Europe HZ (6)7-8(9) mesic forests, cliffs, walls	<u>EG</u> 10-30m ADR	deer , esp. winter (Burroughs+ 2008, Fargione+ 1991, Gill+ 2001, Gonzalez-H.+ 1996, 1999*, Kirby 2001, Masters+ 1991, 2004, McEvoy+ 2006, Metcalf 2005*, Pettorelli+ 2001, 2003, Rackham 2006, Tixier+ 1997, Krafft 2011, Perrin+ 2011; NOT Papageorgiou+ 1981* in summer, Boulanger+ 2009) cattle+ in winter (Metcalf 2005, Troels- Smith 1960, Uytvanck+	TP phenolics monoterpenoids polyacetylenes (falcarinol) saponins (TOX): saponins and falcarinol TAS: strongly bitter pr = 9%*

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			2009) goats (Bartolomé+ 1998, Ingham 2008, Rogosic+ 2006) sheep (Metcalf 2005, Rogosic+ 2007)	
Bignoniaceae <i>Bignonia capreolata</i>	seUSA HZ 7-9 mesic to subxeric forests, thickets	EG 10-25m TEN-ADH (+ADR)	rabbits , esp. winter (Fowler+ 2007, Smith 1982*, Toll+ 1960). deer , moderate use (Atwood 1941, Lay 1967, Wolters+ 1977, Moreland 2003, Wade+ 2010; Blair+ 1980)	?TPS (see other Bignoniaceae) ?phenolics ?triterpenoids ?iridoid- glycosides ?steroids TAS: mealy-sweet (invigorating) pr = 13.5%*
Caprifoliaceae <i>Lonicera sempervirens</i>	seUSA HZ 6-9 submesic thickets, edges	SEG (varies) 3-7m TWI (weak)	deer : high-medium use but variable (Schierenbeck+ 1994, Ashton+ 2008, Jull 2001, Wade+ 2010)	PT phenolics, flavonoids iridoids TAS: slightly bitter, slightly sweet ?pr = 7-11% (Apx. 4)
Caprifoliaceae <i>Lonicera japonica</i> or allies (<i>affinis</i> , <i>hypoglauca</i> etc.)	eChi Jap+ HZ 6-9 AL 8-15 mesic to subxeric, thin	EG-SEG 5+m TWI	deer , esp. winter (Handley 1945, Lay 1967, Moreland 2003, Shierenbeck+ 1994, Ashton+ 2003, Nixon+	TP phenolics, flavonoids saponins iridoid-glycosides sitosterol

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	forests, scrub, rocky ground		1970, Sotala+ 1973, . Wolters+ 1977, Thrift 2007, Beaver 2011*; NOT Shaw 2008* in summer); monkeys (Nakagawa 1989*, Go 2010, Huang+ 2010*); ?squirrels (Setoguchi 1990)	?other glycosides TAS: mealy-bitter (more acrid than <i>sempervirens</i>) pr = 8-15%* (also Segelquist+ 1975, Dyess+ 1993, Jones+ 2008, Kusumoto+ 2012 Lashley 2009; higher includes fertilized)
Caprifoliaceae <i>Lonicera acuminata</i> , or allies (<i>henryi</i> etc.)	sChi Him HZ 6-9? AL 1-32; mesic forests & scrub	SEG 15+m TWI	takin (Schaller+ 1986); monkeys (Bleisch+ 1998*, Xiang+ 2007, Grueter+ 2009)	?TP (see also <i>japonica</i>) phenolics etc. pr = 12%*
MONOCOTS				
Smilacaceae <i>Smilax ferox</i> , <i>S. megalantha</i>	seChi HZ 5-8? AL 9-34 thin forests, thickets	SEG? 5m? TEN+THO Dioecious	cattle+ (Samant 1998); ?monkeys (Huang+ 2010*; <i>Heterosmilax</i> , Fan+ 2009).	?PTSC (based largely on <i>china</i> , <i>chinensis</i> , <i>officinalis</i> , <i>glabra</i>) phenolics (quercetin, glucopyranosides) ?tannins saponins, steroids ??tropane-alkaloid (scopolamine) pr = 13*

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Smilacaceae <i>Smilax</i> species esp. <i>bona-nox</i>	HZ 6-9 bon: seUSA	TEN+THO Dioecious <u>EG-SEG</u> 5-8+ m	deer (Beaver 2011*, Castleberry+ 1999, Dillard+ 2005, Halls 1975, Halls+ 1982*, Klein+ 2010, Korschgen+ 1980, Lay 1967, Lashley+ 2011, Lopes+ 1984, Masters+ 2004, Moreland 2003, Nixon et al. 1970, Shaw+ 2010*, Soper+ 1993*, Sotala+ 1973, Thill+ 1984, 1989, Thrift 2007, Wade+ 2010; BUT Blair+ 1980) cattle (Thill+ 1984, 1989, Wolters+ 1977); rabbits , esp. Apr (Niering+ 1989)	PTCS phenolics tannins ?steroidal saponins TAS: mealy to slightly bitter then slightly sweet (sweetness pronounced in <i>bona-nox</i> but more of an aftertaste in <i>glauca</i>) pr = 9-15+% (more if fertilized or in spring*; also Torgenson+ 1971, Jones+ 2008)
also <i>glauca</i>	gla: seUSA	<u>SEG</u> 5+m		
not <i>hispida</i>	his: seCan-USA	SEG-DEC 7+ m		
not <i>rotundifolia</i>	rot: seUSA+ thin forests, thickets, old fields (esp. on subxeric to xeric sites in north, but also low- lands in south)	SEG-DEC 5-6+m		

Mammalian Herbivory and Secondary Chemistry

The foliage of most evergreen or semi-evergreen vines in mid-temperate regions is eaten by varied mammalian herbivores (Table 5). Only *Hedera* is sometimes reported to be toxic for generalist herbivores, due to its saponins, but even this plant has been much used during the winter within its native range. Cattle, goats and sheep are able to consume large amounts, especially in combination with other plants that contain tannins (Rugosic et al. 2006, 2007). There are no regular published reports of severe disease or mortality from mammals eating *Hedera*—although its bitterness would probably prevent excessive consumption by mammals not adapted to the plant. Some potentially toxic compounds are known from *Celastrus* and *Euonymus* (Table 5), but there are no regular reports of toxicity from the few evergreen vines within these two genera. In the case of *E. fortunei*, there are zero published reports of toxicity to wild mammals or livestock, despite much exposure during recent decades within North America.

For an initial assessment of chemical variation in these plants, it is useful to recognize the following general classes of compounds, as detailed in Table 5; see also Harborne (1991a, 1999), Wink & Schimmer (2010), etc., for general background.

Phenolics/flavonoids and derived glycosides (including tannic acids, stilbenes, lignans, catechols, quinones). Varied types are known in most of the ten genera with evergreen or semi-evergreen mid-temperate vines, but they appear less developed in *Akebia* and *Hedera*. These compounds have diverse physiological effects on mammals, but severe to fatal reactions are

uncommon in the wild. Lignans are estrogen-like and some are toxic. Catechols, quinones and their derivatives are more often highly irritant or toxic to mammalian physiology.

Condensed tannins (polyphenols based on flavone units). These have been reported only from *Actinidia*, *Lonicera* and *Smilax*. Tannins—especially condensed tannins—are large molecules that bind to proteins and generally reduce the digestibility of vegetation (e.g., Adams et al. 2009). However, ruminants such as deer may seek to ingest small amounts of hydrolysable tannins (esters of sugars and tannic acids) for benefits in digestion (Verheyden-Tixier & Duncan 2000).

Terpenoids and derived glycosides/lactones (including saponins and iridoids). These diverse deterrent compounds have been found in all 10 genera except *Decumaria* and *Bignonia* (*sensu stricto*), where they are expected. Secondary chemistry remains virtually unexplored in those two genera, but terpenoids are known from close tropical relatives of *Decumaria* in *Pileostegia* and *Schizophragma* (Hufford et al. 2001, Li 2011), and of *Bignonia* in *Tanaecium* and *Dolichandra*, etc. (Von Poser et al. 2000, Mitaine-Offer 2002, Olmstead et al. 2009, Choudhury et al. 2011). As I can attest for *Akebia* and *Hedera*, terpenoids tend to be more aggressively bitter than tannins. Negative physiological effects in mammals are well-known (e.g., Harbourne 1991b, Langenheim 1994, Nyahangare et al. 2012). Sesquiterpene lactones occur widely in vascular plants (from *Ginkgo* to *Artemisia*), and are often foul-smelling. Saponins are triterpenoids that have been shown to limit mammalian herbivory in many cases. Yet saponins can often inhibit urease in the gut, reducing production of ammonia and methane (e.g., Veit et al. 2011). Ruminants—especially goats—are much less affected by saponins, although digestive problems and abortions can still result from large amounts in the diet (e.g., Cheeke 1971, Francis et al. 2002, Rugosic et al. 2006, 2007). Iridoids are monoterpenes typical of

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asterid taxa that have much general deterrent activity and other physiological effects in mammals.

Sterols/steroids and derived glycosides. These are generally derived from triterpenoid precursors, and share some biosynthetic steps with saponins. Among genera considered here, such chemicals are known from *Celastrus*, *Euonymus*, *Lonicera* and *Smilax*, and may also be expected in *Bignonia*. Some of these compounds have intense physiological effects on mammals, especially the cardiac glycosides (including cardenolides)—which stop hearts. However, specific analyses of the vines listed here have revealed no more than benign sterols/steroids: none in *C. hindsii* (e.g., Sung et al. 2008); sitosterols in *E. fortunei* and *L. japonica* (or allies), which can reduce cholesterol (e.g., Qu 2001); and steroidal saponins in several species of *Smilax*, which can reduce blood sugar, dementia and cancer (Challinor et al. 2012, Zhang et al. 2012).

Alkaloids and diverse other nitrogen-containing compounds. These are unknown in the 10 temperate genera, except *Actinidia* (e.g., Maddumage et al. 2013), *Celastrus* (e.g., Feng et al. 2007) and *Euonymus* (e.g., Ying et al. 2011), which have pyridine derivatives (actinidine etc.) or sesquiterpene-pyridine alkaloids. These types of compounds tend to have significant inhibitory effects on insects, tumor cells and viruses. Some exhibit attractant, irritant or neuroleptic-like effects on mammals, but without mortality (e.g., Sousa & Almeida 2005). Other groups of alkaloids, not found at all in these 10 genera, are well-known to reduce herbivory or cause mortality in mammals (Levin & York 1978, Harborne 1999).

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Relationships of Chemistry to Life-form

Evergreen versus deciduous taxa. Genera with vines that are largely deciduous in mid-temperate zones have a different chemical spectrum and a greater variety, especially in alkaloids or other nitrogen-containing compounds (Table 6). Such compounds are known or suspected in about 9 of the 21 genera, mostly belonging to Magnoliid or Ranunculid families and Fabaceae. However, there are no reports of pyridine alkaloids as found in Celastraceae. Among phenolics, condensed tannins are known only in *Humulus* and *Wisteria*; tannic acids are common in Vitaceae and *Toxicodendron*; and the latter is also infamous for its allergenic derivatives of catechol. Ten of the 21 genera have well-developed terpenoid chemistry, but concentrated in Asterid families. *Berchemia*, *Clematoclethra*, *Cocculus* and *Humulus* are known to produce sterols or steroids. These trends are summarized further below (Table 7).

Vines in general versus shrubby relatives. Within the following families or genera with woody plants that mostly have a climbing habit (evergreen or deciduous), current information on phylogenetic patterns indicates a halt or loss in evolution of some significant defensive compounds. This trend can be explored through comparisons with their closest non-climbing relatives, following Gianoli (2004), Stevens (2012) and other relevant sources.

(1) **Schisandraceae.** All species are vines, comprising deciduous *Schisandra* and evergreen *Kadsura*. There are no reports of alkaloids or mammalian toxicity; moderate use by deer is reported from Japan (Yokoyama et al. 2000). The closely allied Illiciaceae (*Illicium*) are all shrubs or small trees with mammalian toxicity, due to sesquiterpenes or the protoalkaloid skimmianine (Nakamura et al. 1996, Tsuji & Takatsuki 2004, Wang et al. 2011).

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(2) **Lardizabalaceae.** Almost all species are vines, mostly high-climbing evergreen-tending species, and they lack benzylisoquinoline alkaloids. But these alkaloids are generally found in the allied clade that comprises the much shorter climbers in Menispermaceae (deciduous in temperate regions) plus non-climbers in Berberidaceae and Ranunculaceae (Stevens 2012).

(3) **Celastraceae: *Euonymus*.** Section Illicifoliae (including *fortunei*) are mostly scandent shrubs to high climbers, generally evergreen and with diverse secondary chemicals, but there are no reported significant problems or intense physiological effects in mammalian herbivores (Table 5). Other sections of the genus are all non-climbing shrubs and small trees (mostly deciduous in temperate regions), including two well-known Old World species that have added chemical defenses such as cardenolides. *E. europaeus* also produces gut-damaging lectins that may reduce herbivory by rabbits; it is just moderately preferred by generalist herbivores (Hart et al. 1988, Motta 1996, Boulanger et al. 2009, Thomas et al. 2011). *E. alatus* can have varied medicinal to cytotoxic effects in mammals, partly due to cardenolides (e.g., Kitanaka et al. 1996, Kim et al. 2009). It may be preferred much less than *fortunei* during the winter in North America (Conover & Kania 1988; but see Fargione et al., 1991), although moderate use by deer has been reported in its native range (Yokoyama et al. 2000). As noted above, however, the North American shrubs remain highly palatable (*americana*, *atropurpurea*, *obovatus*), so this trend from climbers to shrubs within *Euonymus* is not consistent.

(4) **Anacardiaceae: *Toxicodendron*.** The only climber in this genus is *radicans* (sensu lato)—a widespread deciduous vine of southern China and eastern North America. Despite the dermatitis in humans that is often caused by its catechol derivatives, “Cattle, horses, sheep, hogs, goats, birds and many other animals appear completely immune to poison ivy” (Mulligan & Junkins 1977, and their citations). Moreover, deer and other herbivores (moose, cattle, goats, deer,

rabbits, muskrat) often eat the plant with low to moderate preference, although there is much variation in the degree of selection (e.g., Atwood 1941, Habeck 1960, Sotala & Kirkpatrick 1973, Wolters et al. 1977, Korschgen et al. 1980, Ludewig & Bowyer 1985, Paul 1993, Castleberry et al. 1999, Pederson and Wallis 2004, Ward 2000, Dillard et al. 2005, Forrester et al. 2006, Thrift 2007, Shaw 2008, Beaver 2011); see also Mohan et al. (2008) and their citations for recent review. *T. radicans* was among the preferred woody species in taste-tests performed with captive American bison in Kentucky (personal trials of the author).

In contrast, the other species of *Toxicodendron* are mostly tall shrubs or trees (*parviflorum*, *potaninii*, *succedaneum*, *sylvestre*, *vernicifluum*, *vernix*, etc.), and they tend to have stronger toxicity (Vogl & Mitchell 1998, Frankel 1999). From literature reviewed here, there are few reports of mammalian herbivory: one case of rabbits on *vernix* in a severe winter (Siegler 1937); one case of deer on *vernix* (Lay 1967); one case of deer on *pubescens*, which is relatively close to *radicans* (Thill & Martin 1989); and an occasional young shoot of *succedanum* by monkeys (Tsuji & Takatsuki 2004).

(5) **Rhamnaceae, section Rhamneae.** *Berchemia* contains just vines, but other temperate genera are small trees or shrubs, sometimes reportedly “scandent” but not high-climbing: *Rhamnus*, *Frangula* and (mostly in drier regions) *Sageretia*. Deeper phylogenetic comparisons may also be appropriate within this complex family (Richardson et al. 2012). The secondary chemistry of *Berchemia* foliage appears largely limited to phenolic compounds, derived glycosides and tannins, plus some non-toxic sterols of medicinal value. But phenyl-ethylamine alkaloids are added in *Sageretia* and *Frangula* (*F. alnus* = *R. frangula*); triterpenes and steroids are added in *Rhamnus alnoides* (Villar et al 1986). Also, the foliage of various *Rhamnus* species, when eaten by mammals, can be strongly laxative—probably due to anthraquinone

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glycosides, or it can depress and potentially damage nervous systems, or it can damage livers (Smyth 1903, Cotton et al. 1933, Lichtensteiger et al. 1997, Mahady 2004, Crowch & Okello 2009). The yellowish inner bark of North American species tends to be ill-scented (E. Wofford, pers. comm.).

In North America, several reports indicate moderate to high preference of deer for *B. scandens* (e.g., Blair 1960, Lay 1967, Wolters et al. 1977, Blair & Brunett 1989, Thill 1984, Nelle 1996, Castleberry et al. 1999, Shaw 2008). Elsewhere, species of *Berchemia* are used locally to feed goats (e.g., Wambui et al. 2011), and shoots are eaten occasionally by monkeys (e.g., Tsuji & Takatsuki 2004). There are no reports of toxic effects in mammals from eating leaves. To compare *Berchemia* with *Frangula*, a meaningful sympatric contrast can be made between *B. scandens* and *F. caroliniana*, which are both well-known species of the southeastern U.S.A. The latter is apparently browsed much less by deer—the few reports indicate generally zero to moderate preference (Lay 1967, Wolters et al. 1977, Lay & Murry 1978, Blair & Brunett 1980, Grabner et al. 2005, Shaw 2008, Wade & Mengak 2010), except in one case that may have included goats (Nelle 1996). The European *Frangula alnus* is often browsed, but typically with just moderate preference (Staines & Welch 1981, Gill 1992, Borkowska & Konopko 1994, Knapp et al. 2008, Aday & Wyckoff 2010).

The common European species, *Rhamnus cathartica*, appears to have little general use by ruminants (Godwin 1943, Heinrich & Predl 1993, Qaderi et al. 2009), except hungry goats (e.g., Decandia et al. 2000). Most species of *Rhamnus* in Mediterranean and Himalayan regions also have generally low but highly variable preference; there is some use by camels and goats, or as fodder prepared for livestock (e.g., LeHou erou 1980, Samant et al. 2007). In eastern North America, foliage of *R. alnoides* is completely avoided by ruminants (e.g., Wright &

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Long 2002) and lagomorphs (e.g., DeVos 1964), and there are no published reports of mammals eating *R. lanceolatus*—although fatal girdling by rodents can occur in arboreta (2 observations of this author).

(6) **Cannabaceae.** The vine *Humulus* has been traditionally used for fodder, salad greens, beer and varied medicinal uses (e.g., Hampton et al. 2009, Al-Mamun et al., 2009, Guo et al. 2009, Srećec et al. 2011). It has diverse chemistry, but is generally not considered toxic. The allied tall annual herb, *Cannabis*, produces some distinct phenolics which have various benign, medicinal or debilitating effects in mammals, especially through the nervous system (e.g., Hall & Degenhardt 2009). There are virtually no reports of *Cannabis* foliage being eaten for food by deer, although fruited material may be rarely used in winter (as cited by Atwood, 1941) and seeds are much used by birds and small mammals. Fresh foliage is generally avoided by livestock, but cattle may rarely consume a toxic amount (Driemeir 1997), and moderate amounts are sometimes processed into silage for livestock (e.g., Fisher 1975).

(7) **Hydrangeaceae.** *Decumaria*, *Pileostegia* (both evergreen) and *Schizophragma* (deciduous) form a clade of vines (Hufford et al. 2001) that appear to have a rather simple, largely phenolic chemistry without significant toxicity (Bohm et al. 1985, Li 2011). The allied deciduous genus *Hydrangea* mostly contains shrubs, and it has more diverse chemistry that includes cyanogenic glycosides and alkaloids in some East Asian species (e.g., Chang et al. 2003, Ishih et al. 2007, Nakamura et al. 2009). There is only sparse chemical information from the few climbers classified within *Hydrangea* (*anomala*, *integrifolia*, *petiolaris*, *integrifolia*, *peruviana*, *seemanii*, *serratifolia*), but it suggests that flavonoids are less developed (Bate-Smith 1978).

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(8) **Bignoniaceae.** The tribe Bignonieae are virtually all climbers (mostly evergreen), and they tend to have low levels of iridoids or less complex forms of them (Von Poser et al. 2000). Iridoids are generally more common in the allied taxonomic groups based on *Tabebuia*, *Tecoma* and *Catalpa*, which are mostly trees (Lohmann 2006, Olmstead et al. 2009). The evergreen vines *Bignonia* (Table 5) and the deciduous vine *Campsis* are both browsed by rabbits or deer at moderate degrees in eastern U.S.A. (e.g., Blair et al. 1977, Wolters et al. 1977, Castleberry et al. 1999, Fargione et al. 2001, Halls & Boyd 1982, Williams & Baxley 2008). In contrast, there are virtually no published reports of deer browsing on the temperate tree, *Catalpa* (Atwood 1941, Heinrich & Predl 1993, Jull 2001), which produces toxic naphthaquinones, iridoids and a phthalide lactone (e.g., McDaniel 1992, Park et al. 2010, Lampert et al. 2011). Further comparison is needed among the tropical genera.

(9) **Caprifoliaceae.** Climbers in *Lonicera* consist of the largely deciduous section *Caprifolium* plus the more evergreen *japonica-macrantha* group. The many species of non-climbers are mostly deciduous shrubs, including the European *xylosteum* and the East Asian invaders in North America: *standishii*, *fragrantissima*, *maackii*, *morrowii* and *tatarica*. Available data (Appendix 4) suggest that species of *Caprifolium* are highly nutritious, and have relatively low levels of phenolics, iridoids and tannins, compared to the shrubs (e.g., LeHouérou 1980, González-Hernández & Silva-Pando 1999, Cabiddu et al. 2000, Nostro et al. 2000, González-Hernández et al. 2003, Peñuelas et al. 2006, Lieurance et al. 2012). Indeed, the old latin taxonomic name—*Caprifolium*—means ‘goat’s leaf’ due to their love of these plants, which have generally been called ‘woodbine’ in England.

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“But chief the flower beyond compare,
the flaunting Woodbine revell’d there,
Sacred to Goats; and bore their name
‘Till Botanists of modern fame
New fangled titles chose to give
To almost all the plants that live.”
(Whitehead 1777)

However, increases in nutrient supply, damage or stress in the climbers can promote iridoids that tend to deter herbivory, especially by insects.

Deer appear to have only moderate preference for the shrubby species of *Lonicera*, in general (Appendix 4). Leaves of *maackii* produce relatively high concentrations of the flavonoids, apigenin and luteolin plus their glycosides (Cipollini et al. 2008), and varying levels of these chemical have been found in several other species of the genus. Apigenin stimulates apoptosis in cells (programmed death), including red blood cells (Zbidah et al. 2012), and it has many physiological effects in animals and plants. Luteolin is less toxic and it has many potential medical uses, but high doses tend to be nauseous. Both chemicals can have oestrogenic effects and disrupt molting cycles (as reviewed by Cipollini et al.). Further comparisons among genera of Caprifoliaceae and Adoxaceae would be useful, since there are significant differences in chemistry and use by ungulates.

(10) **Smilacaceae (*Smilax*)**. Most species are woody or herbaceous climbers, with diverse secondary chemicals but little or no mammalian toxicity (Table 5). They are sister to Liliaceae

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(sensu stricto), which are strictly non-climbing and contain many species with alkaloids or derivatives of methylene-butanoates that have mammalian toxicity (Keeler 1979).

In other families with vines, there are no clear differences in chemistry or herbivory between climbers and allied non-climbers based on available information. However, relevant data remain sparse or absent in several cases.

(1) **Aristolochiaceae**. Appropriate comparisons need more attention, given the complex taxonomy (Ohi-toma et al. 2006). Both *Isotrema* and *Aristolochia* (sensu stricto) contain vines, shrubs and herbs. All are alkaloidal and probably avoided by mammalian herbivores.

(2) **Ranunculaceae**: *Clematis* versus *Anemone* (sensu lato). Both tend to be somewhat toxic due to anemonin or other compounds, often with a sharp-peppery taste. But there are reports of local browsing, especially by deer (Habeck 1980, Fargione et al. 1991, Nelle 1996, González-Hernández & Silva-Pando 1999, Samant et al. 2006, Xiang et al. 2007, Wallach et al. 2009). Much chemistry in *Clematis* involves saponins rather than nitrogenous alkaloids, and despite many medicinal uses severe mammalian toxicity has rarely been reported (Pei et al. 2009, Hao et al. 2012).

(3) **Celastraceae, tribe Celastreae**. Appropriate comparisons need more attention, given the complex phylogeny and chemistry (Simmons et al. 2001).

(4) **Vitaceae**: Vitoideae versus Leeoideae (*Leea*). There is much documented browsing by deer on *Vitis* (except perhaps *rotundifolia*), *Ampelopsis* and *Parthenocissus*, though perhaps less on the latter (e.g., Atwood 1941, Blair & Burnett 1980, Yokoyama et al. 2000, Samant et al. 2007, Shaw 2008, Williams & Baxley 2008, Beaver 2011; see also Borchard et al., 2010). Their tropical climbing relatives include *Tetrastigma*, which is eaten by varied mammals including

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humans (e.g., Sawian et al. 2007; D. Nickrent, pers. comm.), and *Cissus*, which can have sublethal toxicity and varied medicinal effects (as reviewed by Nyahangare et al., 2012). Little available information exists on herbivory of *Leea*, which is more arboreal and strictly tropical; there is a report of human consumption (Barua et al. 2007).

(5) **Rosaceae**: climbers/trailers versus more shrubby species within *Rosa* and perhaps *Rubus*. There is much browsing by deer in both of these complex genera, which are defended largely by tannins and thorns.

(6) **Fabaceae**: Phaseoleae (including *Lackeya* and *Pueraria*) plus Milleteae (including *Wisteria*) versus Indigoferae, or perhaps deeper comparisons to be established within these complex groups. The toxic amino acid canavanine is typical of *Dioclea* and some other largely tropical genera within Phaseoleae, but it is absent from *Pueraria*; the chemistry of *Lackeya* remains unknown (Lackey 1977). *Wisteria* is quite toxic due to other compounds (Table 6).

(7) **Actinidiaceae**: *Actinidia* and *Clematoclethra* are climbers that should be compared to non-climbing *Saurauia*, but the latter is tropical. Within Ericales as a whole, it may be reasonable to just compare other temperate taxa—mostly Ericaceae, which have well-known toxicities in many evergreen species (benzo- and naphthoquinones) but less in deciduous (e.g., *Vaccinium*).

(8) **Gentianales**: Oleaceae (*Jasminium*), Loganiaceae (*Gardneria* etc.), Gelsemiaceae (*Gelsemium*), Apocynaceae (*Trachelospermum*, *Vinca*, etc.). Appropriate comparisons for temperate vines in these families remain unclear, given their complex phylogeny (Livshultz et al. 2007, Frasier 2008). See “Notes on individual taxa” above; although alkaloidal, some of these plants can be browsed much in the winter.

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(9) **Araliaceae**: *Hedera* versus *Acanthopanax*, *Schefflera* and allies. Sublethal toxicity due to varied terpenoids has been reported from these plants; deeper review is needed.

But there is, remarkably, almost no opposite trend among temperate plants—that is, more defensive secondary chemistry in a largely climbing clade compared to the most closely allied non-climbing clade. **Ranunculales** might present the only example: Menispermaceae have a greater diversity of alkaloids and terpenoids than their largely non-climbing relatives in Berberidaceae plus Ranunculaceae (Barboso-Filho & Leitão da-Cunha 2000). However, all three of these families are somewhat toxic to mammalian herbivores, and the chemistry of some temperate Menispermaceae remains poorly known (especially *Calycocarpum*). Moreover, some *Cocculus* and other species are reported to have moderate or high usage by goats and deer (Nelle 1996, Shinde et al. 2000, Bhatta et al. 2001, Samant et al. 2007). It would be interesting to compare general chemistry of vines in Menispermaceae with the few derived shrubby lineages within this family (Ortiz et al. 2007, Jacques et al. 2011).

A deeper and broader analysis will eventually be interesting to pursue, including herbaceous vines as well as woody. Chemical evolution within Solanales is particularly complex, but possible associations with the frequent herbaceous climbing habit in this group do not seem to have been explored (Eich 2008).

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Table 6. Features of secondary chemistry in vine genera that are strictly deciduous within mid-temperate regions.

Genera are listed in phylogenetic order, as in Table 5. Those that contain only vines are underlined. Included here are some largely herbaceous genera with close woody relatives. Several additional genera not listed here have both deciduous and evergreen species (*Actinidia*, *Akebia*, *Celastrus*, *Lonicera*, *Smilax*). Abbreviations are generally the same as for Table 5.

- () : occurrence in mid-temperate zones is marginal, more common in warmer zones
 ? indicates uncertain ecological or chemical assignment
 LF: EG = leaves often evergreen in warmer zones; underlined are often coriaceous
 LF: eg = leaves partially as above, at least somewhat coriaceous
 LF: HE = plants herbaceous in mid-temperate zones but often exceeding 3 m
 LF: he = plants partially as above
 HZ = Hardiness Zones of mid-temperate species (excluding species of warmer zones)
 Predominant types of chemistry are indicated by the following symbols (see also Table 5).
 C = condensed tannins
 N = alkaloids or other nitrogen-containing compounds
 P = phenolics/flavonoids and derived glycosides
 T = terpenoids (with saponins, iridoids, etc.) and derived glycosides
 S = sterols/steroids and derived glycosides
 TOX: indicates genera with reported severe toxicity to mammals.
 TAS: provisional descriptors of taste to humans.
 pr = typical protein contents of leaf dry weight, with references.

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FAMILY: GENUS	LF	HZ	SECONDARY CHEMISTRY
MAGNOLIID FAMILIES			
Schisandraceae: <u><i>Schisandra</i></u> TWI Monoecious	eg?	7-8	PT: polyphenols, ?flavanols, ?tannins/acid? lignans, triterpenoids pr = 9-13% (Nakagawa 1989, Huang+ 2010, ?Kusumoto+ 2012)
Aristolochiaceae: <u><i>Isotrema</i></u> TWI	eg?	6-8	N: unusual alkaloids* (aristolochic acid)/2.38 TOX* ?pr = 11-22% (Bostan+ 2012, Kusumoto+ 2012)
Menispermaceae: <u><i>Calycocarpum</i></u> TWI Dioecious		7-8	?NP: unknown but probably as in <i>Cocculus</i> ?TOX
Menispermaceae: <u><i>Cocculus</i></u> TWI Dioecious	he <u>EG</u>	6-9	NPS: flavonoids, alkaloids of aporphine, erythrina, or isoquinoline* type/3.68, ?sterols, ?tannins/acids ?TOX* ?pr = 11% (Bhatta+ 2001)
Menispermaceae: <u><i>Menispermum</i></u> TWI Dioecious	HE	4-7	NPT: flavonoids, alkaloids (aporphine, chlorinated, isoquinoline* or phenolic types)/3.0, cyanogenic glucosides, terpenoid lactones TOX* TAS: "rank"
RANUNCULID FAMILIES			
Ranunculaceae: <u><i>Clematis</i></u> (s.s.) TWI Partly dioecious	he <u>EG</u>	6-9	PNT: anemonin* (pentadienoic dimer), aporphine alkaloids, ?triterpenoids/saponins/glycosides (roots) TOX* TAS: acrid-peppery pr = 13-19% (Nakagawa 1989, Papageorgiou+ 1981)

ROSID FAMILIES			
Vitaceae: <i>Ampelopsis</i> TEN		6-10	P: phenolics (ampelopsin, myrecetin, gallates, quercetin; lupeol, ethyl gallate, stilbenes), flavanols (catechin) pr = 12% (Iwamoto 1982)
Vitaceae: <i>Parthenocissus</i> ADR +TEN		3-10	P: phenolics (leucantho-cyanidins, stilbenes), tannic acids, flavanols (catechin) pr = 10-16% (Conklin-B+ 1999, Shaw 2008, Lashley 2009, Beaver 2011)
Vitaceae: <i>Vitis</i> TEN Polygamo-dioecious		3-10	P: phenolics (caffeic acid), phenolic acids (gallic acid, stilbene trimers), flavonoids (quercetin), ?flavanoids pr = 10-20% (Torgenson+ 1971, Conklin-B. 1999, Shaw 2008, Lashley 2009, Beaver 2011)
Anacardiaceae: <i>Toxicodendron</i> ADR Polygamo-dioecious		4-10	P: phenolics, flavonoids, tannic acids (gallotannic acid) ALLERGENS: pentadecyl-catechols, biflavonoids pr = 10-16% (Torg.+ 1971, Lash. 2009, Beaver 2011)
Rosaceae: <i>Rosa</i> THO Rarely dioecious (<i>setigera</i> is cryptically dioecious)	eg	4-8	PC: phenolics, flavonoids/glycosides, tannins TAS: mealy-slightly astringent pr = 8-15% (Torgenson+ 1971, Nakagawa 1989, Hedtcke+ 2009, Ammar+ 2004)
Rhamnaceae: (<i>Berchemia</i>) TWI	eg	7-10	P(S): benzoquinones, tetralones*, glucosyl-oxybenzoates, lignans, ?sterols; ?TOX* pr = 12-15% (Shaw 2008; Kusumoto+ 2012; based partly on <i>floribunda</i> , <i>racemosa</i>)
Cannabaceae: <i>Humulus</i> TWI Dioecious	HE	3-7	PTSC: phenolics, tannins, monoterpenoids, triterpenoids (humulone), prenylated flavonoids (steroid-like), sterols, furans ?pr = 17-23% (Al-Mamun+ 2009, Guo+ 2009)

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Fabaceae: <i>Lackeya</i> (segregate of <i>Dioclea</i>) TWI	he	7-8	N: ?canavanine/similar*, ?lectins* ?TOX* ?pr = 7% (Hecht 1979)
Fabaceae: <i>Pueraria</i> TWI	he	6-10	PTN: isoflavones (genistein, and puerarin—an oestrogenic glucoside), triperpenoids, indole alkaloids (roots); pr = 18-23% (Corley+ 1997, Deguchi+ 2001, Samant+ 2007)
Fabaceae: <i>Wisteria</i> TWI		6-9	NC: wisterine* (uncharacterized glycoside), lectins*, tannins TOX* (lectins = proteins that bind to sugars)
ASTERID FAMILIES			
Hydrangeaceae: (<i>Schizophragma</i>) ADR		7-9?	?TP (based on <i>Pileostegia</i> ; Li 2011) ?pr = 8-9% (Nakagawa 1989, Kusumoto+2012)
Hydrangeaceae: <i>H. anomala</i> , <i>integrifolia</i> , <i>kawakamii</i> , <i>petiolaris</i> ADR		6-9?	PT (?NS): flavonoids, iridoids, ?isoquinoline- and ?quinazalone-alkaloids*/3.0, ?cyanogenic glycosides*, (coumarins), ?tannins, ?steroidal glycosides ?TOX* (or perhaps only shrubs) ?pr = 13-18% (Togenson+ 1971, Deguchi+2001)
Actinidiaceae: <i>Clematoclethra</i> TWI? Dioecious		4-7?	PTS: phenolics (caffeic acid, coumarins, mangiferin), flavonoids (kaemferol), tripterpenoids (betulin), sterols
Apocynaceae: (<i>Trachelospermum</i>), <i>Thyrsanthella</i> * TWI (?ADR; Cai et al. 2005)	eg	7-9	PTN: phenolics, flavonoids, lignan glucosides, triterpenoids, saponins, indole alkaloids* ?TOX* pr = 6-8% (Iwam. 1982, Nakag. 1989, Kusum.+ 2012)
Bignoniaceae: <i>Campsis</i> ADR (nodal)		5-9	TP: iridoids, phenolic acids (ferulic), phenyl propanoid glycosides, flavonoids pr = 8-12% (Jones+ 2008)

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PART THREE: General Discussion of Evolutionary Trends and Ecological Factors

Patterns in Phylogeography, Morphology and Physiology

Phylogeographic context. All species of evergreen-tending vines in mid-temperate regions belong to genera with largely temperate ranges, except *Celastrus*. However, several of them, especially those that often reach 10 m (*) and have relatively coriaceous leaves, are in largely tropical families (Stevens 2012): Bignoniaceae (*Bignonia**), Araliaceae (*Hedera**), Actinidiaceae (*Actinidia**) and Celastraceae (*Celastrus**, *Euonymus**). The largely temperate families represented here are Caprifoliaceae (*Lonicera*), Hydrangeaceae (*Decumaria**) and Lardizabalaceae (*Akebia*, *Holboellia**). Smilacaceae (*Smilax*) are a more widespread family of humid tropical and temperate regions—the only family represented here that occurs in all three major realms of the Northern Hemisphere, but largely semi-evergreen in eastern North America and just short, shrubby and southern within Europe. These families represent all major superorders of angiosperms, except the relatively primitive Magnoliidae, which do have several temperate deciduous vines (see Table 5 and text above). They are concentrated among the relatively advanced Asteridae (with *Decumaria*, *Actinidia*, *Hedera*, *Bignonia* and *Lonicera*).

The 11 genera or subgeneric sections with some vines that are evergreen-tending tend to have more restricted global ranges than deciduous or herbaceous taxa (Tables 7, 8a). The six that are strictly evergreen, or nearly so, are as follows: the Sino-American disjunct *Decumaria*—with two species; the Sino-Himalayan *Holboellia*—with ca. 20 species but reasonably merged into the paleotropical *Stauntonia* (Christenhusz 2012); the East Asian *Euonymus* section *Ilicifolia*—with perhaps only 1–4 vining species; the eastern North American *Bignonia*—with maybe just one species but close to the neotropical *Cydista* (Lohmann 2006,

Olmstead et al. 2009); *Lonicera* section *Nintooa*—with ca. 5–10 East Asian species (plus one short, reportedly deciduous species in Mediterranean regions); and the Eurasian *Hedera*—a distinct genus with at least 13 species (Green et al. 2011). Only *Hedera* has radiated widely from its original center in the Mediterranean region, but was absent in North America until recently introduced. The genera/subgenera with mixed evergreen and deciduous condition include two that are restricted to East Asia (*Akebia*, *Actinidia*), two that are moderately widespread but lacking in most of Europe (*Celastrus*, *Smilax*), and one that occurs in all three regions (*Lonicera* subgenus *Caprifolium*).

Among these 11 evergreen-tending taxa, only *Decumaria* has a disjunct distribution in East Asia and eastern North America. Such disjunctions are generally considered to result from fragmentation of the Arcto-Tertiary Flora about 5–10 million years ago (Wen 1999, 2001, Xiang et al. 2000, Milne & Abbott 2002, Donoghue & Smith 2004, Milne 2006). In contrast, among the 23 genera or subgeneric sections with vines that are largely deciduous to herbaceous (Tables 7, 8a), 11 are largely restricted to East Asia and Eastern North America; six have relatively broad distributions across temperate or montane regions; and only five are restricted to one continent. Deeper phylogeographic analysis may reveal similar disjunctions within some largely deciduous to herbaceous sections of more widespread vining genera centered in warmer zones: *Cocculus*, *Berchemia*, *Trachelospermum* (with its segregate *Thyrsanthella*).

Is evergreenness more recent than the deciduous habit? The more restricted ranges of evergreen-tending mid-temperate genera might be attributed, in part, to a generally more recent origin. Synthesis of phylogenetic evidence—calibrated in some cases with fossils—does indicate that the evergreen-tending taxa are less old than the deciduous (Table 10). Such evidence assumes that currently evergreen or deciduous genera have largely remained so since

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they originated. More comprehensive analysis of phylogeography, fossils and functional traits will be needed eventually to provide a rigorous test of this hypothesis. Another consideration is that repeated cold climatic periods during the past 5–10 million years might have prevented evergreenness from becoming a widespread feature among mid-temperate vines. There is indeed some evidence that disjunctions among evergreen Asian-American plants in general tended to develop earlier than in deciduous plants, presumably caused by more sensitivity to cold (Milne & Abbott 2002).

The oldest evergreen-tending mid-temperate vining genus appears to be *Decumaria*, estimated to have diverged during the Oligocene (ca. 20–30 million years ago) from the subtropical to warm temperate genera, evergreen *Pileostegia* and deciduous *Schizophragma* (Samain et al. 2010, Xiang et al. 2011). As reviewed by Green et al. (2011), molecular and fossil evidence indicates that *Hedera* originated in subtropical to warm temperate forest during the Miocene, about 10–15 million years ago, when disjunctions among the older Arcto-Tertiary genera were being formed. Based on the biogeographic context of *Stauntonia* (sensu lato with *Holboellia*) and *Bignonia* (sensu lato), it is likely that their radiation into temperate zones is no older than the origin of *Hedera* (Burnham & Graham 1999, Wang et al. 2002). Within the largely shrubby genus, *Euonymus*, the evergreen vines—*fortunei* and allies—occur only in section *Ilicifolia*, which is restricted to East Asia. The *fortunei* group are closely related to more shrubby species, suggesting a recent divergence, but a molecular analysis of their phylogeny is not yet available.

Genera with both evergreen and deciduous vines in temperate regions will provide special insight to evolutionary processes when their phylogeny is clarified: *Actinidia*, *Akebia*, *Celastrus*, *Hydrangea* (sensu lato), *Lonicera* and *Smilax*. Based on morphological patterns outlined in Flora of China (1996–2011) plus recent molecular analysis, evergreenness of mid-

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temperate species could be a derived character within most of these genera (Chat et al. 2004, Wang et al. 2002, Mu et al. 2012, Smith 2009, Samain et al. 2010, Xiang et al. 2011). Patterns within the complex genus, *Lonicera*, are particularly interesting, since it contains a largely deciduous vining group—subgenus *Caprifolium*—that is widespread across the Northern Hemisphere, plus a semi-evergreen vining group—section *Nintooa*—that has much less geographic radiation within the largely deciduous and shrubby subgenus *Lonicera* (Appendix 4). As indicated by Smith (2009), *Nintooa* probably had a more recent origin than *Caprifolium* (ca. 20 versus 25–30 million years ago). However, most of the radiation within both groups appears to have occurred within the past 10–15 million years. Another complex genus, *Hydrangea* sensu lato—including *Decumaria*, could also provide an instructive comparison of deciduous and evergreen lineages, which seem to have diverged much earlier than in *Lonicera* (Xiang et al. 2011). The deciduous climbing habit may be ancestral within the Arcto-Tertiary “*Hydrangea* 1” clade of Samain et al. (2010). In *Smilax*, some morphological patterns suggest that evergreenness is derived among the largely temperate clades (Chen et al. 2006), but molecular analyses suggest that it is ancestral within the whole genus (Cameron & Fu 2006).

Despite their smaller ranges and evidence of more recent appearance, the evergreen-tending vines tend to have more species per genus and more current invasiveness than the deciduous vines (Table 8a). These are weak trends, but they affirm that evergreenness has special benefits for these plants in some modern contexts.

Climbing habits. There are diverse climbing patterns in vining plants (Putz & Mooney 1991, Isnard & Silk 2009). The evergreen-tending species of mid-temperate regions—as well as other temperate vines—are loosely divisible into two broadly defined ecological groups, although a

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few species combine features of both. At the generic level, this division has associations with several biological features (e.g., Table 9, Figure 6).

(1) Typical twiners, scramblers and tendrillers. These climb on supporting branches but not vertical surfaces, often to no more than 5–10 m high, without adventitious roots but usually twining around the supports: *Akebia*, *Holboellia*, *Celastrus*, *Lonicera*, *Smilax* and perhaps some *Actinidia*. Although not able to climb up larger trees by themselves, such species—especially Vitaceae among deciduous taxa—can sometimes climb up other vines (Putz 1995) or up smaller trees and shrubs, then move to larger hosts (Ichihashi & Tateno 2011). *Smilax* uses tendrils and thorns to climb, but its shoots do not twine. *Holboellia* is the only strictly evergreen genus in this group. In contrast, the 23 genera or subgeneric sections with strictly deciduous to herbaceous vines in mid-temperate regions (Table 7) mostly belong here, except *Parthenocissus*, *Toxicodendron* (*radicans* group), *Schizophragma*, *Hydrangea* (*anomala* group) and *Campsis*, which belong with the following.

(2) Surface-ascenders. These are able to ascend tree trunks, cliffs or walls, often up to 10–20 m and usually with adventitious roots, at least on lower stem sections: *Decumaria* (both species), *Hedera* (all species), *Euonymus fortunei* and allies, *Actinidia henryi* and *Bignonia capreolata*. Assignment of *A. henryi* remains uncertain since the extent of adventitious roots is not published, but such roots are known in the genus and *A. kolomicta* is reported to climb trees “by means of adventitious roots in the lower parts, or by the twining of long whip-like shoots higher up, or rambling after the fashion of brambles over the undergrowth, or in clearings trailing on the ground by means of adventitious roots in the lower parts” (Stapf 1926). In the case of *Bignonia*, climbing is accomplished by versatile tendrils that are able to curl around narrow stems or to attach in small crevices of most tree barks. Its tendrils “form into irregularly

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shaped attachment pads when they come into contact with a suitable substrate surface. These attachment pads grew around presented fibers and into surface cavities and excreted a resin-like substance” (Seidelmann et al. 2012, summarizing Charles Darwin’s observations of 1875). But only larger lower stem sections of *Bignonia* eventually form adventitious roots.

There has been insufficient synthesis of research on growth rates and shade tolerances to allow thorough physiological comparisons of these two groups, but several studies have provided useful insight (e.g., Forseth & Teramura 1987, Carter et al. 1988, 1989, Schierenbeck & Marshall 1993, Baars & Kelly 1996, Cai 1999, Sakai et al. 2002, Cai et al. 2005, Jiang et al. 2007, Allen 2007, Ichihashi et al. 2009, 2010, 2011, Leicht-Young 2010, Kusumoto et al. 2012). Typical twiners and scramblers appear to have more rapid potential growth and less shade tolerance, on average, often resulting in lower leaf:stem mass ratios and establishment on smaller hosts. *Akebia* and *Lonicera japonica* are somewhat exceptional in having a moderate degree of shade tolerance, and often sending prostrate shoots into understories. But they are still concentrated in young or thin woods with more small trees and shrubs—for example, on the Carolina Piedmont, *L. japonica* has declined in maturing woods where *Vitis rotundifolia* and *Rubus* spp. have increased (Taverna et al. 2005). Also, the thick rhizomes of *Smilax* often maintain low shoots that are thinly scattered in the shade, exposed to herbivory but able to grow up if fresh canopy gaps occur. These shoots are sometimes more thorny or variegated than taller flowering shoots.

In contrast, the ‘surface-ascenders’—especially *Hedera*, *Euonymus* (evergreen) *Toxicodendron* and *Parthenocissus* (deciduous)—can often produce leafy shoots that dominate much ground in shade between trees. In *Hedera* and *Euonymus* the leaves of low creeping ‘juvenile’ shoots often have distinct coloration, morphology, physiology, compared to those of

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root-climbing shoots and flowering ‘adult’ shoots (Metcalf 2005, Bauer & Bauer 2006, Yang et al. 2006). Also, *Decumaria*, *Schizophragma* and *Toxicodendron* are able to produce much thinner and broader leaves in shade. However, *Campsis* is relatively intolerant of shade and among the most rapidly growing species—it is the only ‘surface-ascender’ with pinnate leaves.

To summarize, this initial review of the literature indicates that most of the 11 evergreen-tending genera or subgenera (Table 7) are ‘surface-ascenders’ with simple leaves (or paired in *Bignonia*), moderate to high shade tolerance, relatively low specific leaf area (mm^2/mg dry weight), and generally moderate growth rates. *Holboellia* and *Akebia* (which is partially deciduous) are exceptional in their more twining habit and palmate leaves. In contrast, the 23 largely deciduous to herbaceous taxa are mostly twiners, scramblers or tendrillers (except for five genera), with compound leaves in 10 cases. Deciduous vines appear to have a broader distribution of overall stature, leaf forms, nitrogen contents, shade tolerances and growth rates. Among twiners or tendrillers, *Ampelopsis*, *Berchemia*, most *Vitis* (except *rotundifolia*), most *Actinidia*, most *Celastrus* and most *Lonicera* subgenus *Caprifolium* generally do not grow into more shady understories. In contrast, *Parthenocissus*, *Schizophragma* and *Toxicodendron* are ‘ascenders’ that also have a pronounced ability to grow over the ground under forest canopies—often exhibiting unusually high and variable specific leaf area (e.g., Ichihashi et al. 2009). Moreover, deciduous species with compound leaves have some of the fastest growth rates, in terms of annual extension by both ‘ordinary shoots’ and longer ‘searcher shoots’ in twiners (e.g., *Akebia*, *Pueraria*, *Wisteria*) and in the somewhat anomalous ‘ascender’, *Campsis*.

Hydrological associations. Figure 7 summarizes, for forests of east-central U.S.A., the apparent distribution of all native woody or subshrubby vine species in terms of their typical modal positions along hydrological gradients. The format follows that of Figure 1a, with

abbreviations for vines rather than trees (Appendix 1), but their positions are based on general knowledge and review of the literature rather than data from plots. Although most species have wide overlapping ranges of habitat, there is a concentration of modal positions along the zone between two major sectors: (1) mesic-to-subxeric-to-xeric woods; versus (2) wetlands and open shrubby-to-grassy uplands. Such concentration is confirmed by detailed observations at specific sites (e.g., Waters et al. 1974, Collins & Wein 1993, Goebel et al. 2001, Allen 2007, p. 105). It is suggested that the core habitat for woody vines, in general, has been developed within this broad ‘vining zone’—where deeper woods typical of more hilly terrain come into contact with riparian interruptions, wetlands, browsed or burned uplands and plains.

Figure 7 marks (with underlining) those species that have evergreen-tendency, including some that are largely deciduous within Kentucky but evergreen-tending further south. The few native ‘ascenders’ (group 2)—including evergreen-tending *Decumaria* and *Bignonia*—are mostly positioned in slightly more mesic wooded habitats. The other evergreen-tending species are mostly clustered at the drier end of the vining zone: *Smilax* spp., *Lonicera sempervirens* and *Cocculus*.

Physiology of water-relations. There is a need for deeper comparative review than is possible here. One basic theme would be the extent to which xylem anatomy is correlated with the twining versus trunk-climbing habit, or with deciduous versus evergreen habit. An initial reading of scattered incomplete literature suggests that several of the ‘twining and scramblers’ (group 1) have ‘ring-porous’ clustering of particularly wide vessels in early wood, which can allow rapid growth to be concentrated in a delayed spring to early summer, before onset of droughts—a trait also found in most temperate trees of summer-dry sites (Woodcock 1994, Zanne 2006, Boura & DeFranceschi 2007). Ring-porous vines include *Akebia trifoliata* (Sun et al. 2003), *Clematis* spp. (Schoch et al. 2004), *Celastrus* spp. (Davis & Evert 1970, Tibbets &

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Ewers 2000), *Vitis* spp. (Schoch et al. 2004), *Rosa* spp. (Schoch et al. 2004), *Wisteria floribunda* (Sun et al. 2003), some *Actinidia* spp. (Condon 1991, Xiao et al. 2010), and *Lonicera japonica* (Bell et al. 1988, Chiu & Ewers 1992). *Vitis* maintains unusually wide and late-produced vessels throughout most of its wood, together with strong root-pressure until frost and active phloem into the winter (Tibbets & Ewers 2000).

In contrast, distinctly ring-porous patterns (versus just ‘semi-ring-porous’)—or presence of unusually wide vessels throughout—are not documented among ‘surface-ascenders’ like *Parthenocissus* and *Hedera* (group 2). Further research is needed like that of Bell et al. (1988), who found: “Most significant among the species differences in water relations were the conservative water use patterns of *P. quinquefolia*, and [in contrast] the midday maxima of transpirational water loss measured in *L. japonica* compared to the morning peaks in transpiration for the two deciduous species.” Slower growth, more stomatal control and diffuse-porous wood in the ‘surface-ascenders’ could also reduce problems from cavitation of xylem during droughts or freezes.

There is, however, no evidence so far of general differences in wood anatomy between the evergreen and deciduous vines. In oaks as well, Cavender-Bares & Holbrook (2001) showed no clearcut relationship between hydraulic properties and evergreenness. And additional traits that may be largely independent of evergreenness are the structure, extent and function of rhizomes or other underground storage organs, as in the monocot *Smilax* (without cambial growth). Cobb et al. (2007) concluded that: “Strong root pressure can account for *Smilax*’s survival in temperate regions with severe frosts, where few monocots with persistent aboveground organs are found.” The rhizomes of *Smilax* must also be involved in the remarkable ability of these plants to recover from repeated damage (e.g., Boggs et al. 2012).

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Leaf-longevity in general and its temperate associations. Evergreenness itself deserves more precise definition in most of these vines, ideally based on measurements of longevity in individual leaves (Koyama & Kikuzawa 2008). Most of the evergreen-tending species in mid-temperate zones appear to have leaves that live up to a year, until shortly before or shortly after the next flush of growth in spring—these would be “brevi-deciduous” to “semi-evergreen” in the sense of Kikuzawa & Lechowicz (2011, Chapter One). *Hedera helix*, which has relatively tough frost-resistant leaves, tends to lose them after spring—then providing a pulse of nutrients from their decomposition (Badre et al. 1998). But even in this well-known species, longevity of leaves is not well-documented. Fischer & Feller (1994) worked with *Hedera* leaves up to two years old, which may be expected as a typical mean for broad-leaved evergreen woody plants in humid temperate zones. Yet there is much variation even within species (Escudero & Mediavilla 2003, Hikosaka 2005, Wright et al. 2005).

Within humid temperate regions, the proportion of evergreen woody plants is greatest on relatively acid, infertile soils (e.g., Monk 1966, Reich et al. 1992, Givnish 2002, Ordoñez et al. 2008; Table 1). It has been suggested that this trend simply results from selection of more conservative nutrient-cycling strategies on less fertile soils, where annual shedding of leaves during autumn might allow leaching of critically limiting nutrients during winter or spring. Slower decomposition of evergreen leaves could also be involved (Cornelissen et al. 1999). But a more profound cause may be that the generally slower maximum rates of photosynthesis on infertile soils must directly reduce the potential for summer-growth to offset any lack of leaves during winter.

Moreover, there is a correlated set of foliar traits among all vascular plants (Wright et al. 2004), including temperate vines (e.g., Ichihashi et al. 2009). These traits are greater longevity,

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more thickness (or mass/area), lower N and P contents, and slower maximum photosynthesis (even per area)—but not leaf size. The more long-lived, slower metabolizing leaves in this global ‘spectrum’ are associated with dry climates and infertile soils. Yet much mystery remains concerning the local variation along this continuum, within particular environments or functional groups (Grubb 2002), and there are further relationships with leaf shape. Among woody plants such as temperate vines, evergreen leaves are more often simple versus compound (with the notable exception of *Holboellia*), unlobed (with the notable exception of ‘juvenile’ *Hedera*) and entire-margined, compared to deciduous taxa (Table 9). And although cordate leaf bases are generally associated with the vining habit (Givnish & Vermeij 1976), none of the strictly evergreen mid-temperate vines are distinctly cordate (0/6 versus 11/28 in Table 7); ‘juvenile’ *Hedera*, again, is somewhat exceptional. Possible causes of such associations remain largely untested, although increase in perimeter/area of leaves may enhance photosynthetic rates (with more gas-exchange, cooling and vascular supply), and it may reduce some effects of herbivory (Givnish 1987, Brown & Lawton 1991, Nicotra et al. 2011).

Correlations with leaf nitrogen content. Provisional information on nitrogen contents in mature leaves of several vines are scattered in the literature. These are generally reported as “crude protein” (= $6.25 \times$ N content from the Kjeldahl method), as reviewed by Conklin-Brittain et al. (1999). Among the evergreen-tending vines considered here (Table 5), most have moderate protein contents of about 9–13% dry weight. Leaves of *Hedera* (and perhaps also *Decumaria* and *Holboellia*) have consistently low content (about 9%), in accord with their thickness and slow photosynthetic rates (Carter & Tamura 1988). *Smilax* tends to have relatively high content but quite variable (9–17%), increasing in fresh growth after disturbance (e.g., Dewitt & Derby 1955; and refs. of Table 5). The largely deciduous genera of mid-temperate zones (Table 6), tend to have higher protein contents (mostly 10–18%), but there is

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much overlap with the evergreen-tending taxa. The highest contents (18–22%) have been reported among more herbaceous-to-subshrubby species in *Aristolochia*, *Clematis* and *Pueraria*—which have much nitrogen-based secondary chemistry and rapid weedy growth. However, relatively low contents (6–9%) have been reported from other herbaceous-to-subshrubby genera, especially some with more subtropical or evergreen ancestry (?*Dioclea*, *Schizophragma*, *Trachelospermum*).

Kusomoto et al. (2012) have recently reported nitrogen contents and other functional characters for 20 largely evergreen vines in a subtropical forest of southern Japan, including several species in the same genera as temperate vines. Their data support the concept that thin, nitrogen-rich leaves are loosely associated with a deciduous or herbaceous tendency, and with the twining or scrambling habit rather than climbing with adventitious roots (Figure 6a). Leaf nitrogen content and the deciduous/herbaceous tendency—but not specific leaf area (mm² per mg)—were correlated with concentration of the species on more concave topography (Figure 6b). There were no significant relationships of functional characters with tree basal area or density in the forest. Wood density of the vines was not correlated with the foliar characters, but it was relatively high among the few species of thorny scramblers and nitrogen-fixers.

More collection and analysis of functional data such as these would allow much better understanding of the diversity in vining behaviors. It will be important to add data for expanding displays like Figure 6. A provisional expansion (not shown) using information from Ichihashi et al. (2009, 2010, 2011), Han et al. (2010) and other miscellaneous sources confirms that variation in both leaf nitrogen contents and in specific leaf area tends to increase at higher levels of those parameters. The variation suggests that species are clustered along two branches: towards high N at moderate SLA (with more herbaceous species at the extreme); and

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towards higher SLA at moderate-low N (with deciduous ‘ascender’ *Schizophragma* at the extreme). Thorny scramblers and nitrogen-fixers cluster with the high N branch, but might be divisible. Overlays of secondary chemistry on such figures would be quite interesting.

Concentration of evergreen-tending vines on eutrophic soils. Within humid temperate forests of the Northern Hemisphere, vines are generally most frequent on moist to damp, fertile soils (e.g., Tables 1, 2; Schnitzer & Bongers 2002, Morrissey et al. 2009, Chettri et al. 2010, Kusomoto 2012). The few species of evergreen-tending vine follow this trend. In eastern North America (Table 2, Figure 2), *Bignonia capreolata* and other such species are mostly concentrated on mesic to submesic sites with moderately high pH (ca. 5–7), where the highest overall fertilities tend to occur. In most of Europe, *Hedera helix* (sensu lato) is the only native evergreen vine, and it is most common on nutrient-rich sites with pH of 4.5–7 (Rackham 2003, Metcalfe 2005, Schnitzler & Heuzé 2006). In the Himalayan region, *H. nepalensis* is concentrated on similar soils—together with *Holboellia latifolia* and *Lonicera glabrata* in eastern sections (e.g., Saima et al. 2009, Chettri et al. 2010). Further east in Asia, there is much less published information in English. *Euonymus fortunei*, *E. vagans* and their allies are sometimes associated with base-rich soils, but their associated forest may often be largely evergreen in montane cloud forest with much leaching of minerals from upper soil horizons (Shi & Zhu 2009, Yuan et al. 2009).

This association of evergreen vines with fertile soils is the opposite of trends in other evergreen vascular plants (e.g., Tables 1, 2), and fundamentally different selective factors must have operated. It is likely that the vines are taking advantage of the deciduous season in the tree canopy, continuing to photosynthesize through some of the winter, at least during early or late periods, as shown for *Hedera* (Bauer & Kofler 1987). *Hedera* even flowers and fruits during

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the late summer to fall, an unusual trait among woody plants of mid-temperate zones (also in some *Hamamelis*, *Lonicera*, ?*Viburnum* and Viscaceae). Among evergreen-tending vines of east-central U.S.A., *Smilax glauca* is somewhat exceptional as it occurs mostly on subxeric sites with moderately to strongly acid soils (Figure 2j). But its associated trees are mostly deciduous oaks that form a broad ecological wedge between *Tsuga*-with-*Rhododendron* on more mesic sites and *Pinus*-with-*Kalmia* on more xeric sites.

As noted above, evergreen-tending vines probably spread into temperate zones of the Northern Hemisphere after deciduous trees had largely replaced evergreen trees—including some conifers—during the climatic cooling and drying of mid- to late-Tertiary Eras (Table 10; Axelrod 1966, 1983; Davis 1983, Wolfe 1987, Graham 1999, Manos & Stanford 2001, Manos et al. 2007, Wallander 2008). These vines appeared later than most of the strictly deciduous vines. Moreover, the dominant deciduous tree genera of subxeric sites—*Castanea*, *Quercus*, *Carya*, *Fraxinus*—appear to have proliferated much later than radiation of the original Arcto-Tertiary flora. In the temperate forests of the Southern Hemisphere, one would then expect less shade-tolerant evergreen vines, due to general lack of deciduous trees—this can be tested.

The concept that some life-forms of plants settle on a contrarian strategy to complement the ecology of their dominants, is of course an old idea. Similar selection must have shaped the concentrations of winter annuals and biennials (with overwintering rosettes) in forest on base-rich soils (Table 2). It is also implicated in the much-researched ephemeral flushes of diverse life-forms among vernal perennials, which capture light before new tree leaves expand to cast deep shade, and which capture nutrients from decomposing deciduous litter (e.g., Muller 1978, Grubb & Marks 1989). However, the evolution of evergreen-tending vines and their allies seems to have been generally more recent and more plastic, leaving many questions.

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Evolutionary ‘loose ends’ may include the few odd evergreen-tending ‘subshrubby’ life-forms that can cover ground in deciduous woods—as in *Euonymus* of eastern North America (*obovatus* and, sometimes, *americanus*), and especially the whole genus *Vinca* of Mediterranean regions. Are these constrained evolutionary branches that might have developed into evergreen vines if climates were less harsh during the late Tertiary and Quaternary Eras? Or do these plants still have potential for selection of more rampant forms, as occur in other branches of their families—Celastraceae and Apocynaceae?

Potential interactions with herbivory. Varied direct and indirect effects of herbivory may complicate patterns in evergreen versus deciduous plants, reducing the value of simple theories (Grubb 1992). Even recent global analysis of multiple factors in such patterns leaves much residual variation (VanOmmen-Kloeke et al. 2012). Consumption of evergreen leaves by overwintering herbivores is a problem for many plants (e.g., Karban 2008). Among evergreen woody vines, species like *Euonymus fortunei*, *Bignonia capreolata* and *Hedera helix* can become especially important for mammalian herbivores during the winter (Table 5). Yet browsing on some relatively deciduous *Smilax* species is often most intense during the early growing season, when thick new rapidly growing shoots emerge from the ground—somewhat like bamboos (e.g., Halls 1975).

Uniform addition of herbivory to the landscape might select for more physically- or chemically-defended leaves of the ‘neutral’ type (unspecialized fibre, tannins etc.)—an investment that could lead to mutual benefits with nutritional or photosynthetic strategies for more expensive evergreen leaves on infertile soils. But herbivores—especially larger animals—are expected to concentrate on more productive soils with mineral-rich forage during the winter (Kirby 2001, Jones et al. 2008). Such soils would include valleys, gullies, swales and saddles in

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montane bamboo forests with diverse vines (Igota et al. 2004, Tang 2006, Shi et al. 2012). The direct selective effects of such herbivory could amplify any general trends in nutrient-cycling and photosynthetic strategy that lead to evergreen woody plants being concentrated on infertile soils. Alternatively, young vining shoots on eutrophic sites—more than other woody plants—may often escape from most mammals by growing rapidly into the tree canopy. In that case, there could be special advantages to minimizing structural support and chemical defense, at least temporarily, so that upward growth rate is increased.

For both reasons, defenses of the ‘neutral’ type might be less useful on more fertile soils. It is notable that decreases of foliar phenolic concentrations have been indicated within several woody species along gradients of increasing soil fertility in temperate regions (e.g. Muller et al. 1987, Nicolai 1988). Also, a recent intensive analysis of rain forest in Peru has shown generally lower levels of chemical defenses on more fertile soil, and levels are especially low among woody vines (Asner & Martin 2011).

Patterns in secondary chemistry. As detailed above (at end of Part Two), vining taxa tend to have less defensive chemistry than their closest non-vining relatives. In 10 of these phylogenetic comparisons, there are indications of this trend; in 9 cases there is neutral or uncertain information; and only one case provides a possibly opposing trend. Moreover, shorter woody plants in general may be better defended against mammals than high-climbing vines and trees (Borchard et al. 2011). In particular, most of the species of evergreen shrubs or small trees in largely deciduous forests of north temperate zones have strongly deterrent to toxic chemistry that includes complex terpenoids, steroidal alkaloids or cyanogenic glycosides (Grubb 1992)—*Ilex* species may be exceptional, lacking toxicity, but some of them have prickly leaves instead.

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Most vines still have some chemical defense, and Table 7 summarizes the predominant types of secondary chemicals produced by different groups of vines in mid-temperate regions. Largely herbaceous or subshrubby genera have diverse chemistry that includes alkaloids or other N-containing compounds in most cases (7 of 9 taxa). Largely deciduous woody genera have less diverse chemistry, with relatively simple phenolics predominating (minus condensed tannins or steroidal compounds in almost all cases) and with less occurrence of N-containing compounds (3 of 14 taxa). In genera that have mixed deciduous-evergreen or largely evergreen habit, there is more diverse phenolic chemistry that includes condensed tannins, terpenoids and steroidal compounds in several cases. But again there are relatively few of these genera with N-containing compounds (3 of 11 taxa; $P = 0.02$ with χ^2 test), none of which are acutely toxic to mammals (see notes on herbivory and chemistry above).

Indications of moderate to acute toxicity for mammalian herbivores (Table 7) are concentrated among the largely herbaceous vining genera (6 of 9), plus a few in the largely deciduous woody group (6 of 14). There may be no indications of acute toxicity among the mixed or largely evergreen groups, although a few of these genera are known to cause digestive problems if eaten in quantity (2 of 11; $P = 0.09$ with χ^2 test). Miscellaneous reports of consumption by mammals, especially ruminants, suggest a trend of increasing use from herbaceous to deciduous to evergreen vines (Table 7). These trends remain just suggestive without a more definitive and comprehensive analysis. There is a growing abundance of information on secondary chemicals in vascular plants and their miscellaneous effects. Clearly more synthesis and experimentation could provide answers to the central questions here—which of these chemicals influence consumption by generalist mammalian herbivores, and can the general edibility of many vines in temperate regions be directly attributed to low levels of defensive compounds?

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Table 7. Summary of genera (or subgeneric segregates) with woody or subshrubby vines in mid-temperate zones: herbaceous or deciduous versus evergreen-tending groups.

This relates (with abbreviations) to Tables 5, 6, 8 and 9, plus further literature review for TOX (toxicity) and EAT (edibility) ratings. Growth habit and species numbers are based just on vines in mid-temperate zones. Underlining shows taxa that contain only vines; EG are more evergreen in warmer zones; taxa with minor extent into mid-temperate zones are in ().

HERBACEOUS TO SUBSHRUBBY	SPP	VEG	SEX	RAN	CHE	TOX	EAT
<i>Aristolochia</i> [without <i>Isotrema</i>] (EG) 2n = 6-16, 24, 28, 36 Birthwort	2-4	TWI	Bisexual	Wide+ (AfEu)	N	XX	-?
<i>Cocculus</i> (EG) 2n = 26, 52, 78 Southern Moonseed	2	TWI	Dioec.	Wide (Af)	NPS	X?	+?
<i>Menispermum</i> 2n = 52 Northern Moonseed	2	TWI	Dioec.	AsAm	NPT	XX?	-
<i>Clematis</i> subgenus <i>Clematis</i> (EG) 2n = 16, 32, 48 Clematis	c. 50- 80	TEN	Partly Dioec.	Wide+ (AfEu)	PNT*	X	+?
<i>Humulus</i> 2n = 16-20 (40) Hop	3	TWI	Dioec.	Wide+ (Eu)	PTSC		++
<i>Lackeya</i> [= <i>Dioclea multiflora</i>] 2n = ? <22 in allied genera> Clusterpea	1	TWI	Bisexual	Am	N	X?	+++?
<i>Pueraria</i> ~ 2n = 22-24 (44) Kudzu	5*	TWI	Bisexual	As++	PTN*		+++
(<i>Trachelospermum</i> + <i>Thyrsanthella</i>) (EG) 2n = 20 Star-jasmine	2	TWI	Bisexual	AsAm	PTN*	X?	+?
<i>Smilax</i> subgenus <i>Nemexia</i> 2n = 26 Carrion-flower	5-7*	TEN	Dioec.	AsAm	PTS		+?

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WOODY, LARGELY DECIDUOUS	SPP	VEG	SEX	RAN	CHE	TOX	EAT
(<i>Schisandra</i>) 2n = 28 Magnolia Vine	c. 9	TWI	Monoec.	AsAm	PT		+?
<i>Isotrema</i> [<i>Aristolochia</i> segregate] (EG) 2n = 28-32 Pipe Vine	c. 10	TWI	Bisexual	AsAm	N	XX	-?
<i>Calycocarpum</i> 2n = ? <26 allied genera> Cupseed Vine	1	TWI	Dioec.	Am	?NP	X?	-?
<i>Ampelopsis</i> 2n = 20, 40 Pepper Vine	c. 6*	TEN	Bisexual	AsAm++	P		++
<i>Parthenocissus</i> 2n = 40 Virginia Creeper	3	ADH	Bisexual	AsAm	P		+++?
<i>Vitis</i> [<i>Muscadinia</i> could be split] 2n = 38, 40* Grape Vine	c. 16	TEN	Poly- Dioec.	Wide+ (AfEu)	P		++
<i>Toxicodendron</i> 2n = 30 Poison-ivy	2	ADV	Poly- Dioec.	AsAm	P	X	+++?
<i>Rosa</i> (EG) 2n = 14, 21, 28, 35 (42) Rose	c. 2-5*	THO	Rare Dioec.	Wide++ (AfEu)	PC		++
(<i>Berchemia</i>) (EG) 2n = ? <24 in allied genera> Supplejack	c. 9	TWI	Bisexual	Wide (Af)	P	X?	++
<i>Wisteria</i> 2n = 16 (32) Wisteria	c. 3-6*	TWI	Bisexual	AsAm++	NC	XX	-
(<i>Schizophragma</i>) 2n = 28 Lesser Climbing Hydrangea	1-3	ADV	Bisexual	As	?TP		+?
<i>Hydrangea: anomala</i> group 2n = 36 Climbing Hydrangea	2-4	ADV	Bisexual	AsAm?	PT	X?	+?
<i>Clematoclethra</i> 2n = 48 Lesser Chinese Gooseberry	1-4	TWI?	Dioec.	As	PT		??

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<i>Campsis</i> 2n = 40 Trumpet Creeper	2	ADV	Bisexual	AsAm	TP		++
WOODY WITH MIXED DECID. AND EG	SPP	VEG	SEX	RAN	CHE	TOX	EAT
<i>Akebia</i> (EG) 2n = 16, 32 Chocolate Vine	3-5*	TWI	Monoec.	As+	T		+?
<i>Actinidia</i> (EG) 2n = 58 (116, 174) Chinese Gooseberry	10-15	TWI- ADV	Poly- Dioec.	As+	TPCN*		+?
<i>Celastrus</i> (EG) 2n = 46 Bittersweet	8-12	TWI	Partly Dioec.	Wide++ (Af)	TPNS*	X?	+?
<i>Lonicera</i> subgenus <i>Caprifolium</i> (EG) 2n = 18, 36 Woodbine Honeysuckle	c. 8	(TWI)	Bisexual	Wide+ (Eu)	P(T)		+++
<i>Smilax</i> subgenus <i>Smilax</i> (EG) 2n = 32 (64) Greenbriar	c. 25- 35	TEN	Dioec.	Wide (Af)	TPCS		+++
WOODY, LARGELY EVERGREEN	SPP	VEG	SEX	RAN	CHE	TOX	EAT
<i>Holboellia</i> 2n = 32 China Blue Vine	c. 5*	TWI	Monoec.	As	?TP		+?
<i>Decumaria</i> 2n = 28 Woodvamp	2	ADV	Bisexual	AsAm	?PT		+?
<i>Euonymus</i> section <i>Ilicifolia</i> 2n = 32, ?64 Winter Creeper	1-8*	ADV	Bisexual	As++	TPNS*		++
<i>Hedera</i> 2n = 24, 48, 72, 96, 144, 192 Ivy	5*	ADV	Bisexual	EuAs++	TP	X	++
<i>Bignonia</i> 2n = 40 Crossvine	1	TEN- ADH	Bisexual	Am	?TPS		++
<i>Lonicera</i> section <i>Nintooa</i> 2n = 18 (36) Asian Honeysuckle	5-11	TWI	Bisexual	As++	PT		++

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Table 8a. Comparison of range size, species-richness and invasive tendency between herbaceous-tending, deciduous and evergreen-tending genera of mid-temperate vines.

See Table 7 for list, classes of genera and abbreviated groups. For B, numbers of species (SPP) are estimates; the intermediate class (4–6) is marked with asterisks (*). For C, “some” invasive tendency is “+” under RAN in Table 7; in these cases, the genus is established outside the native range but not widely abundant. Genera with “much” invasive tendency are “++” under RAN; these have become locally abundant in at least one region outside the native range. P is from Fisher’s exact tests with 2×2 cells (a,b,c,d) or extended to all 3×3 cells (Ghent 1972).

A. RANGE WIDTH P= 0.24 (3×3); 0.06 (2×2)	Native to one temperate region	Native to two temperate regions	More widespread (in Africa/Europe)
Herbaceous or subshrubby	2a	3b	4b
Largely deciduous	3a	8b	3b
Largely evergreen or mixed	6c	2d	3d
B. SPECIES-RICHNESS P= 0.27 (3×3); 0.12 (2×2)	1–3 mid-temperate vining species	4–6 mid-temperate vining species	>6 mid-temperate vining species
Herbaceous or subshrubby	6a	2a	1b
Largely deciduous	7a	3a	4b
Largely evergreen or mixed	2c	4c	5d
C. INVASIVE TENDENCY P= 0.31 (3×3); 0.17 (2×2)	No invasive tendency	Some invasive tendency	Much invasive tendency
Herbaceous or subshrubby	5a	3a	1b
Largely deciduous	10a	1a	3b
Largely evergreen or mixed	4c	3c	4d

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Table 8b. Comparison of range size, species-richness and invasive tendency between bisexual/monoecious, partly dioecious & strictly dioecious genera of mid-temperate vines. See Table 7 for list, classes of genera and abbreviated groups. For B, numbers of species (SPP) are estimates; the intermediate class (4–6) is marked with asterisks (*). For C, “some” invasive tendency is “+” under RAN in Table 7; in these cases, the genus is established outside the native range but not widely abundant. Genera with “much” invasive tendency are “++” under RAN; these have become locally abundant in at least one region outside the native range. P is from Fisher’s exact tests with 2×2 cells (a,b,c,d) or extended to all 3×3 cells (Ghent 1972).

A. RANGE WIDTH P= 0.03 (3×3); 0.008 (2×2)	Native to one temperate region	Native to two temperate regions	More widespread (in Africa/Europe)
Bisexual or monoecious	7a	10a	3b
Partly dioecious (various)	1c	0c	5d
Strictly dioecious	2c	2c	3d
B. SPECIES-RICHNESS P= 0.015 (3×3); 0.013 (2×2)	1–3 mid-temperate vining species	4–6 mid-temperate vining species	>6 mid-temperate vining species
Bisexual or monoecious	8a	7c	5c
Partly dioecious (various)	0a	2c	4c
Strictly dioecious	6c	0d	1d
C. INVASIVE TENDENCY P= 0.009 (3×3); 0.09 (2×2)	No invasive tendency	Some invasive tendency	Much invasive tendency
Bisexual or monoecious	11a	3b	6b
Partly dioecious (various)	0a	4b	2b
Strictly dioecious	6c	1d	0d

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Table 9. Tests of associations among some biological features of mid-temperate vines.

See Table 7 for listing and classification of genera, plus abbreviations. P is from Fisher’s exact tests with 2×2 cells (a,b,c,d) or extended to all 3×3 cells (Ghent 1972). Note also that the ‘evergreen-tending’ condition is weakly associated with the ‘surface-ascending’ condition (P = 0.1 in 2×2 test among the 24 woody genera).

1-5 [next page]. Comparisons between genera of herbaceous-subshrubby vines; versus woody twiners, tendrillers or scramblers; versus woody surface-ascenders. Ascenders have ADR or ADH under VEG in Table 7.

6-10 [second page]. Comparisons between genera of herbaceous-subshrubby vines; versus largely deciduous; versus evergreen-tending genera. Note that in 10 the strictly evergreen group are all strictly non-dioecious (see text).

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1. SERRATED LEAVES P= 0.94 (3×3); 0.32 (2×2)	Entire unlobed	Partly serrated or lobed	Consistently serrated
Herbaceous to subshrubby	4a	4a	1b
Woody twiners/tend./scram.	4c	5c	3d
Woody surface-ascenders	3c	4c	3d
2. DIVIDED LEAVES P= 0.80 (3×3); 0.42 (2×2)	Simple unlobed	Lobed or varied	Compound
Herbaceous to subshrubby	3a	2a	4b
Woody twiners/tend./scram.	6c	4c	4d
Woody surface-ascenders	5c	1c	4d
3. OPPOSITE LEAVES P=0.005 (3×2); 0.004 (2×2)	Alternate leaves	Opposite leaves	
Herbaceous to subshrubby	9a	0b	
Woody twiners/tend./scram.	12a	2b	
Woody surface-ascenders	4c	6d	
4. CHEMISTRY (Table 6) P= 0.03 (3×3); 0.13 (2×2)	Alkaloids etc. (N 1st or 2nd under CHE)	Mixed (marked * under CHE)	Terpenoids or phenolics (T/P)
Herbaceous to subshrubby	4a	3a	2b
Woody twiners/tend./scram.	3a	1a	10b
Woody surface-ascenders	0c	2c	8d
5. SEXUALITY (Table 6) P= 0.22 (3×3); 0.05 (2×2)	Bisexual or monoecious	Partly dioecious (various)	Strictly dioecious
Herbaceous to subshrubby	4a	1a	4b
Woody twiners/tend./scram.	9a	3a	3b
Woody surface-ascenders	8c	2c	0d

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6. SERRATED LEAVES P= 0.39 (3×3); 0.07 (2×2)	Entire unlobed	Partly serrated or lobed	Consistently serrated
Herbaceous or subshrubby	4a	4a	1b
Largely deciduous	3c	6c	5d
Largely evergreen or mixed	6a	4a	1b
7. DIVIDED LEAVES P= 0.74 (3×3); 0.12 (2×2)	Simple unlobed	Lobed or varied	Compound
Herbaceous or subshrubby	3a	2b	4b
Largely deciduous	6a	3b	5b
Largely evergreen or mixed	7c	1d	3d
8. OPPOSITE LEAVES P=0.06 (3×2); 0.07 (2×2)	Alternate leaves	Opposite leaves	
Herbaceous or subshrubby	9a	0b	
Largely deciduous	11a	3b	
Largely evergreen or mixed	6c	5d	
9. CHEMISTRY (Table 6) P= 0.004 (3×3); 0.30 (2×2)	Alkaloids etc. (N 1st or 2nd under CHE)	Mixed (marked * under CHE)	Terpenoids or phenolics (T/P)
Herbaceous or subshrubby	4a	3a	2b
Largely deciduous	3a	0a	11b
Largely evergreen or mixed	0c	3c	8d
10. SEXUALITY (Table 6) P= 0.44 (3×3); 0.21 (2×2)	Bisexual or monoecious	Partly dioecious (various)	Strictly dioecious
Herbaceous or subshrubby	4a	1a	4b
Largely deciduous	9a	3a	2b
Largely evergreen or mixed	8c	2c	1d

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Table 10. Geological eras from first appearance up to early radiation of each vine genus.

Estimates vary much in reliability. Darker shading indicates based on fossils and/or dated phylogenetic analysis; lighter, same but weaker evidence. Question marks are estimates only from phylogeny and biogeography. Within each genus, later dates indicate when relatively diverse lineages are likely to have begun, including some in mid-temperate zones. Genera are listed in approximate order of age, but with additional separation as follows.

(a) Genera that are largely deciduous in mid-temperate regions; * evergreen in warmer.

(b) Evergreen-tending genera with at least one modern mid-temperate evergreen vine; asterisks indicate partially evergreen genera (*) or fully evergreen genera (***) today.

(c) Largely herbaceous taxa in mid-temperate zones today (not a comprehensive list); these are included for comparative purposes, since they do have woody relatives.

Indicates genera with at least some ability to climb trees using adventitious roots or pads.

Bottom two rows show first North American indications of genera with deciduous trees in temperate regions, based largely on fossils, in the following two groups.

(d) Largely typical of mesic to submesic or subhydric habitats, mostly diffuse-porous:

1, *Acer* (Renner et al. 2008); 2, *Aesculus* (Manchester 2001); 3, *Alnus* (Forest+ 2005); 4, *Carpinus* (Forest+ 2005); 5, *Liquidambar* (Pigg+ 2004); 6, *Nyssa* (Wen+ 1993); 7, *Platanus* (Feng+ 2005); 8, *Tilia* (Manchester 1994); 9, *Fagus* (Manchester+ 2004); 10, *Liriodendron* (Nie+ 2008); 11, *Morus* (Zerega+ 2005); 12, semi-ring-porous *Catalpa* (Olmstead+ 2009).

(e) Largely typical of more subxeric or disturbed habitats, mostly ring-porous (except *Ostrya*):

1, *Carya*-like pollen (Muller 1981); 2, *Castanea* (Manos+ 2001); 3, *Carya* (Manos+ 2007); 4, *Fraxinus* (Wallander 2008); 5, *Juglans* (Stanford+ 2000); 6, *Gleditsia* & *Gymnocladus* (Schnabel+ 1998); 7, *Ostrya* (Forest+ 2005); 8, *Quercus* sections *Quercus* and *Lobatae* (Manos+ 1999, 2001); 9, *Robinia* (Lavin+ 2003); 10, *Maclura* (Martinez-C.+ 2006).

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GEOLOG. ERA	uCret.	Paleo.	Eoc.	Oligo.	Mio.	Plio.	References
Millions of years	84-65	65-56	56-34	34-23	23-5	5-2.5	St = Stevens 2012 ref.

(a) DECIDUOUS VINES

<i>Schisandra</i>							Denk+ 2006
<i>Aristolochia</i>							St: Wikstrom+ 2004
<i>Cocculus*</i>							Jacques+ 2011
<i>Ampelopsis</i>							Nie+.10; Zecca+.12
<i>Clematoclethra</i>			?				Chat+ 2004
# <i>Parthenocissus</i>							Nie+.10; Zecca+.12
# <i>Campsis</i>							Xiang+ 2000
<i>Isotrema</i>			?	?			Ohi-toma+ 2006
<i>Calycocarpum</i>			?	?			Jacques+ 2011
<i>Vitis</i>							Nie+.10; Zecca+.12
<i>Rosa*</i>							DeVore+ 2005
# <i>Schizophragma</i>							Samain+.10; Xiang+.11
# <i>Hydrangea</i>							Xiang+ 2011
<i>Menispermum</i>							Jacques+ 2011
<i>Clematis*</i>							Xie+ 2011
<i>L. Caprifolium*</i>							Smith 2009
# <i>Toxicodendron</i>							Nie+ 2009
<i>Berchemia</i>							Smiley+.75, Rich+.04
<i>Wisteria</i>				?		?	Wang+ 2006

GEOLOG. ERA	uCret.	Paleo.	Eoc.	Oligo.	Mio.	Plio.	References
Millions of years	84-65	65-56	56-34	34-23	23-5	5-2.5	St = Stevens 2012 ref.

(b) EVERGREEN-TENDING VINES

<i>S. Smilax</i> *							Dilcher+ 2005
<i>Celastrus</i> *			?		?	?	Mu+ 2012
# <i>Actinidia</i> *			?	?	?		Schönenberger+ 2001
# <i>Decumaria</i> **							Samain+.10; Xiang+.11
# <i>Hedera</i> **							Green+ 2011 & cited.
<i>L. Nintooa</i> **							Smith 2009
<i>Akebia</i> *					?		Wang+ 2002
<i>Holboellia</i> **					?	?	Wang+ 2002
# <i>Bignonia</i> **					?	?	Olmstead+ 2009
# <i>E. fortunei</i> s.l.**					?		Li+.84; Simmons+.01

(c) HERBACEOUS VINES

<i>S. Nemexia</i>			?		?		St: Qi+ 2012
<i>Humulus</i>							Zerega+ 2005
<i>Pueraria</i>							Lavin+.05; Wang+.10
<i>Lackeya</i>						?	Kajita+ 2001
<i>Thyrsanthella</i>						?	Livshultz+ 2007

(d,e) DECIDUOUS TREES

Mesic Genera		1-8	9-11	12?			See also: Graham 1999,
Subxeric Genera		1?	2-5	6-9	10?		Muller 1981 etc.

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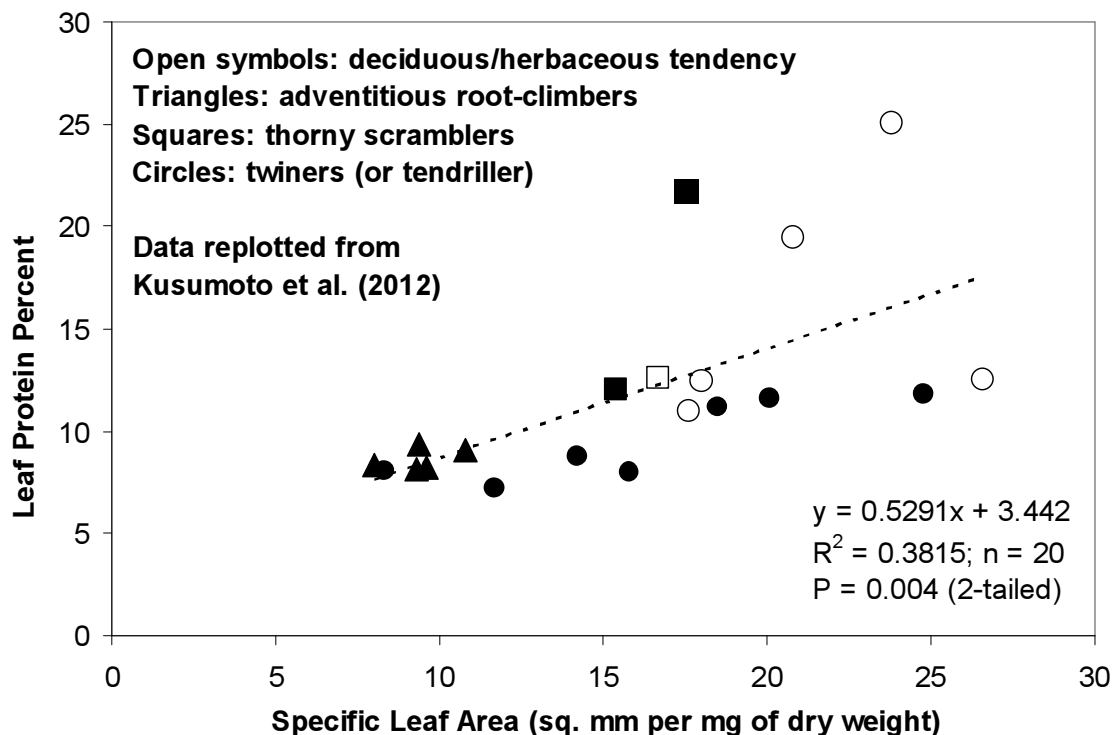


Figure 6a. Estimated leaf protein content (= 6.25 × N% of dry weight) in relation to specific leaf area for the 20 species of vine ('lianas') studied by Kusumoto et al. (2012) in subtropical forest of southern Japan. The three species with >15% protein are the one herbaceous plant with tendrils (at top) and the two N-fixing species among all 20.

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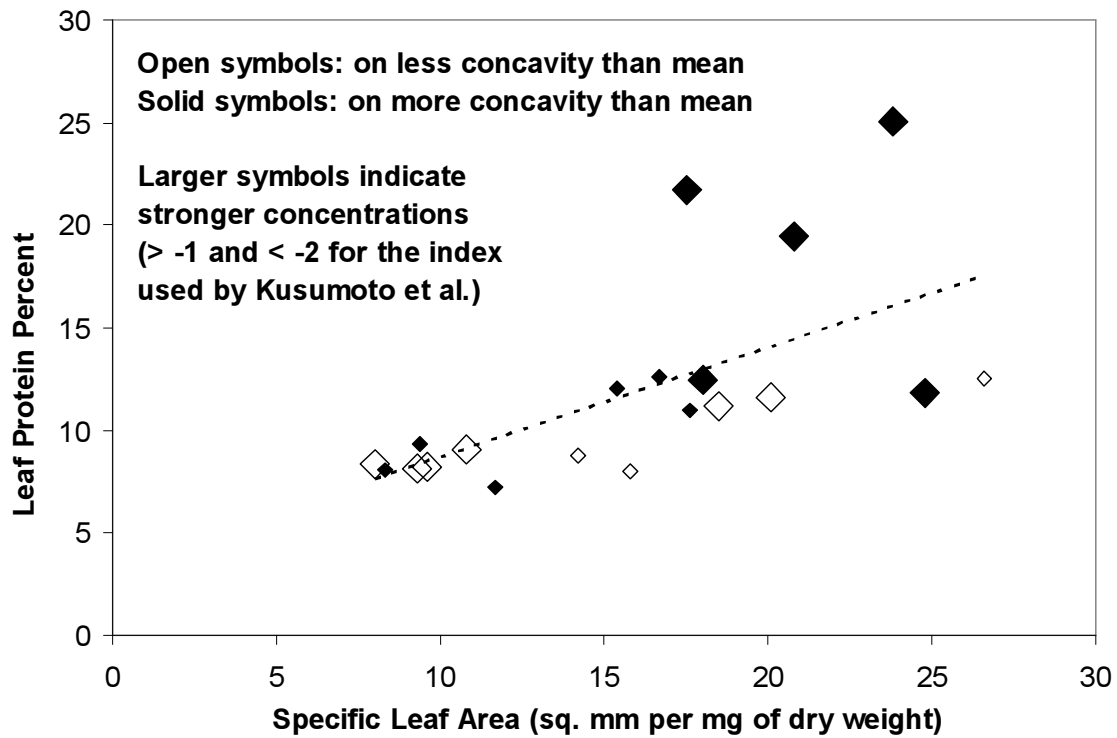


Figure 6b. As in Figure 6a, adding overlay of associations with topographic concavity for each of the species, from the “relief index” calculated by Kusumoto et al. (2012). Their index was significantly correlated with leaf nitrogen ($P < 0.01$) and climbing habit ($P < 0.05$). But there was no correlation of forest basal area with functional characters.

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Figure 7. Approximate modal positions of woody vine species along hydrological gradients of Figure 1. See text for further explanation. Underlined species are evergreen-tending; # climb trees; + have pinnate leaves; () are rare/local species.

				CLIFFS				XERIC EXTREME
					(LONRET)			
MESIC SLOPES (below)	(LONDIO)				VITAES SMIBON			
(SCHGLA)				SMIROT	SMIGLA COCCAR	LONSEM CELSCA		
ISOMAC		+BIGCAP	#PARQUI	VITROT VITBAIL VITVUL	CLEVIR+ MENCAN	(LACMUL+) ROSSET+		
ISOTOM		(HUMLUP*)	#DECBAR #TOXRAD+ SMIHIS	VITLAB VITCIN #CAMRAD+	AMPARB+ (BERSCA)			
		VITRIP* AMPCOR (CALLYO*)	WISFRU+ VITPAL	THYDIF WISMAC+				
RHEIC EXTREME	(VITRUP*)							HYDRIC EXTREME

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Development of Hypotheses, Experiments and Applications

Competition, invasion and herbivory of evergreen-tending vines. There has been much interest in potential interactions of vines with trees, but rather little conclusive experimental work (e.g., Putz & Mooney 1991, Schnitzer & Bongers 2002, Morrisey et al. 2009, Ichihashi & Tateno 2011). Even the effects of smothering species with adventitious roots like *Hedera* remain uncertain, as commented upon by Rackham (1990, p. 24). Some statistical analysis of patterns in woodland has suggested a negative effect of *Hedera* on larger trees (e.g., Garfi & Ficarrotta 2003), and “ivy is often regarded as a problem as it is believed to reduce tree growth rates” (Castagneri et al. 2013). Dillenberg et al. (1993) used a trenching and trellising experiment to show that root competition of *Lonicera japonica* has more effect on growth of *Liquidambar* than does canopy competition, which was only significant in combination with root competition; *Parthenocissus quinquefolia* had much less overall effect. Skullman et al. (2004) showed allelopathic effects of *L. japonica* on pines. There is little evidence that high-climbing vines have distinct ‘host-preferences’ for tree species independent of tree size, succession and environmental factors (Ladwig & Meiners 2010, Leicht-Young et al. 2010, Castagneri et al. 2013, Kusumoto et al. 2012). But Talley et al. (1996) did show that seed-germination of the deciduous vine, *Toxicodendron radicans*, is inhibited by bark chemistry of some trees, especially *Juglans nigra*.

Effects of these vines on ground vegetation within woodland are more obvious in some cases. Across its native and invaded ranges, *Hedera* appears to suppress growth of associated plants (including *Lonicera*) due to direct competition for space and resources—based on long-term observations of succession without ungulates (Harmer et al. 2001) and short-term manipulations (Biggerstaff & Beck 2007a,b). In Kentucky, the local dominance of *Euonymus*

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fortunei has clearly led to much local reduction in cover of native grasses (especially *Elymus* spp.) and herbs (especially summer perennials), as documented in the Shady Lane Walnut Woods at University of Kentucky Arboretum (J.J.N. Campbell & Kim D-H., in prep.). However, Smith & Reynolds (2012) have recently experimented with potted plants, and showed that *Asarum canadense* can depresses growth of *E. fortunei* by ca. 60% through changes in the chemistry of associated soil. They found no significant effect of *Euonymus* soil (versus *Asarum* soil) on growth of *Asarum*.

Rather than invoking direct competition or inhibition (Grubb 1992), parameters of disturbance that are correlated with these vines can sometimes be used to indicate more significant relationships with associated plants than parameters of the vines themselves. For example, Surette & Brewer (2008) found that environmental factors including prescribed fire provided stronger correlations with native plant diversity than did cover of *Lonicera japonica* itself—a species that is reduced by frequent fire. It would be interesting to extend such research to include patterns of browsing by deer—which can also reduce *L. japonica* (Table 8).

Given the apparent competitive ability of some evergreen-tending vines, why do they do not dominate temperate deciduous woodlands in general—what ecological factors could control their populations? The invasion of vines like *Hedera*, *Lonicera* and *Euonymus* species into the southeastern U.S.A. has been partly attributed to release from ‘natural enemies’ that may control them in their native ranges (Schierenbeck et al. 1994, Ding et al. 2006, Ashton & Lerdaun 2008). Perhaps the most obvious type of ‘enemy’ to consider first would be livestock (pigs, cattle, goats, sheep)—which have been a major ecological factor in much woodland of Europe and East Asia for several thousand years, often replacing wild ungulates that roamed before human settlement. Livestock have been especially influential in transitions from

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grassland to woodland on more fertile soils, where invasion by vines is most common. They remain locally important, especially in more hilly regions—although original effects of extinct megafauna were probably concentrated on plains (e.g., Bullock & Pakeman 1997, Pykälä 2000, Vera 2000, Mountford & Peterken 2003, Willson 2006, Melick et al. 2007, Hodder & Bullock 2009, Smith 2010, Garcia 2012). In contrast, although livestock tended to replace large native herbivores in the southeastern U.S.A. during initial centuries of European settlement, open range has been greatly reduced in woodlands here after the 1930s, especially around more densely populated areas. Moreover, fencing generally provides ideal support for vines such as *Lonicera japonica*, while also reducing the movement of larger animals through the tangles.

The ‘herbivore hypothesis’ and its extensions. This hypothesis—with some corollaries—can be stated as follows (Campbell 2012): “Before human civilization, patterns of herbivory in space and time, especially by larger animals, were a major factor maintaining the diversity of woodland on eutrophic soils in temperate regions. If so, the modern decline in naturally-behaving larger animals has often allowed browsing-sensitive plants to increase in situations where they would have been more controlled in the past. Such plants would include some aliens that have now prospered in more populated regions, where even deer and livestock are restricted. In contrast to fire, the original impact of herbivory was concentrated on mineral-rich soils with the most productive vegetation. Moreover, regular browsing as well as rapid decomposition would have reduced fuel-loads in woodland on such soils.”

The general elimination of larger herbivores from civilized landscapes, especially on more eutrophic soils suitable for agriculture, has left fundamental questions about the original ecology, how to conserve remnants, and how to restore functional systems in a post-modern world. These difficult questions have not received sufficient attention around the temperate

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world, except during recent years in Europe (Mitchell & Kirby 1990, VanWieren 1995, Pykälä 2000, Vera 2000, Kirby 2001, Bradshaw et al. 2003, Bakker et al. 2004, Mitchell 2005, Rackham 2006: 90–100, Takatsuki 2008, Hodder et al. 2009, Johnson 2009, Hédli et al. 2010, Kuijper 2011, Garcia 2012, Garcia et al. 2013). In eastern North America, the ‘herbivore hypothesis’ can be developed to help explain much vegetational pattern on more base-rich soils (e.g., Campbell 1989). A central concept is that prolonged intensive browsing is expected eventually to thin out relatively palatable plants on productive soils, but also to select for more tolerant or deterrent species that can predominate in the recovery from disturbance. Thus, a somewhat cyclical process can be envisaged—especially if trails and glades maintained by larger animals form a continually shifting dynamic network over the landscape, responding to changes in the browsable quality of vegetation (Campbell 2012). Such trails and glades might cover a small proportion of the landscape, but have a large influence on regeneration of trees.

Productivity, herbivory, morphology and chemistry. In eastern North America, effects of mammalian herbivores may have been most influential on the most fertile, productive soils—usually with moderate to high pH (ca. 5.5–6.5). Among large trees, in particular, there is a concentration on fertile soils of most species with thorns (Table 1) or with strongly repellent to toxic chemistry: these include *Aesculus*, *Carya* Sect. *Apocarya*, *Gymnocladus*, *Juniperus*, *Maclura*, *Platanus*, *Prunus* and *Robinia* on more fertile soils, versus *Pinus* and *Liquidambar* on less fertile soils (e.g., Atwood 1941, Burrows & Tyril 2001). There is a similar concentration of trees with large fruits and seeds that appear adapted to dispersal in guts of large animals (Table 1)—cattle and deer in the central Ohio Valley still disperse some seeds of the ungulate-adapted tree, *Gymnocladus dioica*, despite a recent claim to the contrary (Zaya & Howe 2009). In addition, herbivory might have contributed to some trends in the gross vegetative morphology of trees, such as the tendency for clonal spread and compound leaves to be more frequent on

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more fertile soils (Table 1). But the potential selective effects of herbivory on morphology (Brown & Lawton 1991, Grubb 1992) will be difficult to separate from the effects of other disturbances.

As summarized in Table 7 and text above, many mid-temperate vines have deterrent chemistry, especially relatively short, herbaceous or deciduous species, but vining taxa tend to have less defensive chemistry than their closest non-vining relatives. The evergreen-tending genera typically lack toxic alkaloids and other complex, specialized compounds that do occur in the deciduous taxa. Instead, their deterrent chemistry is largely based on tannins, terpenoids (including saponins) and steroidal compounds. Such trends would accord with the concept that these evergreen-tending vines are more ‘accessible’ or ‘apparent’ to generalist herbivores during the winter, causing selection for more ‘quantitative’ unspecialized defenses (Feeney 1976, Grubb 1992, Stamp 2003).

Most temperate vines also lack ‘spines’—woody thorns, cauline and foliar prickles. Among evergreen-tending species, spines form only in *Smilax*. A few deciduous species of *Rosa* and *Rubus* are somewhat vining and densely prickly, but also relatively short. Several species of herbaceous vines have rasping prickles (e.g., *Humulus*, some *Mimosa*) or stinging hairs on stems (e.g., *Tragia*), but only *Humulus* is a robust climber. Spines increase among vines of warmer zones (adding species of *Elaeagnus*, *Saurauia* and *Zanthoxylum* in East Asia), especially in the tropics (adding *Calamus* and many other genera)—where hooked spines enhance climbing ability as well as defense from herbivores (e.g., Putz & Mooney 1981, Grubb 1992, Kusumoto et al. 2012). One can hypothesize that most mid-temperate vines have been able to suffer the seasonal herbivory of larger mammals by recovering with vigorous growth in general, escaping into the canopy (Ashton & Lerdaun 2008). The few spiny species (*Smilax*,

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Rosa, *Rubus*) are mostly short and concentrated in brushy transitions from woodland to grassland where larger herbivores may be more frequent. The many shrubs and trees with a thorny tendency (especially deciduous Araliaceae, Berberidaceae, Fabaceae, Rosaceae, Rutaceae) or with a prickly tendency (including the evergreen leaves of some *Ilex*, *Juniperus*, *Mahonia*, *Ulex*) are also typical of such habitats. The relatively defenseless *Ilex vomitoria* of southeastern U.S.A. is much browsed by deer (Halls & Boyd 1982).

An unusual feature in a few vines is variegated coloration on lower leaves, with yellowish to whitish stripes or mottles. Among temperate vines, this occurs only in some species of *Euonymus* (which also becomes purplish during the winter), *Hedera* (*helix* more than *hibernica*) and *Smilax* (especially *bona-nox* and *aspera*)—all evergreen-tending (Figure 1). Variegation of leaves is usually associated with relatively narrow or lobed shape, forming a ‘juvenile’ syndrome that is generally restricted to low shoots in the woods. Such shoots are more exposed to herbivores, especially during winter, and their coloration may have been selected as camouflage from larger animals with poor visual acuity (Givnish 1990). However, in *Smilax* these shoots often also extend into more open sunny areas, where their potential for warning coloration has been suggested (Lev-Yadun 2009). As reviewed by Givnish and Lev-Yadun, one can conceive of various interactions between leaves like these and the structure, chemistry or herbivory of vegetation, but there has been almost no experimental investigation.

Sexuality. Another character that may be influenced by herbivory is the dioecious tendency, including various subdioecious or polygamo-dioecious conditions (with bisexual plus unisexual individuals). Such sexual separation occurs in only 5-10% of all vascular plants but it is relatively frequent among vines (Renner & Ricklefs 1995). In the complete native plus alien flora of the Carolinas, Conn et al. (1980) reported an increase in dioecy (broadly defined) from

12.3% of 173 tree species, to 13.8% of 282 shrubs, to 15.7% of 102 woody or subshrubby vines. This trend is enhanced in Kentucky (Campbell & Medley 2012), with higher proportions of species overall: 21.5% of 107 trees, 16.7% of 156 shrubs, and 47.7% of 44 vines (or 41.7% of 24 vining genera). Included here as dioecious vines are *Celastrus scandens* (Mu et al. 2012) and *Rosa setigera*, a somewhat vining plant (Kemp et al. 1993), plus the eight species of *Vitis*—which are generally described as dioecious or polygamo-dioecious (Moore 1991, Olien 2001, Aradya et al. 2012). If the eight alien vines in Kentucky are excluded (of which only *C. orbiculatus* is dioecious-tending), the proportion is 55.6% (or 47.6% of 21 genera). These high proportions—as elsewhere in the Ohio Valley—may be part of a general association with nutritional factors. Among trees of Kentucky, at least, the dioecious tendency is associated with moderate to high soil fertility (Table 1).

Dioecious percentages of 15–45% (including polygamous transitions) may be typical for vines in subtropical to mid-temperate regions. High proportions among woody plus herbaceous vines are also reported from subtropical to mid-temperate, mineral-rich regions of China: 31.5% of 130 species in part of Zhejiang (Cai 1999); 32.4% of 330 in part of Hunan (Yan 2007); 35.5% of 62 in part of north Yunnan (Chen & Li (2008b); and 35.5% of all 661 in southeast China (Cai & Song 2000). Lower proportions have been reported from some of the most tropical regions in China: 28.7% of 115 in part of south Yunnan (Chen & Li 2008a); 22.1% of 340 in Taiwan (Tseng et al. 2008). At the depauperate extreme, in Europe plus the Mediterranean region, there are only about eight native woody vines, of which two southern species are dioecious (*Smilax aspera*) or partially so (*Vitis vinifera*); the proportion could be higher if herbaceous vines are included (with several species of *Bryonia* and *Tamus* but also *Aristolochia*). In contrast, only 1–20% of vines (lianas) in most fully tropical floras are reported

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to be dioecious—proportions that are generally lower than among the trees (e.g., Renner & Ricklefs 1995, Gillespie 1999, VanDulmen 2001, Matallana et al. 2005, Machado 2006).

It is notable, then, that none of the six genera with strictly evergreen vines in mid-temperate regions have any degree of dioecy (Table 7)—despite several other strictly evergreen woody plants of mid-temperate zones being strictly dioecious (hollies, junipers, mistletoes, yews). In contrast, 3/5 of the genera with mixed evergreen and deciduous condition are dioecious or partially so; 4/14 of the largely deciduous genera; and 4/9 of the herbaceous to subshrubby taxa considered here ($P = 0.08$ with χ^2 test of strictly evergreen versus the rest). There is also a negative association between strict dioecy and the ‘ascending’ growth form with adventitious roots or pads (Table 9.5). Globally, vining taxa tend to have higher numbers of species than sister taxa (Gianoli 2004) and dioecious taxa tend to have fewer (Heilbut 2000), but there are interesting exceptions. Among mid-temperate taxa, strictly dioecious genera have few species and are rarely invasive, but dioecious-tending genera tend to have wide global ranges (Table 8b). Moreover, genera with more than 10 vining species (Table 7) all have labile sexuality (partially dioecious or polygamo-dioecious: *Clematis*, *Vitis*, *Actinidia*, *Celastrus*) except for *Smilax*, which is strictly dioecious. Among woody plants of Kentucky, there are also intriguing associations between dioecy and habitat gradients (Table 1, Appendix 3).

How might these varied trends in sexuality be explained? Renner & Ricklefs (1995) suggested that successful fruit production by vines is particularly expensive—in terms of resources diverted from critical upward vegetative growth—and that this functional problem has led to delayed femaleness and then sexual specialization. Their hypothesis might apply to dioecious deciduous species, especially typical twiners, scramblers and tendrillers that are relatively light-demanding. But it may not apply to more the more shade-tolerant, slow-

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growing, ground-covering, evergreen-tending tree-ascenders. Moreover, the obligate outcrossing from sexual specialization in deciduous vines might be critical for rapid local genetic segregation (micro-evolution) in the more unpredictable biotic environments of larger forest gaps and transitions to open land. Instead, some of the evergreen-tending genera have relatively high chromosome numbers (Table 6), suggesting that polyploidy or other duplication of genome sections has been more important in their success.

It is also conceivable that intense herbivory in vine-rich eutrophic woodland has contributed to dioecy through sexual selection, with more defense in females than males. Again, evergreen-tending species can be excepted since they are more ‘apparent’ to larger herbivores during winter, increasing selection for general chemical defense. In those species, it is likely that any unusual male-tending genotypes with less defense would be extirpated by continual consumption for several months of the year. Some studies have shown that males in dioecious plant species often suffer higher degrees of herbivory than females, and tend to grow faster or have other vegetative differences (e.g., Ågren 1987, 1988, Jing & Cooley 1990, Dawson & Ehleringer 1993, Ashman 2002, Stevens & Esser 2009; but see, Niesenbaum 1992, Retuerto et al. 2006). Differential responses to pathogens may also be involved, which needs much more investigation (Williams et al. 2011, Vegas-Frutis et al. 2012). While varied theories linking dioecy with consumers exist, it will be impossible to make real advances in understanding without directly measuring the effects of consumption.

Effects of larger herbivores on vines and associated vegetation. Table 11 summarizes evidence from studies of browsing effects of deer or livestock on evergreen-tending vines, which indicate general reductions of at least 25–90% within periods of several years. Deciduous vines and shrubs were much reduced in most cases as well. But statistical

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significance is lacking for some individual results, and most of these observations refer just to *Smilax* species, *Lonicera japonica* or *Hedera helix* in the southeastern U.S.A. or western Europe. Much more long-term research, in diverse regions and with a wide variety of vegetational parameters, will of course be essential for deeper understanding (Hester et al. 2000, Russell et al. 2001).

In a more general review of deer effects, Rooney (2009) showed that there is a general shift from dicot herbs and woody plants towards graminoids, ferns or lycopods. It should not be surprising that large herbivores can reduce palatable plants, but there are more fundamental issues in overall ecology and conservation—what plants replace the more sensitive species; are the replacements native (or otherwise desirable); and can the effects of herbivores be used to restore functioning systems? Details of the research summarized in Table 11 do provide some initial insights—abundant replacing species are native in some cases (e.g. *Asimina*, *Impatiens*, *Pteridium*, *Toxicodendron*) but weedy aliens have invaded in other cases. It is likely that results vary much depending on the timing and intensity of browsing. In particular, the appropriate seasons for more intense browsing deserve special consideration.

Remarkable insight to some montane East Asian bamboo forests was provided by Takatsuki (2009), who found that enclosure from dense populations of sika deer led to large increases in woody vines, shrubs and tree regeneration. The vines—deciduous *Actinidia arguta* and *Celastrus orbiculatus*—were virtually absent from the browsed forest. Initial work in the U.S.A. has confirmed that the highly invasive vine, *C. orbiculatus*, can be greatly reduced by deer (Rossell et al. 2007, Averill 2012). The associated evergreen bamboo in Takatsuki’s study (*Sasa nipponica*) also increased initially but after 12 years its biomass had become more or less equalized and leafy cover was 70% lower than the browsed condition, apparently due to strong

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competition from the released woody dicots. His observations—plus evidence that deer concentrate in temperate bamboo forests during winter (Igota et al. 2004)—accord with the idea that large herbivores have enhanced the cover of bamboo in such zones. Similar ungulate effects are suggested for much of the *Arundinaria gigantea* that used to cover lowlands in the southeastern U.S.A. (Campbell 1989, Platt & Brantley 1997). Yet other research on East Asian mountains has shown that browsing can sometimes allow relatively unpalatable weeds, shrubs and trees (such as *Picea*) to grow up within bamboo thickets (Takahashi & Kaji 2001, Nomiya et al. 2003, Darabant et al. 2007, Seki et al. 2012).

How might browsing effects vary in space and time? Dennis (1997) has provided perhaps the only detailed published research on effects of different seasons for browsing in temperate woodland. Although not based on a true experiment, her observations of woodlots compared those with no recent browsing by cattle (but probably more deer), cattle mostly in Apr to Nov, cattle mostly in late Jun to Nov, and cattle mostly in Dec to Mar; there were two sampled transects of 50 m² per treatment. The ‘deferred’ browsing of cattle in late Jun to Nov appeared to support the most native species among grasses, herbs and shrubs (ca. 83 versus 48 in the controls); there were few additional aliens (8 versus 4)—but *Lonicera japonica* was most abundant in this treatment, compared to all others. Woodlots browsed by cattle in Dec to Mar had the lowest cover of vines, shrubs and tree seedlings, with *Lonicera* completely absent, but alien grasses and other weeds were relatively frequent (19 species).

Patterns of herbivory may be substantially different on the modern landscape, compared to earlier periods. For example, historical records of “buffalo” (*Bison bison*) during 1750–86 in north-central Kentucky show distinct concentrations during May–Jun and Nov (Campbell 2012), and some of the pioneer literature suggested seasonal migrations from north in summer

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to south in winter (Roe 1951, Belue 1996). Extinct megafauna of eastern North America probably migrated in similar ways (Hoppe & Koch 2007, and their citations)—expanding licks into glades and moving much matter (Haynes 2012). In contrast, the current effects of white-tailed deer on woodland in Kentucky tend to be spread through the whole year, or may be most intense during winter. It is likely that the modern abundance of food for deer in farmland and along edges for much of the year—especially during summer—has substantial effects on browsing within many adjacent wooded areas. And the general lack of predators today, except for humans during hunting seasons, probably allows deer to forage in openings that would have been more exposed, dangerous, avoided places before Virginian settlement.

It is hypothesized here that (Aug–)Sep to Nov(–Dec) would be the most effective season for intense browsing to shift the balance from alien to native plants in eutrophic woodlands of east-central U.S.A. During those months, most natives have stopped growing new shoots or are dormant, while most of the more problematic alien plants tend to remain exposed and some keep growing—especially evergreen-tending vines like *Euonymus fortunei*, tardily deciduous shrubs like *Lonicera maackii*, biennials with rosettes like *Alliaria petiolata*, and fall-germinating winter annuals like *Stellaria media*. Tests of such hypotheses will be as important to restoration of eutrophic woodlands in east-central states as research on the effects of different fire seasons in pine-oak woodland of more southeastern states (e.g., Glitzenstein et al. 1995, Sparks et al. 1998, Flory & Lewis 2009).

It is also important to pursue more systematic research into how herbivory varies from deeply shaded forest interiors to tree-fall gaps to more open woodland, because the regeneration (and natural selection) of many vines, shrubs, trees and other plant species has probably been concentrated in gaps and openings (Grubb 1977, 1992, Hulme 1996). Although

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much information has been accumulated on spatial patterns in deer browsing, there have been surprisingly few studies that have linked this information to models of forest dynamics (Russell et al. 2001). Gap-related patterns appear to be highly varied. Some recent research has indicated little or no concentration of herbivory by deer in smaller gaps (e.g., Moser et al. 2008). But browsing on woody plants can be greatly influenced by presence of alternative higher quality shrubs (especially *Rubus*), herbs and grasses (e.g., Moser et al. 2006, Rackham 2006: 538, Abbas et al. 2012). Deer often forage into nearby farmland rather than focussing on small interior forest-gaps, where unusually dense populations may nip regenerating plants in the bud before substantial forage grows up. And sheep are clearly lured away in the winter from deeper deciduous woods with evergreen-tending undergrowth, if there is better forage nearby amongst cool-season grasses and herbs (e.g., Garin et al. 2000).

Comparison of browsing with burning effects. A corollary of the general ‘herbivore hypothesis’ for eutrophic woodland (as stated above) is that burning should decrease in relative importance with more browsing on damper, richer soils. Again, there have been surprisingly few studies in temperate forests that explore such relationships. However, general knowledge of forests in the east-central U.S.A. indicates that fires are most frequent on somewhat dry infertile soils—in the upper left sector of Figure 2 (e.g., Campbell et al. 1991). In contrast, effects of bison, elk, deer and beaver appear to have been most intense on more moist and fertile soils—in the lower right sector of Figure 2 (e.g., Campbell 1989).

It is well known that *Lonicera japonica* often occurs on moderately dry or infertile soils sites (Figure 1f), and that it can be reduced by repeated burning (Barden & Mathews 1980, Nuzzo 1997, Kush et al. 2000, Munger 2000, Wang et al. 2012). However, its vigorous resprouting after fires can produce more accessible browse for deer (Stransky 1984), and the

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potential for interaction with deer deserves further investigation. *Hedera* (ivy) provides an interesting contrast with *Lonicera*. Metcalfe (2005) noted: “Cutting and grazing generally reduce the competitive abilities of ivy, and it shows low tolerance of fire when it does burn [see also Úbeda et al., 2006]. However, being evergreen and with a relatively high water content, ivy is slow to burn and will not readily spread fire well. Consequently, ground planting with ivy has been proposed to reduce fire risk in seasonally dry areas (e.g. Utah Bureau of Land Management 2001).” Species of *Vinca* have also been recommended for living firebreaks in some states (e.g., Lippi & Kuypers 1998).

Potential importance of smaller herbivores and pathogens. Could consumers other than large herbivores influence vine populations? This question remains largely unanswered. For example, there might be considerable effects of small mammals on evergreen vines that cover the ground. In western Europe, the bank vole (*Clethrionomys glareolus*) is often associated with *Hedera*, but its potential use of the plant in winter does not seem to have been investigated (Alain et al. 2006). Larger birds (Anseriformes, Galliformes) can also feed much on these vines, which provide relative palatable forage that can sometimes balance effects of more toxic evergreen plants in the diet (e.g., Hewitt & Kirkpatrick 1977). It is likely that turkeys and geese, in particular, can influence establishment of woody seedlings in some habitats (Clark & Gage 1995, Hulme 1996, Rinkes 2004, Vera 2009). One might expect invertebrates to play an insignificant role during winter, compared to warm-blooded animals, but do evergreen leaves allow populations to build up on them in dormant states? Some types of insects, such as adelgids and scales, are often associated with evergreen plants (McClure 1989, Ward et al. 1995).

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Ding et al. (2006) outlined prioritized “natural enemies” for trials in biological control of East Asian plants in North America, including *Euonymus fortunei* and some deciduous vines (*Ampelopsis brevipaniculata*, *Celastrus orbiculatus*, *Pueraria montana*). They identified 40 species of arthropods, fungi and microbial pathogens on *Euonymus* in China, 13 of which are limited to this genus and thus potential agents to develop. For *Ampelopsis*, these numbers are 22 and 4; for *Celastrus*, 9 and 5; for *Pueraria*, 200+ and perhaps 3+, but some of these can also feed on *Amphicarpaea* and *Glycine* (Frye et al. 2007, Imai et al. 2011, Ruberson et al. 2012). For *Hedera helix*, they are 122+ and perhaps 22+ (Metcalf 2005); for *Lonicera japonica*, at least 16 (Larson et al. 2006).

There is little published evidence that any of the evergreen-tending vine species are controllable to a large extent by arthropods or pathogens, but further investigation is needed. Most information comes from cultivated plants. The ‘euonymus scale’ insect (*Unaspis euonymi*) is an Asian pest of *E. fortunei* and other Celastraceae, that was estimated to cause about 10% annual mortality in planted *E. fortunei* across southern New England (VanDriesche et al. 1998)—considered an economic problem for the horticultural industry! Also, a cosmopolitan anthracnose fungus (*Colletotrichum gloeosporoides*) has caused much loss to *fortunei* in nurseries (Ningen et al. 2005). Both the scale and the anthracnose generally cause more damage in sun than shade, and there is no evidence that they have caused significant damage to populations of the plant that have escaped into wooded areas. Several ‘leaf spot’ fungi are common aesthetic problems on *Hedera helix*, but, again, none are documented to cause significant problems for wild plants (Metcalf 2005, Waggy 2010).

There has been much recent interest in potential biological control of the largely herbaceous vine, *Pueraria montana* (= *P. lobata*, kudzu). This species has been common in the

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southeastern U.S.A. for 90 years, with virtually no reduction by pests or pathogens until spread of the Asian plataspid (shield bug), *Megacopta cribraria*, after 2009. In Georgia, Zhang et al. (2012) found that feeding of this bug led to a total biomass reduction of 32.5 % during the first year of infestation. Frye et al. (2012) showed that such levels of damage could cause reduction but only if repeated for several years. However, the bug also damages soybeans and some native legumes, so will not be promoted for biological control of *Pueraria*.

Integration of experimental trials, woodland restoration and land economy. Several vines have become problematic invasive species outside their native ranges, especially among the evergreen-tending genera (Table 7, 8a). Is there any evidence that substantial control of alien vines—evergreen or deciduous—is possible, as a cost-effective enterprise within the context of modern society?

Many papers have been written about these aliens, much has been recommended, and considerable effort has been made at control within some tracts of land. Leaders in the U.S.A. have included organizations such as the The Nature Conservancy, National Park Service, U.S. Forest Service and their partners in state governments. Methods for manual, mechanical or chemical control have been generally determined. But the potential for biological control is still uncertain, and trials with livestock have been rare. Moreover, the fundamental ecological and economic questions remain largely unanswered.

- (1) Can naturalistic ‘macro-management’ of habitats shift the composition from alien to native species, so that expensive ‘micro-management’ of individual species is minimized?
- (2) Can any management plan be implemented across large areas and be sustained by regular income and fees from that land, or (less likely) by donations and taxes from afar?

There has been remarkably little published evaluation of success versus failure in our varied options for management to control these vines across North America. For *Hedera*, Biggerstaff & Beck (2002, 2007a,b) studied the effectiveness of manual versus chemical control and subsequent recovery of native plants. Ingham (2008) compared control of *Hedera* by goats with mowing. However, these projects covered only a few years, and did not address deeper issues of economics or ecological sustainability. The direct or indirect costs of invasive species to society in general have sometimes been estimated (Pimentel et al. 2005). But while mechanical and chemical control are often prescribed for control of alien plants, there have been few publications on the direct continuing costs of using these methods for keeping populations down to an acceptably low standard—other than general reviews and theories (D’Antonio 2004, Olson 2006, Kim et al. 2007, Epanchin-Neill & Hastings 2010, Kettenring et al. 2011). Even simple statistics are inconsistently published in most regions, if at all, such as annual expenditures per acre and degrees of reduction in aliens at specific sites. And there appears to have been no proper analysis of the costs and benefits from using livestock—or perhaps wildlife management—to help control unwanted plants within temperate woodlands designed for native biological diversity. A more systematic approach is needed, as is being developed for rangelands in western states (Masters & Sheley 2001, Finnoff et al. 2008).

Even at well-known sites with much visibility in eastern states, background research and repeated effort, there does not seem to be an adequate stream of public information. For example, parks in or near Washington D.C. have had much invasion of *Hedera* that has been studied for over 30 years (Thomas 1980, 1988, Putz 1995, Swearingen & Diedrich. 2004). Organized control has been tried for up to a decade (e.g., Young et al. 2012). The National Park Service is now reportedly cooperating on *Hedera* reduction with the Rock Creek Conservancy (2012). And in addition to staff and volunteers, a private company was contracted for at least

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five years to work on 176 acres (Invasive Plant Control Inc. ca. 2008). Yet the recent General Management Plan for the park made no mention of invasive plant species (NPS 2007 and associated documents), and there does not appear to be any freely available official report about invasive plant reduction. Moreover, the recent Environmental Impact Statement for control of deer in Rock Creek Park (NPS 2012) makes virtually no reference to one potential benefit of high deer density in some places—reduction of some alien plants. A photograph was shown (p. 22), with caption: “Deer enclosure/fenced plot at edge of forest overrun by invasive plants. The plot was discarded because the number of nonnative plants biased the data.”

There is currently a dearth of institutional interest in using large herbivores to reduce alien plants, recover natives, and restore structure within eutrophic deciduous woodlands. There have been a few experiments, with promising results (Brockway & Lewis 2003, Compton et al. 2003, Darabant et al. 2007, Harrington & Kathol 2009, Hedtcke et al. 2009). But there has been virtually no extension to larger scales on a continuing schedule. A few conservationists have urged deeper involvement (Cramer 1991, Dennis 1997, Williams 1997, Brower & Dennis 1998), and agriculturalists have sometimes outlined concepts of ‘agroforestry’ (Rule et al. 1994, Buffum et al. 2009, Guyer & Ponder 2012), but there appears to be general disinterest or resistance. Critics usually point to the ecological damage that extended use of livestock can cause (Dambach 1944a,b; Johnson 1952), without considering the value from short periods of intensive browsing that could simulate the original seasonal patterns of larger herbivores. With deer alone, there are well-documented cases of high densities promoting some aliens—such as *Alliaria petiolata* or *Microstegium vimineum* (Webster et al. 2008, Eschtruth & Battles 2009, Knight et al. 2009). More basic problems are the lack of generally accepted ecological models for original vegetation, a decline of knowledge and experience with livestock in modern society, and general aversion to dealing with large animals.

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In the Bluegrass region of Kentucky, with unusually fertile soil on phosphatic limestone, there is evidence of much ancient interaction between larger herbivores and native vegetation—but no evidence of regular fires before Virginian settlement (Campbell 1989, McEwan & McCarthy 2008). Griffith Woods is a 745 acre farm that presents the best opportunity to restore something like the original woodland. The Nature Conservancy and University of Kentucky initiated a partnership here in 2003 for research and management, including comparisons of browsing, burning and mowing effects (Crowley 2002, Berry 2007). In association with this project and the National Fish & Wildlife Foundation, a cooperative weed committee was supposed to be established for the region (Campbell 2004b). *Euonymus fortunei* is one of the most problematic invasive plants in Bluegrass Woodlands, together with bush-honeysuckle (*Lonicera maackii*) and garlic-mustard (*Alliaria petiolata*). Yet there is much circumstantial evidence that cattle have kept the *Euonymus* out of woodland pastures at Griffith Woods and elsewhere (Figure 8). Ideally, cooperative planning for this site will enable long-term studies of how livestock can be used to advance ecological goals, integrated into a sustainable economy.

Just as a return to prescribed fire has proliferated within recent decades, it is reasonable to expect diverse benefits from a careful return to more management of ungulates. For example, the concept of regular browsing by sheep and goats in fenced blocks of larger urban parks has considerable traction—to reduce some alien plants at their most sensitive season, to provide local meat for the community, to allow long-term research with much visibility, to reconnect people with more ecological and economic heritage (Berry 1990). Both animals are known to relish vines like *Hedera* (Table 5), and sheep appear to consume *Alliaria* (garlic mustard). The latter is virtually absent from pastures of all types in Britain (Tansley 1939), and W. Berry (pers. comm.) observed substantial reduction of it after autumnal grazing by sheep in a large enclosed plot near Port Royal, Kentucky.

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It is suggested here that satisfactory control of invasive plants in the remnants of temperate woodland on eutrophic soils of the eastern U.S.A. will only be possible with three basic organizing principles among human beings.

- (1) Regional networks at a moderate spatial scale, large enough to allow a critical mass of local knowledge focussed on relatively homogeneous regions, not spread over too many states.
- (2) Regular sharing and pooling of data on native composition and trends in invasive species—especially at managed sites, providing transparent accounting for costs and benefits.
- (3) Reasonable discussion and debate about how to approach methods for long-term sustainability that includes overall well-being of human communities.

We should avoid sentimental attachments to the past, but still try to understand it better. With the ‘post-modern’ age of conservation, which has actually been brewing for millenia, a deeper functional understanding is needed to manage the reassembled plants and animals—retaining as much of the old as possible but applying the new towards reasonable balances. For clear thinking in North America (e.g., Porter & Underwood 1999, Wright 1999), it is good to compare notes from around the world (e.g., Vera 2009, Hughes et al. 2012). Some of the most difficult lessons may come from New Zealand, where extinct large avian herbivores were replaced by mammals but left many endemic spiny or cryptic plants (Brown & Lawton 1991, Grubb 1992). Conservationists and ecologists there are grappling with some of the most radical current changes within temperate forests anywhere (Parkes 2008, Tanentzap et al. 2009).

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Table 11 [next page]. Summarized apparent effects of cattle or deer on evergreen-tending vines, from studies of exclosures or from comparisons of different browsing histories.

Studies are listed alphabetically by authors (right column). Note that statistical significance is lacking in several individual cases, and only the studies of exclosures include proper controlled experiments; see text for discussion. See Appendix 1 for abbreviations of vine species.

Responses indicated here are estimates based on varied measures of stem density or cover (not frequency in plots), unless otherwise stated; ? indicates unclear or varied response. The entries from Griggs et al. (2006) and Webster et al. (2005) summarize trends in importance values for exclosures and changes in Cades Cove attributed to deer over 20 years.

Species	Treatment summary	Decrease amount	Other vine responses	Other changes in ground vegetation	Author, date, state
<i>Lonicera japonica</i>	deer for 12 years versus exclosures	>90%	–Toxrad 80% – Parqui 60%	general decrease but increases in <i>Asimina</i> , other native tolerators	Asnani+ 2006 OH
<i>Lonicera japonica</i>	high versus low deer density	>90% in winter	– Smirot 80% – Toxrad 70% ? Parqui ? Vitspp	much less shrub layer (<i>Rosa multiflora</i> , <i>Ligustrum sinense</i> , etc.) esp. summer	Beaver 2011 TN
<i>Smilax glauca</i>	high versus low deer density	ca 90% in summer	as above	as above	Beaver 2011 TN
<i>Smilax bona-nox</i>	high versus low deer density	>90% in summer	as above	as above	Beaver 2011 TN
<i>Euonymus fortunei</i>	cattle for 20 years in open woods versus excluded	>90%	decreases in Camrad Parqui Toxrad Smihis Vitvul	general decrease in shrub layer (<i>Rubus</i> , <i>Lonicera maackii</i> etc.) but more saplings of <i>Carya laciniosa</i>	Berry 2007 KY; + author's observations
<i>Lonicera japonica</i>	cattle in winter for >15 years	>90%	+Toxrad 300% –Vitrip 30%	much less shrub layer; more annuals, weeds	Dennis 1997 IL
<i>Hedera helix</i>	numbers of trends outside 6 deer-exclosures	decrease in 83%	– Lonper in 2 + in other 2	general decrease in <i>Quercus</i> , <i>Salix</i> , <i>Rubus</i> , <i>Carpinus</i> , <i>Sorbus</i> etc.	Gill+ 2001 EU (review)

<i>Smilax glauca</i>	deer for 8+ years versus exclosures and initial data	>90%	less vines in general – Smirot>50% – Lonjap (?)	much less shrubs, (Euoame, Gayurs, Vacspp etc.) and seedlings, but increases in Acerub, Tsucan, Rhomax	Griggs+ 2006; Webster+ 2005
<i>Hedera helix</i>	deer for 2 years versus exclosures	ca 50%		shrubs reduced by half (<i>Rubus</i> , <i>Vaccinium</i>); but <i>Erica</i> increased	Gonzalez-Herdandez + 1996 SP
<i>Smilax</i> spp.	deer for 7 years versus exclosures	ca 80%		also ca 90% reduction in <i>Rubus</i> ; less of most trees except <i>Robinia</i>	Harlow+ 1970,1975
<i>Hedera helix</i>	deer for several years versus exclosures	ca 40%	– Lonper>90%	also reduction in <i>Rubus</i> ; increases in <i>Pteridium</i> , grasses	Kirby 2001 EN Nagshead
<i>Hedera helix</i>	deer for 8 years versus exclosures	ca 10%	– others 40%: Ampbr Celorb Euofor Lonjap Parqui Smispp Toxrad Vitaes	general reduction of shrub layer by about half (incl. <i>Euonymus</i> , <i>Rubus</i> , <i>Vaccinium</i>)	Krafft+ 2011 DC
<i>Lonicera japonica</i>	cattle for 4 years (in 2 months/year)	>90%		much less shrub layer (<i>Robinia</i> , <i>Rosa multiflora</i> , <i>Rubus</i>)	Luginbuhl+ 2000 NC

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<i>Hedera helix</i>	ponies, cattle and deer for 22 years versus deer only	60% less in plot frequency	– Lonper 60% less in plot frequency	>90% decrease in <i>Rubus</i> frequency; 40% increase in <i>Pteridium</i> ; 100%+ in grasses	Putman+ 1989
<i>Lonicera japonica</i>	deer for 2 years versus exclosures	>90%	– Parqui 60% ? Smispp ? Vitrot	5-10% decrease in woody plants <1.4 m; esp. <i>Rubus</i> ; increases in <i>Fagus</i> , <i>Vaccinium</i>	Thrift 2007 SC
<i>Lonicera japonica</i>	high versus low deer density (many years)	>90% ?	– Parqui 30% – Smispp 15% – Vitrot 15%	ca 40% decrease in woodies <13 cm dbh; esp. <i>Quercus</i> , <i>Prunus</i> , <i>Liriodendron</i> , <i>Oxydendrum</i> ; increases in <i>Cornus</i> , <i>Nyssa</i>	Thrift 2007 SC
<i>Decumaria barbara</i>	high versus low deer density	>50%	as above	as above	Thrift 2007 SC
<i>Smilax</i> spp.	deer for 2-5 years versus exclosures	2–40% but recovered	– Vitspp 70%?	ca 15% decrease in woody plants <1.4 m; esp. <i>Acer saccharum</i> , <i>Prunus serotina</i> , <i>Betula lenta</i> , <i>Rubus</i> ; but increase in <i>Fagus</i>	Thrift 2007 WV
<i>Hedera helix</i>	cattle continual for 4 years	> 90%	–Vinmin>90%	much less <i>Rubus</i> ; less vernal herbs	Uytvanck+ 2009 BE

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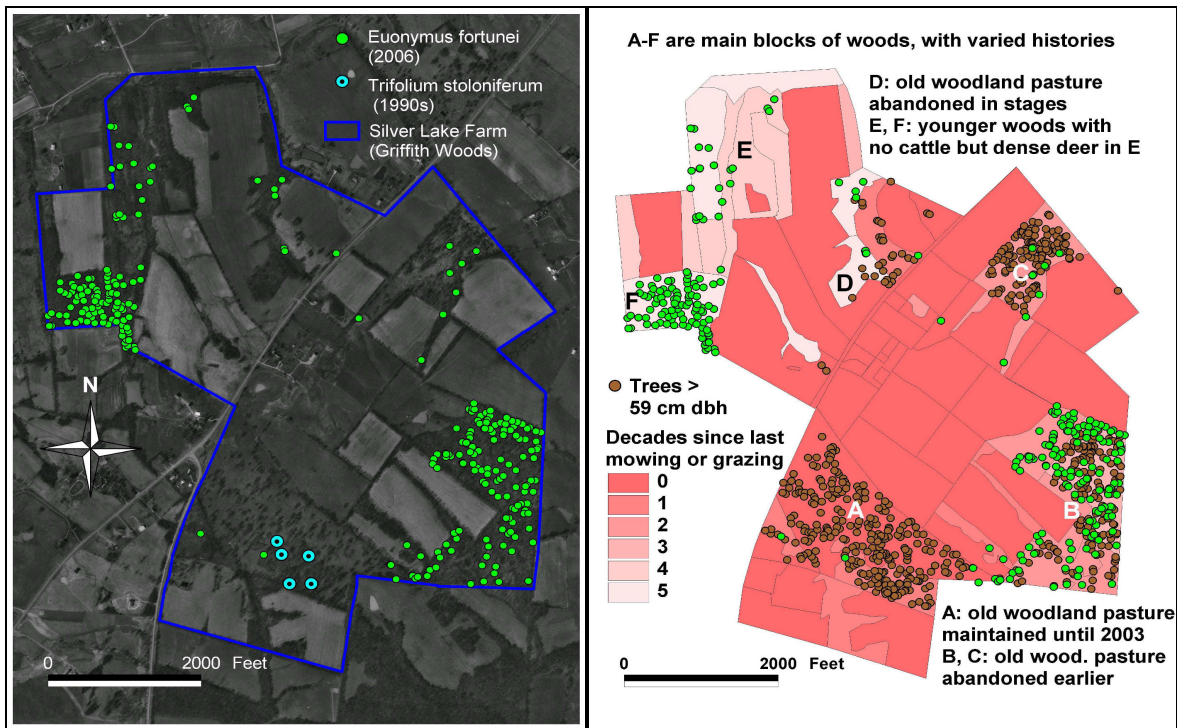


Figure 8. Maps of Griffith Woods (Harrison Co., Kentucky) showing locations of *Euonymus fortunei* and *Trifolium stoloniferum* in relation to management history. These data were assembled in 2003-2007, with assistance of Berry (2007). The old 'savanna' to south (A) was partly unmowed since 1986, leading to thickets of *Carya lacinosalovata* (which cattle do not eat) plus patches of the endangered clover.



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Literature Cited

[*abstracts only seen, full papers not yet available in Kentucky; ** priorities to secure]

- Abbas, F., D. Picot, J. Merlet, B. Cargnelutti, B. Lourtet, J. Angibault, T. Daufresne, S. Aulagnier & H. Verheyden. 2012. A typical browser, the roe deer, may consume substantial quantities of grasses in open landscapes. *European Journal of Wildlife Research* DOI 10.1007/s10344-012-0648-9.
- Aciego-Pietri, J.C., & P.C. Brookes. 2009. Substrate inputs and pH as factors controlling microbial biomass, activity and community structure in an arable soil. *Soil Biology and Biochemistry* 41: 1396–1405.
- Ackerfield, J., & J. Wen. 2003. Evolution of *Hedera* (the ivy genus, Araliaceae): insights from chloroplast DNA data. *International Journal of Plant Sciences* 164: 593–602.
- *Adams, J.M., B. Rehill, Zhang Y-J. & J. Gower. 2009. A test of the latitudinal defense hypothesis: herbivory, tannins and total phenolics in four North American tree species. *Ecological Research* 24: 697–704.
- Adams, W.W., & B. Demmig-Adams. 1995. The xanthophyll cycle and sustained thermal energy dissipation activity in *Vinca minor* and *Euonymus kiautschovicus* in winter. *Plant, Cell & Environment* 18:117–27
- Aday, J., & P. Wyckoff. 2010. White-tailed deer (*Odocoileus virginianus*) as facilitators of a European buckthorn (*Rhamnus cathartica*) invasion into western Minnesota forests. Abstract, 95th Ecological Society of America Annual Meeting.
- Ågren, J. 1987. Intersexual differences in phenology and damage by herbivores and pathogens in dioecious *Rubus chamaemorus* L. *Oecologia* (Berlin) 72: 161–169.
- Ågren, J. 1988. Sexual differences in biomass and nutrient allocation in the dioecious *Rubus chamaemorus*. *Ecology* 69: 962–973.
- Alain, B., P. Gilles & D. Yannick. 2006. Factors driving small rodents assemblages from field boundaries in agricultural landscapes of western France. *Landscape Ecology* 21: 449–461.
- Allen, B.P. 2007. Vegetation dynamics and response to disturbance, in floodplain forest ecosystems with a focus on lianas. Ph.D. dissertation, Ohio State University, Columbus. 242 pages.
- Al-Mamun, M., K. Goto, S. Chiba & H. Sano. 2009. Responses of plasma acetate metabolism to hop (*Humulus lupulus* L.) in sheep. *International Journal of Biological Sciences* 5: 287–292.

- Ammar, H., S. López, J.S. González & M.J. Ranilla. 2004. Chemical composition and in vitro digestibility of some Spanish browse plant species. *Journal of the Science of Food and Agriculture* 84: 197–204.
- **Apsley, D.K., D.J. Leopold & G.R. Parker. 1984. Tree species response to release from domestic livestock grazing. *Proceedings of the Indiana Academy of Sciences* 94: 215–226.
- Aradhya, M., Y. Wang, M.A. Walker, B.H. Prins, A.M. Koehmstedt, D. Velasco, J.M. Gerrath, G.S. Dangl & J.E. Preece. 2012. Genetic diversity, structure, and patterns of differentiation in the genus *Vitis*. *Plant Systematics and Evolution* DOI 10.1007/s00606-012-0723-4
- Ashman, T-L. 2002. The role of herbivores in the evolution of separate sexes from hermaphroditism. *Ecology* 83: 1175–1184.
- Ashton, I.W., & T.M. Lerda. 2008. Tolerance to herbivory, and not resistance, may explain differential success of invasive, naturalized, and native North American temperate vines. *Diversity and Distributions* 14: 169–178.
- Asnani, K.M., R.A. Klips & P.S. Curtis 2006. Regeneration of woodland vegetation after deer browsing in Sharon Woods Metro Park, Franklin County, Ohio. *Ohio Journal of Science* 106: 86–92.
- Asner, G.P., & R.E. Martin. 2011. Canopy phylogenetic, chemical and spectral assembly in a lowland Amazonian forest. *New Phytologist* 189: 999–1012.
- Atwood, E.L. 1941. White-tailed deer foods of the United States. *Journal of Wildlife Management* 5: 314–332.
- Averill, K.M. 2012. Exploring the role of deer browsing preference in plant invasion. Abstract, Ecological Society of America, 97th Annual Meeting.
- Axelrod, D.I. 1966. Origin of deciduous and evergreen habits in temperate forests. *Evolution* 20: 1–15.

163

- Axelrod, D. I. 1983. Biogeography of oak in the Arctotertiary Province. *Annals of the Missouri Botanical Garden* 70:629–657.
- Baars, R., & D. Kelly. 1996. Survival and growth responses of native and introduced vines in New Zealand to light availability. *New Zealand Journal of Botany* 34: 389–400
- Badre, B., P. Nobelis & M. Trémolières. 1998. Quantitative study and modelling of the litter decomposition in a European alluvial forest. Is there an influence of overstorey tree species on the decomposition of ivy litter (*Hedera helix* L.)? *Acta Oecologia*, 19, 491–500.
- **Bajracharya, D., S.J.B. Rana & A.K. Shrestha. 1978. General survey and biochemical analysis of fodder plants found in Nagarjun Hill Forest of Kathmandu Valley. *Journal of the Natural History Museum (Kathmandu)* 2: 105–116.
- **Bajracharya, D., T.B. Bhattarai, M.R. Dhakal, T.N. Mandal, M.R. Sharma, S. Sitaula & B.K. Vimal. 1985. Some feed values for fodder plants from Nepal. *Journal Angewandte Botanik* 59: 357–365
- Bakker, E.S., H. Olf, C. Vandenberghe, K. De Maeyer, R. Smit, J.M. Gleichman & F.W.M. Vera. 2004. Ecological anachronisms in the recruitment of temperate light-demanding tree species in wooded pastures. *Journal of Applied Ecology* 41: 571–582.
- Barbosa-Filho, J.M., & E.V. Leitão da-Cunha. 2000. Alkaloids of the Menispermaceae. *The Alkaloids: Chemistry and Biology* 54: 1–190.
- Barden, L.S. 1978. Regrowth of shrubs in grassy balds of the Southern Appalachians after prescribed burning. *Castanea* 43: 238–246.
- Barden, L.S., & J.F. Matthews. 1980. Change in abundance of honeysuckle (*Lonicera japonica*) and other ground flora after prescribed burning of a piedmont pine forest. *Castanea* 46: 257–260.
- Bartolomé, J., J. Franch, J. Plaixats & N. Seligman. 1998. Diet selection by sheep and goats on mediterranean heath-woodland. *Journal of Range Management* 51: 383–391.

164

- Barua, U., D.K. Hore & R. Sarma. 2007. Wild edible plants of Majuli Island and Darrang districts of Assam. *Indian Journal of Traditional Knowledge* 6: 191–194.
- Bate-Smith, E.C. 1978. Astringent tannins of *Viburnum* and *Hydrangea* species. *Phytochemistry* 17: 267–270.
- Bauer, H., & U. Bauer. 1980. Photosynthesis in leaves of the juvenile and adult phase of ivy (*Hedera helix*). *Physiologia Plantarum* 49:366–372
- Bauer, H., & R. Kofler. 1987. Photosynthesis in frost-hardened and frost-stressed leaves of *Hedera helix* L. *Plant, Cell & Environment* 10: 339–346.
- Bauer, H., & W. Thoni. 1988. Photosynthetic light acclimation in fully developed leaves of the juvenile and adult life phases of *Hedera helix*. *Physiologia Plantarum* 73: 31–37.
- Bazzaz, F.A. 1968. Succession on abandoned fields in the Shawnee Hills, southern Illinois. *Ecology* 49: 924–936.
- Beals, E.W., G. Cottam, R.J. Vogel. 1960. Influence of deer on vegetation of the Apostle Islands, Wisconsin. *Journal of Wildlife Management* 24: 68–80.
- Beaver, J.T. 2011. An evaluation of population estimators and forage availability and nutritional quality for white-tailed deer in Tennessee. M.Sc. thesis, University of Tennessee, Knoxville. 94 pages.
- Beckstrom-Sternberg, S.M., & J.A. Duke. 1994. The Phytochemical Database. ACEDB, Version 4.3 -data version July 1994 [<http://probe.nal.usda.gov:8300/cgi-bin/browse/phytochemdb>; updated at “Dr. Duke's Phytochemical and Ethnobotanical Databases”, <http://www.ars-grin.gov/duke/>].
- Bell, D.J., I.N. Forseth & A.H. Teramura. 1988. Field water relations of three temperate vines. *Oecologia* 74: 537–545.
- Belue, T.F. 1996. *The Long Hunt: Death of the Buffalo East of the Mississippi*. Stackpole Books, Mechanicsburg, Pennsylvania. 288 pages.

165

- Berry, A. 2007. Creation of a Geodatabase for Griffith Woods. M.Sc. thesis, University of Kentucky. 193 pages.
- Berry, W. 1990. *What are People for?* North Point Press, San Francisco. 210 pages.
- Bhatta, R., A.K. Shinde, S.K. Sankhyan & D.L. Verma. 2001. Nutrition of range goats in a shrubland of western India. *Asian & Australian Journal of Animal Science* 15: 1719–1724.
- Biggerstaff, M.S., & C.W. Beck. 2002. Effects of English Ivy (*Hedera helix* L.) and differences in its removal on regeneration of native vegetation in a southeastern Piedmont Forest. Summer Undergraduate Research Program at Emory University [http://www.cse.emory.edu/sciencenet/undergrad/SURE/Posters/2002_biggerstaff.html].
- Biggerstaff, M.S. & C.W. Beck. 2007a. Effects of method of English ivy removal and seed addition on regeneration of vegetation in a southeastern piedmont forest. *American Midland Naturalist* 158: 206–220.
- Biggerstaff, M.S., & C.W. Beck. 2007b. Effects of English Ivy (*Hedera helix*) on seed bank formation and germination. *American Midland Naturalist* 157: 250–257.
- Blair, R.M. 1960. Deer forage increased by thinnings in a Louisiana loblolly pine plantation. *Journal of Wildlife Management* 24: 401–405.
- Blair, R.M., H.L. Short & E.A. Epps. 1977. Seasonal nutrient yield and digestibility of deer forage from a young pine plantation. *Journal of Wildlife Management* 41: 667–676.
- Blair, R.M., & L.E. Brunett. 1980. Seasonal browse selection by deer in a southern pine-hardwood habitat. *Journal of Wildlife Management* 44: 79–88.
- Bledsoe, B.P., & T.H. Shear. 2000. Vegetation along hydrologic and edaphic gradients in a North Carolina coastal plain creek bottom and implications for restoration. *Wetlands* 20: 126–47.
- Bleisch, W., Liu Z., E. Dierenfeld & Xie J. 1998. Selected nutrient analysis of plants in the diet of the Guizhou snub-nosed monkey (*Rhinopithecus [Rhinopithecus] brelichi*). Pages 241–

166

- 254 in: N. Jablonski (ed). *The Natural History of the Doucs and Snub-nosed Monkeys*. World Scientific Publishing, Singapore.
- Boggs, L.L., J.P. Muir & J.W. Dunn. 2012. Greenbriar suppression with goat mob browsing. *Livestock Research for Rural Development* Vol. 24, No. 89.
- Bohm, B.A., K.W. Nichols & U.G. Bhat. 1985. Flavonoids of the Hydrangeaceae Dumortier. *Biochemical Systematics and Ecology* 13: 441–445.
- Borchard, F., H.-J. Berger, M. Bunzel-Drüke & T. Fartmann. 2011. Diversity of plant–animal interactions: possibilities for a new plant defense indicator value? *Ecological Indicators*, in press.
- Borkowska A., & A. Konopko. 1994. The winter browse supply for moose in different forest site-types in the Biebrza Valley, Poland. *Acta Theriologica* 39: 67–71.
- Bostan, C., A. Moisuc & M. Butnariu. 2012. Evaluation of nutritive value in some plants that contain allelopathic compounds of Banat region (western Romania). *Studia Universitatis “Vasile Goldiș”*, Seria Științele Vieții 22: 179–183.
- Boulanger, V., C. Baltzinger, S. Saïd, P. Ballon, J-F. Picard & J-L. Dupouey. 2009. Ranking temperate woody species along a gradient of browsing by deer. *Forest Ecology and Management* 258: 1397–1406.
- Boura, A. & D. DeFranceschi. 2007. Is porous wood structure exclusive of deciduous trees? *Comptes Rendus Palevol* 6: 385–391.
- Boyle, M.F., R.K. Peet, T.R. Wentworth, M.P. Schafale & M. Lee. 2009. Natural vegetation of the Carolinas: classification and description of plant communities of the Upper Tar, Roanoke, Meherrin, Chowan, and Cashie Rivers. Ecosystem Enhancement Program, North Carolina Department of Environment and Natural Resources. 67 pages.
- Bradshaw, R.H., G.E. Hannon & A.M. Lister. 2003. A long-term perspective on ungulate–vegetation interactions. *Forest Ecology and Management* 181: 267–280.

167

- Brady, N., & R. Weil. 2002. *The Nature and Properties of Soils*. 13th edition. 960 pages.
- Breckle, S.W. 2002. *Walter’s Vegetation of the Earth: the Ecological Systems of the Geo-Biosphere*, 4th edition. Springer, Berlin. 547 pages.
- Braun, E.L. 1950. *Deciduous Forests of Eastern North America*. Blakiston, Philadelphia. 596 pages.
- Brockway, D.G., & C.E. Lewis. 2003. Influence of deer, cattle grazing and timber harvest on plant species diversity in a longleaf pine bluestem ecosystem. *Forest Ecology and Management* 175: 49–69.
- Brothers, T.A., & A. Springarn. 1992. Forest fragmentation and alien plant invasion of central Indiana old-growth forests. *Conservation Biology* 6:91–100.
- **Brower, B., & A. Dennis. 1998. Grazing the forest, shaping the landscape? Continuing the debate about forest dynamics in Sagarmantha National Park. Pages 184–208 in K. S. Zimmerer, K. R. Young (eds.), *Nature’s geography: New lessons for conservation in developing countries*. University of Wisconsin Press, Madison, Wisconsin.
- Brown, V.K., & J.H. Lawton. 1991. Herbivory and the evolution of leaf size and shape. *Philosophical Transactions of the Royal Society of London B* 333: 265–272.
- Buffum, B., G. Gratzler & Y. Tenzin. 2009. Forest grazing and natural regeneration in a late successional broadleaved community forest in Bhutan. *Mountain Research and Development* 29: 30–35.
- Bullock, J.M., & R.J. Pakeman. 1997. Grazing of lowland heath in England: management methods and their effects on heathland vegetation. *Biological Conservation* 79: 1–13.
- Burnham, R.J., & A. Graham. 1999. The history of neotropical vegetation: new developments and status. *Annals of the Missouri Botanical Garden* 86: 546–589.
- Burroughs, J.P., & T.A. Dudek. 2008. Deer-resistant plants for homeowners. *Extension Bulletin E-3042*, Michigan State University. 4 pages.

168

- Burrows, G.E., & R.J. Tyrl. 2001. Toxic plants of North America. Iowa State Univ Press, Ames. [More recent listings based partly on this original work are available; e.g., http://www.cnr.uidaho.edu/range/toxicplants_horses/Toxic%20Plant%20Database.html; <http://www.ces.ncsu.edu/depts/hort/consumer/poison/Euonysp.htm>; <http://www.accessdata.fda.gov/scripts/plantox/index.cfm>.]
- Cabiddu, A., M. Decandia, M. Sitzia & G. Molle. 2000. A note on the chemical composition and tannin content of some Mediterranean shrubs browsed by Sarda goats. *Cahiers Options Méditerranéennes* 52: 175–178
- **Cai Y-L. 1999. Fundamental characteristics of vines in Tiantong National Forest Park of Zhejiang Province, China. *Journal of East China Normal University (Nat. Sci.)* 1999-02.
- **Cai Y-L. & Song Y-C. 2000. Diversity of vines in subtropical zone of east China. *Plant Science Journal* 18: 390–396
- **Cai Y-L. & Song Y-C. 2005. Adaptive ecology of lianas in an evergreen broad-leaved forest of Tiantong National Forest Park, Zhejiang. II. Climbing capacity and climbing efficiency. *Acta Phytocologica Sinica* 2005-03.
- Cameron, K.M., & Fu C-X.. 2006. A nuclear rDNA phylogeny of *Smilax* (Smilacaceae). Pages 598–605 in: J.T. Columbus, E.A. Friar, J.M. Porter, L.M. Prince & M.G. Simpson (eds). *Monocots: Comparative Biology and Evolution. Excluding Poales*. Rancho Santa Ana Botanical Garden, Claremont, California [Aliso 22: 598–605.]
- Campbell, J.J.N. 1980. Present and presettlement forest conditions in the Inner Bluegrass of Kentucky. Ph.D. dissertation, University of Kentucky. 209 pages.
- Campbell, J.J.N. 1982. Pears and persimmons: a comparison of temperate forests of Europe and eastern North America. *Vegetatio* 49: 85–101.

169

- Campbell, J.J.N. 1987. Gradients of species composition in the Central Hardwood Forest. Pages 325–346 in: R.L. Hay, F.W. Woods & H. DeSelm (eds.). *Proceedings of the Sixth Central Hardwood Forest Conference*. University of Tennessee, Knoxville, Tennessee.
- Campbell, J.J.N. 1989. Historical evidence of forest composition in the Bluegrass region of Kentucky. Pages 231–246 in: G. Rink & C. A. Budelsky (eds.). *Proceedings of the Seventh Central Hardwoods Forest Conference*. Southern Illinois University, Carbondale, Illinois.
- Campbell, J.J.N., D.D. Taylor, M.E. Medley & A.C. Risk. 1991. Floristic and historical evidence of fire-maintained, grassy pine-oak “barrens” before settlement in southeastern Kentucky. Pages 359–375 in: S.C. Nodvin and T.A. Waldrop (eds.). *Fire and the Environment: Ecological and Cultural Perspectives*. Southeastern Forest Experiment Station, Asheville, North Carolina.
- Campbell, J.J.N. 2004a. Comparative ecology of warm-season (C4) versus cool-season (C3) grass species in Kentucky, with special reference to Bluegrass Woodlands. Pages 96–115 in: T.G. Barnes (ed). *Proceedings of the Fourth Eastern Native Grass Symposium*. University of Kentucky, Lexington, Kentucky.
- Campbell, J.J.N. 2004b. Invasive plant reduction in Bluegrass Woodlands; establishment of program in Lexington (Kentucky) area, focused on critical species and effective demonstration sites. Proposal to National Fish & Wildlife Foundation (Pulling Together Initiative), funded and transferred to University of Kentucky (Department of Forestry).
- Campbell, J.J.N. 2012. The herbivore hypothesis for Bluegrass Woodland. Published by the author at http://bluegrasswoodland.com/uploads/Herbivore_Hypothesis.pdf. 6 pages.
- Campbell, J.J.N., D.D. Taylor, M.E. Medley, & A.C. Risk. 1991. Floristic and historical evidence of fire-maintained, grassy pine-oak “barrens” before settlement in southeastern Kentucky. Pages 359–375 in: S.C. Nodvin and T.A. Waldrop (eds.). *Fire and the*

170

- Environment: Ecological and Cultural Perspectives. Southeastern Forest Experiment Station, Asheville, North Carolina.
- Campbell, J.J.N., & J. Grubbs. 1992. Natural plant communities of Hopkins County, Kentucky. *Transactions of the Kentucky Academy of Science* 53: 29–38.
- Campbell, J.J.N., & W.R. Seymour. 2011. A review of native vegetation types in the Black Belt of Mississippi and Alabama, with suggested relationships to the catenas of soil series. *Journal of the Mississippi Academy of Science* 56: 166–184.
- Campbell, J.J.N., & M.E. Medley. 2012. Atlas of the Vascular Plants of Kentucky. Current draft available at http://bluegrasswoodland.com/Kentucky_Plants_Flora.html. 3+ vols.
- Carter, G.A., & A.H. Teramura. 1988. Vine photosynthesis and relationships to climbing mechanics in a forest understory. *American Journal of Botany* 75: 1011–1018.
- Carter, G.A., A.H. Teramura & I.N. Forseth 1989. Photosynthesis in an open field for exotic versus native vines of the southeastern United States. *Canadian Journal of Botany* 67: 443–446.
- Castagneri, D., M. Garbarino & N. Nola. 2013. Host preference and growth patterns of ivy (*Hedera helix* L.) in a temperate alluvial forest. *Plant Ecology* 214: 1-9.
- Castleberry, S.B., W.M. Ford, K. V. Miller & P. Winston. 1999. White-tailed deer browse preferences in a Southern Bottomland Hardwood Forest. *Southern Journal of Applied Forestry* 23: 78–82.
- Cavender-Bares, J., & N.M. Holbrook. 2001. Hydraulic properties and freezing-induced xylem cavitation in evergreen and deciduous oaks with contrasting habitats. *Plant, Cell & Environment* 24: 1243–1256
- Challinor, V.L., P.G. Parsons, S. Chap, E.F. White, J.T. Blanchfield, R.P. Lehmann & J.J. DeVoss. 2012. Steroidal saponins from the roots of *Smilax* sp.: structure and bioactivity. *Steroids* 77: 504–511.

171

- **Chang F-R., Lee Y-H., Yang Y-L., Hsieh P-W., A.T. Khalil, Chen C-Y. & Wu Y-C. 2003. Secoiridoid glycoside and alkaloid constituents of *Hydrangea chinensis*. *Journal Natural Products* 66: 1245–1248
- Chaovanalikit, A., M.M. Thompson & R.E. Wrolstad. 2004. Characterization and quantification of anthocyanins and polyphenolics in blue honeysuckle (*Lonicera caerulea* L.). *Journal of Agricultural and Food Chemistry* 52: 848–852.
- Chat, J., B. Jáuregui, R.J. Petit & S. Nadot. 2004. Reticulate evolution in kiwifruit (*Actinidia*, Actinidiaceae) identified by comparing their maternal and paternal phylogenies. *American Journal of Botany* 91: 736–747.
- Cheeke, P.R. 1971. Nutritional and physiological implications of saponins: a review. *Canadian Journal Animal Science* 51: 621–632.
- **Chen K. & Wang X-D. 2008. Physiological indices of three liana species (*Mucuna sempervirens*, *Hedera nepalensis* var. *sinensis* and *Euonymus fortunei*) with drought resistance. *Journal of Zhejiang Forestry College* 2008-03.
- Chen S-C., Qiu Y-X., Wang A-L. & Fu C-X. 2006. A phylogenetic analysis of the Smilacaceae based on morphological data. *Acta Phytotaxonomica Sinica* 14: 113–125.
- **Chen Z-M., Li Q-R. & Du G-J. 2007. Resistance and absorbency to gaseous SO₂ of 42 landscaping plants in Zhejiang. *Journal of Zhejiang Forestry Science and Technology* 2007-06.
- Chen, X.-S. & Li, Q.-J. 2008a. Sexual systems and ecological correlates in an azonal tropical forests, SW China. *Biotropica* 40: 160–167.
- **Chen, X.-S. & Li, Q.-J. 2008b. Patterns of plant sexual systems in subtropical evergreen broad-leaved forests in Ailao Mountains, SW China. *Journal of Plant Ecology* 1: 179–185.

172

- **Chettri, A., S.K. Barik, H.N. Pandey & M.K. Lyngdoh. 2010. Liana diversity and abundance as related to microenvironment in three forest types located in different elevational ranges of the Eastern Himalaya. *Plant Ecology and Diversity* 3: 175–185.
- Chiu, S.T., & F.W. Ewers. 1992. Xylem structure and water transport in a twiner, a scrambler, and a shrub of *Lonicera* (Caprifoliaceae). *Trees* 6: 216–224.
- Choudhury, S., S. Datta, A.D. Talukdar & M.D. Choudhury. 2011. Phytochemistry of the family Bignoniaceae—a review. *Biological and Environmental Sciences* 7: 145–150.
- **Christenhusz, M.J.M. 2012. An overview of Lardizabalaceae. *Curtis's Botanical Magazine* 29: 235–276.
- Christensen, E.M. 1963. Herbaceous vegetation in lowland winter habitats of white-tailed deer in northern Wisconsin. *Ecology* 44: 411–414.
- Chumbalov, T.K., O.V. Fadeeva & E. I. Chesnokova. 1978. Polyphenols of *Lonicera microphylla*. *Chemistry of Natural Compounds* 14: 446–447.
- Chytrý, M., L. Tichý, & J. Roleček. 2003. Local and regional patterns of species richness in Central European vegetation types along the pH/calcium gradient. *Folia Geobotanica* 38: 429–442.
- Cipollini, D., R. Stevenson, S. Enright, A. Eyles & P. Bonello. 2008. Phenolic metabolites in leaves of the invasive shrub, *Lonicera maackii*, and their potential phytotoxic and anti-herbivore effects. *Journal of Chemical Ecology* 34: 144–152.
- Clark, M.S., & S.H. Gage. 1996. Effects of free-range chickens and geese on insect pests and weeds in an agroecosystem. *American Journal of Alternative Agriculture* 11: 39–47.
- Clarke, M.M., S.H. Reichard & C.W. Hamilton. 2006. Prevalence of different horticultural taxa of ivy (*Hedera* spp., Araliaceae) in invading populations. *Biological Invasions* 8: 149–157
- Clemants, S.E., & G. Moore. 2005. The changing flora of the New York metropolitan region. *Urban Habitats* 3 (1) ISSN 1541-7115. 19 pages.

173

- *Clinton, B.D., J.M. Vose & W.T. Swank. 1993. Site preparation burning to improve southern Appalachian pine-hardwood stands: vegetation composition and diversity of 13-year-old stands. *Canadian Journal of Forest Research* 23: 2271–2277.
- Cobb A.R., B. Choat & N.M. Holbrook. 2007. Dynamics of freeze–thaw embolism in *Smilax rotundifolia* (Smilacaceae). *American Journal of Botany* 94:640–649.
- Collins, B.S. & G.R. Wein. 1993. Understory vines: distribution and relation to environment on a southern mixed hardwood site. *Bulletin of the Torrey Botanical Club* 120: 38–44.
- Compton, P., J. Hedtcke & J. Harrington. 2003. Integrating livestock production and conservation: use of cattle in oak savanna restoration. University of Wisconsin Extension, Environmental Resources Center. 20 pages.
- Condon, J.M. 1991. Aspects of comparative vegetative morphology as an aid to *Actinidia* taxonomy. Ph.D. thesis, University of Canterbury, New Zealand. 241 pages.
- Conklin-Brittain, N.L., E.S. Dierenfeld, R.W. Wrangham, M. Norconk & S.C. Silver. 1999. Chemical protein analysis: a comparison of Kjeldahl crude protein and total ninhydrin protein using wild, tropical vegetation. *Journal of Chemical Ecology* 25: 2601–2622.
- Conn, J.S., T.R. Wentworth & U. Blum. 1980. Patterns of dioecism in the flora of the Carolinas. *American Midland Naturalist* 103: 310–315.
- Conover, M.R., & G.S. Kania. 1988. Browsing preference of white-tailed deer for different ornamental species. *Wildlife Society Bulletin* 16: 175–179.
- Conover, M.R., & G.S. Kania. 1995. Annual variation in white-tailed deer damage in commercial nurseries. *Agricultural, Ecosystems and Environment* 55: 213–217.
- Corley, R.N., A. Woldeghebriel & M.R. Murphy. 1997. Evaluation of the nutritive value of kudzu (*Pueraria lobata*) as a feed for ruminants. *Animal Feed Science and Technology* 68: 183–188.

174

- Cornelissen, J.H.C., N. Pérez-Harguindeguy, S. Díaz, J.P. Grime, B. Marzano, M. Cabido, F. Vendramini & B. Cerabolini. 1999. Leaf structure and defence control litter decomposition rate across species and life forms in regional floras on two continents. *New Phytologist* 143: 191–200.
- Cornett, M.W., L.E. Frelich, C. Puettmann & P.B. Reich. 2000. Conservation implications of browsing by *Odocoileus virginianus* in remnant upland *Thuja occidentalis* forests. *Biological Conservation* 93: 359–369.
- Cotton, A., & L.K. Darbaker. 1933. The laxative properties of *Rhamnus alnifolia*, a preliminary report. *Journal of the American Pharmaceutical Association* 22: 131.
- **Cramer, C. 1991. Cattle and woodlots can mix: livestock, native vegetation and wildlife all benefit with careful management. *The New Farm* 13: 40–41.
- Crawford, H.S., R.A. Lautenschlager, M.R. Stokes & T.L. Stone. 1993. Effects of forest disturbance and soil depth on digestible energy for moose and white-tailed deer. USDA Forest Service Research Paper NE-682. 13 pages.
- Crimmins, S.M., J.W. Edwards, W.M. Ford, P.D. Keyser & J.M. Crum. 2010. Browsing patterns of white-tailed deer following increased timber harvest and a decline in population density. *International Journal of Forestry Research* 592034. 7 pages.
- Crowch, C.M. & E.J. Okello. 2009. Kinetics of acetylcholinesterase inhibitory activities by aqueous extracts of *Acacia nilotica* (L.) and *Rhamnus prinoides* (L’Hér.). *African Journal of Pharmacy and Pharmacology* 3: 469–475.
- Crowley, P.H. 2002. Bluegrass savanna-woodland restoration. Grant application from University of Kentucky to the Kentucky Heritage Land Conservation Fund. Energy and Environment Cabinet, Kentucky State Government, Frankfort.

175

- Daly, C., M.P. Widrlechner, M.D. Halbleib, J.I. Smith & W.P. Gibson. 2012. Development of a new USDA Plant Hardiness Zone Map for the United States. *Journal of Applied Meteorology and Climatology* 51: 242–264.
- Dambach, C.A. 1944a. A ten-year ecological study of adjoining grazed and ungrazed woodlands in northeastern Ohio. *Ecological Monographs* 255–270.
- Dambach, C.A. 1944b. Comparative productiveness of adjacent grazed and ungrazed sugar-maple woods. *Journal of Forestry* 42: 164–168.
- D’Antuono, L.F., & A. Lovato. 2003. Germination trials and domestication potential of three native species with edible sprouts: *Ruscus aculeatus* L., *Tamus communis* L. and *Smilax aspera* L. In: E. Düzyaman & Y. Tüzel (eds). *Proceedings of the International Symposium on Sustainable Use of Plant Biodiversity*. *Acta Horticulturalae* 598: 211–218.
- Darabant, A., P.B. Rai, K. Kenzin, W. Roder, G. Gratzner. 2007. Cattle grazing facilitates tree regeneration in a conifer forest with palatable bamboo understory. *Forest Ecology and Management* 252: 73–83.
- Darcy, A.J., & M.C. Burkhart. 2002. Allelopathic potential of *Vinca minor*, an invasive exotic plant in west Michigan forests. *Bios* 73: 127–132.
- Darnley-Gibbs, R. 1974. *Chemotaxonomy of Flowering Plants*. 4 vols. McGill-Queen’s University Press, Montreal, Canada.
- Davis, J.D., & R.F. Evert. 1970. Seasonal cycle of phloem development in woody vines. *Botanical Gazette* 131: 128–138.
- Davis, M.B. 1983. Quaternary history of deciduous forests of eastern North America and Europe. *Annals of the Missouri Botanical Garden* 70: 550–563.
- Dawson, T.E., & J.R. Ehleringer. 1993. Gender-specific physiology, carbon isotope discrimination, and habitat distribution in boxelder, *Acer negundo*. *Ecology* 74, 798–815.

176

- Decandia, M., M. Sitzia, A. Cabiddu, D. Kababya & G. Molle. 2000. The use of polyethylene glycol to reduce the anti-nutritional effects of tannins in goats fed woody species. *Small Ruminant Research* 38: 157–164
- DeGaetano, A.T., & M.D. Schulman. 1990. A climatic classification of plant hardiness in the United States and Canada. *Agricultural and Forest Meteorology* 51: 333–351.
- Deguchi, Y., S. Sato & K. Sugawara. 2004. Relationship between some chemical components of herbage, dietary preference and fresh herbage intake rate of Japanese serow. *Applied Animal Behaviour Science* 73: 69–79.
- Denk, T., & Oh I-C. 2005. Phylogeny of Schisandraceae based on morphological data: evidence from modern plants and the fossil record. *Plant Systematics and Evolution* 256: 113–145.
- Dennis, A. 1997. Effects of livestock grazing on forest habitats. Pages 313–341 in: M.W. Schwartz (ed). *Conservation in Highly Fragmented Landscapes*. Chapman & Hall, New York.
- DeVore, M.L., & K.B. Pigg. 2007. A brief review of the fossil history of the family Rosaceae with a focus on the Eocene Okanogan Highlands of eastern Washington State, USA, and British Columbia, Canada. *Plant Systematics and Evolution* 266: 45–57.
- DeVos, A., 1964. Food utilization of snowshoe hares on Manitoulin Island, Ontario. *Journal of Forestry* 62: 238–244.
- DeWitt, J.B., & J.V. Derby. 1955. Changes in nutritive value of browse plants following forest fires. *Journal of Wildlife Management* 19: 65–70.
- Dilcher D.L., & T.A. Lott. 2005. A middle Eocene fossil plant assemblage (Powers Clay Pit) from western Tennessee. *Bulletin of the Florida Museum of Natural History* 45: 1–43.

- Dillard, J., S. Jester, J. Baccus, R. Simpson & L. Poor. 2005. White-tailed deer food habits and preferences in the Cross Timbers and Prairies regions of Texas. Texas Parks and Wildlife Department, PWD RP W7000-1017 (03/06).
- Dillenberg, L.R., D.F. Whigham, A.H. Teramura & I.N. Forseth. 1993. Effects of belowground and aboveground competition from the vines *Lonicera japonica* and *Parthenocissus quinquefolia* on the growth of the tree host *Liquidambar styraciflua*. *Oecologia* 93: 48–54.
- Ding J-Q., R. Reardon, Wu Y., Zheng H. & Fu W-D. 2006. Biological control of invasive plants through collaboration between China and the United States of America: a perspective. *Biological Invasions* 8: 1439–1450.
- Dirr, M.A. 1998. *Manual of Woody Landscape Plants. Their Identification, Ornamental Characteristics, Culture, Propagation and Uses*. 5th edition. Stipes Publishing, Champaign, Illinois.
- Donoghue, M.J., & S.A. Smith. 2004. Patterns in the assembly of temperate forests around the Northern Hemisphere. *Philosophical transactions of the Royal Society of London Series B—Biological Sciences* 359: 1633–1644.
- Driemeier, D. 1997. Marijuana (*Cannabis sativa*) toxicosis in cattle. *Veterinary and Human Toxicology* 39: 351–352.
- Duncan, W.H. 1967. Woody vines of the Southeastern States. *Sida* 3: 1–76.
- *Dyess, J.G., M.K. Causey, H.L. Stribling & B.G. Lockaby. 1993. Effects of fertilization on production and quality of Japanese honeysuckle deer browse. *Highlights of Agricultural Research* 40: 9.
- Ehrman, T.M., D.J. Barlow & P.J. Hyland. 2007. Phytochemical databases of Chinese herbal constituents and bioactive plant compounds with known target specificities. *Journal of Chemical Information and Modeling* 47: 254–263.

- *Eich, E. 2008. Solanaceae and Convolvulaceae: Secondary Metabolites. Biosynthesis, Chemotaxonomy, Biological and Economic Significance (A Handbook). Springer, Berlin. 625+ pages.
- Ellenberg, H. 1988. Vegetation Ecology of Central Europe. 4th edition. Cambridge University Press, Cambridge. 731 pages.
- Enari, H., & H. Sakamaki. 2010. Abundance and morphology of Japanese mulberry trees in response to the distribution of Japanese macaques in snowy areas. *International Journal of Primatology* 31: 904–919.
- Epanchin-Niell, R.S., & A. Hastings. 2010. Controlling established invaders: integrating economics and spread dynamics to determine optimal management. *Ecology Letters* 13: 528–541.
- Ertsen, A.C.D., J.R.M. Alkemade, & M.J. Wassen. 1998. Calibrating Ellenberg indicator values for moisture, acidity, nutrient availability and salinity in the Netherlands. *Plant Ecology* 135: 113–124.
- Escudero, A., & S. Mediavilla. 2003. Decline in photosynthetic nitrogen use efficiency with leaf age and nitrogen resorption as determinants of leaf life span. *Journal of Ecology* 91: 880–889.
- Ewald, J. 2003. The sensitivity of Ellenberg indicator values to the completeness of vegetation releves. *Basic and Applied Ecology* 4: 507–513.
- Eschtruth, A.K., & J.J. Battles. 2009. Assessing the relative importance of disturbance, herbivory, diversity, and propagule pressure in exotic plant invasion. *Ecological Monographs* 79: 265–280.
- Evens, Z.N., & S.J. Stellpflug. 2012. Holiday plants with toxic misconceptions. *Western Journal of Emergency Medicine* DOI: 10.5811/westjem.2012.8.12572. 5 pages.

179

- Fan P-F., Ni Q-Y., Sun G-H., Huang B. & Jiang X-L. 2009. Gibbons under seasonal stress: the diet of the black crested gibbon (*Nomascus concolor*) on Mt. Wuliang, Central Yunnan, China. *Primates* 50: 37–44.
- **Fang J-Y., Wang Z-H., & Tang Z-Y. 2011. Atlas of Woody Plants in China: Distribution and Climate. Springer, Heidelberg, Germany. Vol. I. 1368 pages.
- Fargione, M.J., P.D. Curtis, & M.A. Richmond 1991. Resistance of woody ornamental plants to deer damage. Cornell Cooperative Extension Fact Sheet 800.00, Cornell University, Ithaca, New York. 4 pages.
- Feeny, P. 1976. Plant apparency and chemical defense. Pages 1–40 in: J.W. Wallace & R.L. Mansell (eds). *Recent Advances in Phytochemistry*, Vol. 10. Plenum Press, New York.
- **Feng W-S., Hao Z-Y. & Zheng X-K. 2007. Chemical constituents of genus *Celastrus*. *Chinese Journal of New Drugs* 2007-09.
- Feng, Y., Oh S-H., & P.S. Manos. 2005. Phylogeny and historical biogeography of the genus *Platanus* as inferred from nuclear and chloroplast DNA. *Systematic Botany* 30: 786–799.
- Finnoff, D., A. Strong & J. Tschirhart J. 2008. A bioeconomic model of cattle stocking on rangeland threatened by invasive plants and nitrogen deposition. *American Journal of Agricultural Economics* 90: 1074–1090.
- Fisher J. 1975. Cannabis in Nepal: an overview. Pages 247–256 in: Rubin Vera (ed). *Cannabis and Culture*. Mouton Publishers, The Hague, Belgium.
- Fisher, R.F., & D. Binkley. 2000. *Ecology and Management of Forest Soils*. 3rd edition. John Wiley & Sons, New York. 489 pages.
- Fischer, A., & U. Feller. 1994. Seasonal changes in the pattern of assimilatory enzymes and the proteolytic activities in leaves of juvenile ivy. *Annals of Botany* 74: 389–396.
- Flamini, G., A. Braca, P.J. Cioni & I. Morelli. 1997. Three new flavonoids and other constituents from *Lonicera implexa*. *Journal of Natural Products* 60: 449–452.

180

- Flora Europaea Editorial Committee [T.G. Tutin et al.]. 1964–80. Flora Europaea. 5 vols. Cambridge University Press, Cambridge, England.
- Flora of China Editorial Committee [Z. Wu, P. Raven et al.]. 1996–2011 [continuing]. Flora of China. 25 vols. Science Press and Missouri Botanical Garden, St. Louis, Missouri [see also <http://hua.huh.harvard.edu/china/>].
- Flora of North America Editorial Committee. 1993–2010 [continuing]. Flora of North America north of Mexico. 30 vols. Oxford University Press, New York.
- Flory, S.L., & J. Lewis. 2009. Nonchemical methods for managing Japanese Stiltgrass (*Microstegium vimineum*). Invasive Plant Science and Management 2: 301–308.
- Forest, F., V. Savolainen, M.W. Chase, R. Lupia, A. Bruneau & P.R. Crane. 2005. Teasing apart molecular- versus fossil-based error estimates when dating phylogenetic trees: a case study in the birch family (Betulaceae). Systematic Botany 30: 118–133.
- Forrester, J.A., D.J. Leopold, & H.B. Underwood. 2006. Isolating the effects of white-tailed deer on the vegetation dynamics of a rare maritime American holly forest. American Midland Naturalist 156: 136–150.
- Forseth, I.N., A.H. Teramura. 1987. Field photosynthesis, microclimate and water relations of an exotic temperate liana, *Pueraria lobata*, kudzu. Oecologia 71: 262–267
- Fowler, A., & R.E. Kissell. 2007. Relative abundance and habitat associations of swamp rabbits in eastern Arkansas. Southeastern Naturalist 6: 247–258.
- Francis, G., Z. Kerem, H.P.S. Makkar & K. Becker. 2002. The biological action of saponins in animal systems: a review. British Journal of Nutrition 88: 587–605.
- Frankel, D.H. 1999. Field guide to clinical dermatology. Lippincott, Williams & Wilkins, Philadelphia.

- Frasier, C.L. 2008. Evolution and systematics of the angiosperm order Gentianales with an in-depth focus on Loganiaceae and its species-rich and toxic genus *Strychnos*. Ph.D. dissertation, Rutgers University, New Brunswick, New Jersey. 124 pages.
- Frederick, R.D., & M.L. Kennedy. 1995. Use of fertilized honeysuckle by white-tailed deer in western Tennessee. Proceedings of the Annual Conference of the Southeastern Association of Fish and Wildlife Agencies 49: 383–388.
- Frye, M.J., J. Hough-Goldstein & J. Sun. 2007. Biology and preliminary host range assessment of two potential kudzu biological control agents. Environmental Entomology 36: 1430–1440.
- Frye, M.J., J. Hough-Goldstein & K.A. Kidd. 2012. Response of kudzu (*Pueraria montana* var. *lobata*) seedlings and naturalized plants to simulated herbivory. Invasive Plant Science and Management 5: 417–426.
- García, R.R., R. Celaya, U. García & K. Osoro. 2012. Goat grazing, its interactions with other herbivores and biodiversity conservation issues. Small Ruminant Research 107: 49–64.
- García, R.R., D. Fraser, R. Celaya, L.M.M. Ferreira, U. García, K. Osoro. 2013. Grazing land management and biodiversity in the Atlantic European heathlands: a review. Agroforestry Systems 87: 19–43.
- **Garfi, G., & S. Ficcarotta. 2003. Influence of ivy (*Hedera helix* L.) on the growth of downy oak (*Quercus pubescens* s.l.) in the Monte Carcaci Nature Reserve (central-western Sicily). Ecologia Mediterranea 29: 5–14.
- Garin, I., A. Aldezabal, J. Herrero & A. García-Serrano. 2000. Understorey foraging and habitat selection by sheep in mixed Atlantic woodland. Journal of Vegetation Science 11: 863–870.
- Geyer, W.A., & F. Ponder. 2012. Transferring site information for black walnut from native woodlands in southeastern Kansas USA to identify sites for agroforestry practices. Agroforestry Systems, in press.

- Ghent, A.W. 1972. A method for exact testing of 2X2, 2X3, 3X3, and other contingency tables, employing binomial coefficients. *American Midland Naturalist* 88: 15-27.
[Calculation available at <http://www.danielsoper.com/statcalc3/>.]
- Gianoli, E. 2004. Evolution of a climbing habit promotes diversification in flowering plants. *Proceeding of the Royal Society of London, Series B* 271: 2011–2015.
- Gill, R.M.A. 1992. A review of damage by mammals in north temperate forests: 1. Deer. *Forestry* 65: 145–69.
- Gill, R.M.A., & V. Beardall. 2001. The impact of deer on woodlands: the effects of browsing and seed dispersal on vegetation structure and composition. *Forestry* 74: 209–218.
- Gillespie, T.W. 1999. Life history characteristics and rarity of woody plants in tropical dry forest fragments of Central America. *Journal of Tropical Ecology* 15: 637–649.
- Givnish, T.J., 1987. Comparative studies of leaf form—assessing the relative roles of selective pressures and phylogenetic constraints. *New Phytologist* 106: 131–160.
- Givnish, T.J. 1990. Leaf mottling: relation to growth form and leaf phenology and possible role as camouflage. *Functional Ecology*. 4: 463–474.
- Givnish, T.J. 2002. Adaptive significance of evergreen vs. deciduous leaves: solving the triple paradox. *Silva Fennica* 36: 703–743.
- Givnish, T.J., & G.J. Vermeij. 1976. Sizes and shapes of liane leaves. *American Naturalist* 100: 743–778.
- Glasby, J.S. 1992. *Dictionary of Plants Containing Secondary Metabolites*. Günter Adam. CRC Press. 488 pages.
- Glennie, C.W. 1969. A comparative phytochemical study of Caprifoliaceae. Ph.D. dissertation, University of British Columbia, Vancouver. 123 pages.
- Glitzenstein, J.S., W.J. Platt & D.R. Streng. 1995. The effects of fire regime and habitat on tree dynamics in North Florida longleaf pine savannas. *Ecological Monographs* 65: 441–76.

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- Go, M. 2010. Seasonal changes in food resource distribution and feeding sites selected by Japanese macaques on Koshima Islet, Japan. *Primates* 51: 149–158.
- Godwin, H. 1943. Biological Flora of the British Isles: *Rhamnaceae*. *Journal of Ecology* 31: 66–92
- Goebel, P.C., B.J. Palik, L.K. Kirkman, M.B. Drew, L. West & D.C. Pederson. 2001. Forest ecosystems of a lower Gulf Coastal Plain Landscape: multifactor classification and analysis. *Journal of the Torrey Botanical Society* 128: 47–75.
- Goldstein, S.J., & A. F. Richard. 1989. Ecology of rhesus macaques (*Macaca mulatta*) in Northwest Pakistan. *International Journal of Primatology* 10: 531–567.
- González-Hernández, M.P., & F.J. Silva-Pando. 1996. Grazing effects of ungulates in a Galician oak forest (northwest Spain). *Forest Ecology and Management* 88: 65–70
- González-Hernández, M.P., & F.J. Silva-Pando. 1999. Nutritional attributes of understory plants known as components of deer diets. *Journal of Range Management* 52: 132–138.
- González-Hernández, M.P., J. Karchesy & E.E. Starkey. 2003. Research observation: hydrolyzable and condensed tannins in plants of northwest Spain forests. *Journal of Range Management* 56: 461–465.
- Grabner, K.W., M. Struckhoff & D.A. Buhl. 2005. Evaluating the impacts of white-tailed deer (*Odocoileus virginianus*) on vegetation within Pea Ridge National Military Park. U.S. Geological Survey, Northern Prairie Wildlife Research Center, Jamestown, North Dakota. 58 pages.
- Graham, A. 1999. *Late Cretaceous and Cenozoic History of North American Vegetation*. Oxford University Press, New York. 370 pages.
- Graves, G. 1940. Fruiting forms and variegations in the wintercreeper. *Journal of the New York Botanical Garden* 41: 285-287.

184

- Green, A.F., T.S. Ramsey & J. Ramsey. 2011. Phylogeny and biogeography of ivies (*Hedera* spp., Araliaceae), a polyploid complex of woody vines. *Systematic Botany* 36: 1114–1127.
- Greller, A.M. 1989. Correlation of warmth and temperateness with the distributional limits of zonal forests in eastern North America. *Bulletin of the Torrey Botanical Club* 116: 145–163.
- Greller, A.M. 2003. A review of the temperate broad-leaved evergreen forest zone of southeastern North America: floristic affinities and arborescent vegetation types. *Botanical Review* 69: 269–299.
- Griggs, J.A., J.H. Rock, C.R. Webster, M.A. Jenkins. 2006. Vegetative legacy of a protected deer heard in Cades Cove, Great Smokey Mountains National Park. *Natural Areas Journal* 26: 126–136.
- Grubb, P.J. 1977. The maintenance of species-richness in plant communities: the importance of the regeneration niche. *Biological Reviews of the Cambridge Philosophical Society* 52: 107–145.
- Grubb, P.J. 1992. A positive distrust in simplicity—lessons from plant defences and from competition among plants and among animals. *Journal of Ecology* 80: 585–610.
- Grubb, P.J. 2002. Leaf form and function—towards a radical new approach. *New Phytologist* 155: 317–320.
- Grubb, P.J., & P.L. Marks. 1989. Spring flowers of eastern North America and European deciduous forests. *Plants Today* 2: 89–95.
- Grueter, C.C., Li D-Y., Ren B-P., Wei F-W. & C. P. van Schaik. 2009. Dietary profile of *Rhinopithecus bieti* and its socioecological implications. *International Journal of Primatology* 30: 601–624.
- *Guo W-H., Gu Z-L., Wei K-P., Liu Y-J. & Wang Z-L. 2009. Effect of *Humulus scandens* for traditional fiber sources on digestion, diarrhea, and performance of growing rabbits. *Agricultural Sciences in China* 8: 497–501.

185

- *Guvenalp, Z., H. Ozbek, A. Kuruuzum-Uz, C. Kazaz, L.O. Demirezer. 2012. Chemical constituents of *Lonicera etrusca*. *Chemistry of Natural Compounds* 48: 693–695.
- Habeck, J.R., 1960. Winter deer activity in the white cedar swamps of northern Wisconsin. *Ecology* 41: 327–333.
- Hall, W., & L. Degenhardt. 2009. Adverse health effects of non-medical cannabis use. *Lancet* 374: 1383–1391.
- Halls, L.K. 1975. Browse use by deer in an east Texas forest. *Proceedings of the Annual Conference of Southeastern Association of Game and Fish Commissioners (1974)* 28: 557–562.
- Halls, L.K., & C.E. Boyd. 1982. Influence of managed pine stands and mixed pine/hardwood standson well-being of deer. U.S.D.A. Forest Service, Southern Forest Experiment Station. Publication SO-183. 21 pages.
- Hampton, R., E. Small & A. Haunold. 2009. Habitat and variability of *Humulus lupulus* var. *lupuloides* in upper midwestern North America: A critical source of American hop germplasm. *Journal of the Torrey Botanical Society* 128: 35–46.
- Han, L., Xie L-J., Dai K-J., Yang Q. & Cai Z-Q. 2010. Contrasting leaf characteristics of trees and lianas in secondary and mature forests in southwestern China. *Photosynthetica* 48: 559–566.
- Handley, C.O. 1945. Japanese honeysuckle in wildlife management. *Journal of Wildlife Management* 9: 261–264.
- Hao, D-C., Gu X-J., Xiao P-G. & Peng Y. 2013. Chemical and biological research of *Clematis* medicinal resources. *Chinese Science Bulletin* 58: in press, doi 10.1007/s11434-012-5628-7.
- Harborne, J.B. 1991a. The chemical basis of plant defense. Pages 46–59 in: R.T. Pale & C.T. Robbins (eds.). *Plant Defenses against Mammalian Herbivory*. CRC Press, London.

186

- Harborne, J.B. 1991b. Recent advances in the ecological chemistry of plant terpenoids. Pages 399–426 in: J.B. Harborne and F.A. Tomes-Barberan (eds.). *Ecological Chemistry and Biochemistry of Plant Terpenoids*. Clarendon Press, Oxford.
- Harborne, J.B., H. Baxter, & G.P. Moss (eds). 1999. *Phytochemical Dictionary*, 2nd edition. Taylor and Francis, London. 976 pages.
- Hardt, R.A. 1986. Japanese honeysuckle: from “one of the best” to ruthless pest. *Arnoldia* 46: 27–34.
- Harlow, R.F., & R.L. Downing. 1970. Deer browsing and hardwood regeneration in the southern Appalachians. *Journal of Forestry* 68: 298–300.
- Harlow, R.F., J.B. Whelan, H.S. Crawford & J.E. Skeen. 1975. Deer foods during years of oak mast abundance and scarcity. *Journal of Wildlife Management* 39: 298–300.
- Harmer, R., G. Peterken, G. Kerr & P. Poulton. 2001. Vegetation changes during 100 years of development of two secondary woodlands on abandoned arable land. *Biological Conservation* 101: 291–304.
- Harrington, J.A., & E. Kathol. 2009. Responses of shrub midstory and herbaceous layers to managed grazing and fire in a North American savanna (oak woodland) and prairie landscape. *Restoration Ecology* 17: 234–244.
- Harrison, S.P., I.C. Prentice, D. Barboni, K.E. Kohfeld, J. Ni & J-P. Sutra. 2009. Ecophysiological and bioclimatic foundations for a global plant functional classification. *Journal of Vegetation Science* 21: 300–317.
- Hart, C.A., R.M. Batt, J.R. Saunders & B. Getty. 1988. Lectin-induced damage to the enterocyte brush border: an electron-microscopic study in rabbits. *Scandinavian Journal of Gastroenterology* 23: 1153–1159.
- Harry, G.B. 1957. Winter food habits of moose in Jackson Hole, Wyoming. *Journal of Wildlife Management* 21: 53–57.

187

- Haynes, G. 2012. Elephants (and extinct relatives) as earth-movers and ecosystem engineers. *Geomorphology* 157–158: 99–107.
- Hecht, S.B. 1979. Spontaneous legumes of developed pasture of the Amazon and their forage potential. Pages 65–78 in: P.A. Sanchez & L.E. Tergas (eds.). *Pasture production in acid soils of the tropics*. CIAT (International Center Trop. Agric.), Santiago de Cali, Columbia.
- Hedtcke, J., J. Posner, M. Rosemeyer & K. Albrecht. 2009. Browsing for conservation: springtime forage value of midstory shrubs of degraded oak savannas in southern Wisconsin. *Renewable Agriculture and Food Systems* 24: 293–299.
- Hegnauer, R., & M. Hegnauer. 2002. *Chemotaxonomy of Plants*. 13 vols. Birkhäuser, Basel.
- Hédli, R., M. Kopecký & J. Komárek. 2010. Half a century of succession in a temperate oakwood: from species-rich community to mesic forest. *Diversity and Distributions* 16: 267–276.
- Heilbuth, J.C. 2000. Lower species richness in dioecious clades. *American Naturalist* 156: 221–241.
- Heinrich, H., & S. Predl. 1995. Can we landscape to accommodate deer? The Tracy Estate Research Garden. Pages 102–112 in: *Proceedings of the Sixth Eastern Wildlife Damage Control Conference*. University of Nebraska Lincoln, School of Natural Resource Science.
- Hester, A.J., L. Edenius, R.M. Buttenschön & A.T. Kuiters. 2000. Interactions between forests and herbivores: the role of controlled grazing experiments. *Forestry* 73: 381–91
- Heuzé, P., J.L. Dupouey & A. Schnitzler. 2009. Radial growth responses of *Hedera helix* L. To hydrological changes and climatic variability in the Rhine floodplain. *River Research and Application* 25: 393–404.
- Hewitt, D.G., & R.L. Kirkpatrick. 1997. Ruffed grouse consumption and detoxification of evergreen leaves. *Journal of Wildlife Management* 61: 129–139.

188

- Hikosaka, K. 2004. Leaf canopy as a dynamic system. *Ecophysiology and optimality in leaf turnover*. *Annals of Botany* 95: 521–533.
- Hodder, K.H., P.C. Buckland, K.K. Kirby & J.M. Bullock. 2009a. Can the pre-neolithic provide suitable models for re-wilding the landscape in Britain? *British Wildlife* 20:4–15.
- Hodder, K.H. & J.M. Bullock. 2009b. Really wild? Naturalistic grazing in modern landscapes. *British Wildlife* 20:37–43.
- Hoppe, K.A., P.L. Koch. 2007. Reconstructing the migration patterns of late Pleistocene mammals from northern Florida, USA. *Quaternary Research* 68: 347–352.
- Hou, D. 1955. A revision of the genus *Celastrus*. *Annals of the Missouri Botanical Garden* 55: 205–302.
- Howard, W.J. 1937. Notes on winter foods of Michigan deer. *Journal of Mammalogy* 18: 77–80.
- Huang Z-P., Huo S., Yang S-G., Cui L-W. & Xiao W. 2010. Leaf choice by black-and-white snub-nosed monkeys *Rhinopithecus bieti* is related to physical and chemical properties. *Current Zoology* 56: 643–649.
- Hufford, L., M.L. Moody & D.E. Soltis. 2001. A phylogenetic analysis of Hydrangeaceae based on sequences of the plastid gene *matK* and their combination with *rbcL* and morphological data. *International Journal of Plant Sciences* 162: 835–846.
- Hughes, F.M.R. 2012. When is open-endedness desirable in restoration projects? *Restoration Ecology* 20: 291–295.
- Hulme, P. E. 1996. Herbivory, plant regeneration, and species coexistence. *Journal of Ecology* 84: 609–615.
- Ichihashi, R., H. Nagashima & M. Tateno. 2009. Morphological differentiation of current-year shoots of deciduous and evergreen lianas in temperate forests in Japan. *Ecological Research* 24: 393–403.

189

- Ichihashi, R., H. Nagashima & M. Tateno. 2010. Biomass allocation between extension- and leaf display-oriented shoots in relation to habitat differentiation among five deciduous liana species in a Japanese cool-temperate forest. *Plant Ecology* 211: 181–190.
- Ichihashi, R., & M. Tateno. 2011. Strategies to balance between light acquisition and the risk of falls of four temperate liana species: to overtop host canopies or not? *Journal of Ecology* 99: 1071–1080.
- Igota, H., M. Sakuragi, H. Uno, K. Kaji, M. Kaneko & R. Akamatsu. 2004. Seasonal migration patterns of female sika deer in eastern Hokkaido, Japan. *Ecological Research* 19: 169–78.
- Imai, K., K. Miura, H. Iida & K. Fujisaki. 2011. Herbivorous insect fauna of invasive vine, kudzu, *Pueraria montana* (Lour.) Merr. var. *lobata* (Willd.) Maesen et S. Almeida (Leguminosae), in its native range. *Japanese Journal of Applied Entomology and Zoology* 55: 147–154
- Ingham, C.S. 2008. Himalayan blackberry (*Rubus armeniacus*) and English ivy (*Hedera helix*) response to high intensity-short duration goat browsing. Ph.D. dissertation, Oregon State University. 126 pages.
- Invasive Plant Control Inc. 2008 [approx.]. Rock Creek National Park, DC. [Progress report posted at <http://www.invasiveplantcontrol.com/pdflinks/RCPNP.pdf>.]
- Ishih, A., T. Miyase, T. Suzuki, F.W. Muregi & M. Terada. 2007. Seasonal variation in the content of a febrifugine and isofebrifugine alkaloid mixture in aerial parts of *Hydrangea macrophylla* var. *otaksa*, with special reference to its antimalarial activity. *Journal of Natural Medicine* 61: 213–216.
- Isnard, S., & W.K. Silk. 2009. Moving with climbing plants from Charles Darwin's time into the 21st century. *American Journal Botany* 96: 1205–1221.
- Iwamoto, T. 1982. Food and nutritional condition of free ranging Japanese monkeys on Koshima Islet during winter. *Primates* 23: 153–170.

190

- Jacques, F.M.B., W. Wang, R.D.C. Ortiz, H.L. Li, Z.K. Zhou & Z.D. Chen. 2011. Integrating fossils in a molecular-based phylogeny and testing them as calibration points for divergence time estimates in Menispermaceae. *Journal of Systematics and Evolution* 49: 25–49.
- Jett, J.W. 1995. Resistance of ornamentals to deer damage. West Virginia University Extension Service Fact Sheet 655 (revised).
- **Jiang D., Xiao W., Ding P., Wu Y-H., Yang Y-H. & Sun X-G. 2007. A study on the adaptability of fourteen types of shade-tolerant plants in Lanzhou City. *Journal of Northwest Forestry University* 2007-02.
- *Jiang G.Y., Qian Z-M., Zhang T.-D. & Li P. 2008. Chemical constituents in aerial parts of *Lonicera chrysantha* Turcz. *Chemistry and Industry of Forest Products* 2008-06.
- Jing, S.W. & P.D. Coley. 1990. Dioecy and herbivory: the effect of growth rate on plant defense in *Acer negundo*. *Oikos* 58: 369–377.
- Johnson, E.A. 1952. Effects of farm woodland grazing on watershed values in the southern Appalachian Mountains. *Journal of Forestry* 50: 109–113.
- Johnson, C.N. 2009. Ecological consequences of Late Quaternary extinctions of megafauna. *Proceedings of the Royal Society Series B* 276: 2509–2519.
- Jones, S.M., D.H. Van Lear & S.K. Cox. 1984. A vegetation-landform classification of forest sites within the upper Coastal Plain of South Carolina. *Bulletin of the Torrey Botanical Club* 111: 349–360.
- Jones, P.D., S. Demarais, B.K. Strickland & S.L. Edwards. 2008. Soil region effects on white-tailed deer forage protein. *Southeastern Naturalist* 7: 595–606.
- JSPS [Japanese Society for Plant Systematics]. 2012. Flora of Japan [<http://foj.c.u-tokyo.ac.jp/gbif/>].
- Jull, J.G. 2001. Plants not favored by deer. University of Wisconsin Extension Publication A3727. 4 pages.

- Kajita, T., H. Ohashi, Y. Tateishi, C.D. Baile & J.J. Doyle. 2001. rbcL and legume phylogeny, with particular reference to Phaseoleae, Millettieae, and allies. *Systematic Botany* 26: 515–536.
- Karban, R. 2008. Leaf drop in evergreen *Ceanothus velutinus* as a means of reducing herbivory. *Ecology* 89: 2446–2452.
- Kartesz, J.T. 2012. BONAP's North American Plant Atlas (NAPA) [http://www.bonap.org/Map_Switchboard.html]. Biota of North America Program (BONAP), Chapel Hill, NC.
- Keeler, R.F., 1979. Toxins and teratogens of the Solanaceae and Liliaceae. Pages 59–82 in: A. D. Kinghorn (ed.). *Toxic Plants*. Columbia University Press, New York.
- Kemp, J.R., P.G. Kevan & U. Posluszny. 1993. Morphological differences and changes of the gynoecium in short-lived flowers of *Rosa setigera* Michaux and their relationship to dioecy. *International Journal of Plant Science* 154: 550–556.
- Kettenring, K.M., & C. Reinhardt-Adams. 2011. Lessons learned from invasive plant control experiments: a systematic review and meta-analysis. *Journal of Applied Ecology* 48, 970–979.
- Kidjo, N., G. Feracci, E. Bideau, G. Gonzalez, C. Mattéi, B. Marchand & S. Aulagnier. 2007. Extirpation and reintroduction of the Corsican red deer (*Cervus elaphus corsicanus*) in Corsica. *Oryx* 41: 488–494.
- Kienle, G.S., R. Grugel & H. Kiene. 2011. Safety of higher dosages of *Viscum album* L. in animals and humans—systematic review of immune changes and safety parameters. *BioMed Central Complementary and Alternative Medicine* DOI:10.1186/1472-6882-11-72.
- Kikuchi, M., & N. Matsuda. 1996. Flavone glycosides from *Lonicera gracilipes* var. *glandulosa*. *Journal of Natural Products* 59: 314–315.
- Kikuzawa, K., & M.J. Lechowicz. 2011. Ecology of Leaf Longevity. *Ecological Research Monographs*, Springer. 147 pages.

- Kim, C.S., D. Lee, G. Schaible & U. Vasavada. 2007. Multiregional invasive species management: theory and an application to Florida's exotic plants." *Journal of Agricultural and Applied Economics* 39: 111–124.
- Kim K-W., Suh S-J., Kim J-D., Kim S-S., Lee I-S., Kim J-K., Chang G-T., Kim D-S. & Kim C-H. 2009. Effects on lipid peroxidation and antioxidative enzymes of *Euonymus alatus* in cultured rat hepatocytes. *Basic & Clinical Pharmacology & Toxicology* 104: 60–70.
- Kirby, K.J. 2001. The impact of deer on the ground flora of British woodland. *Forestry* 74: 219–229.
- *Kitanaka, S., M. Takido, K. Mizoue & S. Nakaike. 1996. Cytotoxic cardenolides from woods of *Euonymus alata*. *Chemistry and Pharmacy Bulletin (Tokyo)* 44: 615–617.
- Klein, J., & D. Conover. 2010. Restoring biodiversity by lowering deer numbers at Shawnee Lookout (Ohio). *Ecological Restoration* 28: 131–133.
- Knapp, L.B., J.H. Fownes & R.A. Harrington. 2008. Variable effects of large mammal herbivory on three non-native versus three native woody plants. *Forest Ecology and Management* 255: 92–98.
- Knight, T.M., J.L. Dunn, L.A. Smith, J-A. Davis & S. Kalisz. 2009. Deer facilitate invasive plant success in a Pennsylvania forest understory. *Natural Areas Journal* 29: 110–116.
- Knox, G.W. 2007. French Hydrangea for gardens in north and central Florida. Publication ENH1069, Florida Cooperative Extension Service, University of Florida. 7 pages.
- Korschgen, L.J., W.R. Porath & O. Torgerson. 1980. Spring and summer foods of deer in the Missouri Ozarks. *Journal of Wildlife Management* 44: 89-97.
- **Koyama, K., & K. Kikuzawa. 2008. Intraspecific variation in leaf life span for the semi-evergreen liana *Akebia trifoliata* is caused by both seasonal and aseasonal factors in a temperate forest. *Journal of Ecology and Field Biology* 31: 207–211.

193

- Krafft, C.C., & J.S. Hatfield. 2011. Impacts of deer herbivory on vegetation in Rock Creek Park, 2001–2009. U.S. Department of the Interior, National Park Service. Natural Resource Report NPS/NCR/NCRO/NRTR, 2011/001. 41 pages.
- Krefting, L.W. 1941. Methods of increasing deer browse. *Journal of Wildlife Management* 5: 95–102.
- Kuijper, D.P.J. 2011. Lack of natural control mechanisms increases wildlife–forestry conflict in managed temperate European forest systems. *European Journal of Forest Research* 130: 895–909.
- Kupfer, J.A., K.M. Meitzen & A.R. Pipkin, 2010. Controls of early post-logging successional pathways in a southern floodplain forest. *Forest Ecology and Management* 259: 1880–1889.
- Kush, J.S., R.S. Meldahl, W.D. Boyer. 2000. Understory plant community response to season of burn in natural longleaf pine forests. Pages 32–39 in: W.K. Moser & C.F. Moser (eds). *Fire Proceedings of the 21st Tall Timbers Fire Ecology Conference (1998)*. Tall Timbers Research, Inc., Tallahassee, Florida.
- Kusumoto, B., T. Enoki & Y. Kubota. 2012. Determinant factors influencing the spatial distributions of subtropical lianas are correlated with components of functional trait spectra. *Ecological Research* DOI 10.1007/s11284-012-0993-x. 11 pages.
- **Lackey, J.A. 1977. A revised classification of the tribe Phaseoleae (Leguminosae: Papilionoideae), and its relation to canavanine distribution. *Botanical Journal of the Linnean Society* 74: 163–178.
- Ladwig, L.M., & S. J. Meiners. 2010. Liana host preference and implications for deciduous forest regeneration. *Journal of the Torrey Botanical Society* 137: 103–112.
- Lampert, E.C., L.A. Dyer, M.D. Bowers. 2011. Chemical defense across three trophic levels: *Catalpa bignonioides*, the caterpillar *Ceratomia catalpae*, and its endoparasitoid *Cotesia congregata*. *Journal of Chemical Ecology* 37: 1063–1070.

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- Langenheim, J.H. 1994. Higher plant terpenoids: a phytocentric overview of their ecological roles. *Journal of Chemical Ecology* 20: 1223–1280.
- Larson, B.M.H., P.M. Catling & G.E. Waldron. 2006. The biology of Canadian weeds. 135. *Lonicera japonica* Thunb. *Canadian Journal of Plant Science* 87: 423–438.
- Larson, K. 2000. Circumnutation behavior of an exotic honeysuckle vine and its native congener: influence on clonal mobility. *American Journal of Botany* 87: 533–538.
- Lashley, M.A. 2009. Deer forage available following silvicultural treatments in upland hardwood forests and warm-season plantings. M.Sc. thesis, University of Tennessee, Knoxville. 55 pages.
- Lashley, M.A., C.A. Harper, G.E. Bates & P.D. Keyser. 2011. Forage availability for white-tailed deer following silvicultural treatments in hardwood forests. *Journal of Wildlife Management* 75: 1467–1476.
- Lavin, M., M. F. Wojciechowski, P. Gasson, C. Hughes & E. Wheeler. 2003. Phylogeny of robinoid legumes (Fabaceae) revisited: *Coursetia* and *Gliricidia* recircumscribed, and a biogeographical appraisal of the Caribbean endemics. *Systematic Botany* 28: 387–409.
- Lavin, M., P. Herendenn & M.F. Wojciechowski. 2005. Evolutionary rates analysis of Leguminosae implicates a rapid diversification of lineages during the Tertiary. *Systematic Biology* 54: 575–594.
- Lay, D.W. 1967. Deer range appraisal in eastern Texas. *The Journal of Wildlife Management* 31: 426–432.
- Lay, D.W., & R.E. Murry. 1978. Comments on browse inventories in Louisiana: Pearson and Sternitzke. *Journal of Wildlife Management* 42: 169–171.
- **Lee, K-J., Jo J-C., Lee B-S. & Lee D-S. 1990. The structure of plant community in Kwangnung (Korea) forest (I): Analysis of the forest community of Soribong area by the

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- classification and ordination techniques. *Journal of the Korean Forestry Society* 79: 173–186.
- **Lee Y-N. 1997. *Flora of Korea*. Kyohak Publishing Company, Seoul, Korea.
- LeHouérou, H.N. 1980. Browse in North Africa. Pages 55–82 in: *Browse in Africa: The Current State of Knowledge*. International Livestock Centre for Africa. Addis Ababa, Ethiopia. 491 pages.
- Leicht-Young, S.A., N.B. Pavlovic, K.J. Frohnapple & R. Grundel. 2010. Liana habitat and host preferences in northern temperate forests. *Forest Ecology and Management* 260: 1467–1477.
- Lemke, D. P.E. Hulme, J.A. Brown & W. Tadesse. 2012. Distribution modelling of Japanese honeysuckle (*Lonicera japonica*) invasion in the Cumberland Plateau and mountain region, USA. *Forest Ecology and Management* 262: 139–149.
- Leuzinger, S., A. Hartmann & C. Körner. 2011. Water relations of climbing ivy in a temperate forest. *Planta* 233: 1087–1096.
- Lev-Yadun, S. 2009. Müllerian and Batesian mimicry rings of white-variegated aposematic spiny and thorny plants: a hypothesis. *Israel Journal of Plant Sciences* 57: 107–116.
- Levin, D.A., & B.M. York. 1978. The toxicity of plant alkaloids: an ecogeographic perspective. *Biochemical Systematics and Ecology* 6: 61–76.
- *Li H-L., Huang J-H., Zhang J-X. & Wang W-B. 1984. Discovery of fossil plants in the Yuhuatai Formation in Nanjing. *Geological Review* 30: 575–577.
- **Li Ming. 2011. Preliminary tests for chemical components of *Pileostegia viburnoides* Hook.f. et Thoms. leaves. *Guiding Journal of Traditional Chinese Medicine and Pharmacy* 2011-07.
- *Li Y-M., Wang T-Z. & Wang Z-X. 2001. Studies on chemical constituents in dried buds of *Lonicera similis* Hemsl. [Chinese]. *China Journal of Chinese Materia Medica* 26: 45–47.

196

- Lichtensteiger, C.A., N.A. Johnston & V.R. Beasley. 1997. *Rhamnus cathartica* (buckthorn) hepatocellular toxicity in mice. *Toxicology and Pathology* 25: 449–452.
- Lieurance, D., S. Chakraborty, P. Bonello, S.R. Whitehead, D.M. Bowers & D. Cipollini. 2012. Does variation in defensive secondary metabolites in native and non-native *Lonicera* species explain invasive success? Abstract, 97th Annual Meeting, Ecological Society of America.
- *Lippi, C., & M. Kuypers. 1998. Dealing with wildfires in Florida's urban-wildland interface. *Proceedings of the Florida State Horticultural Society* 111: 232–234.
- Littlefield, K.A., J.P. Mueller, J.P. Muir & B.D. Lambert. 2011. Correlation of plant condensed tannin and nitrogen concentrations to white-tailed deer browse preferences in the Cross Timbers. *Texas Journal of Agriculture and Natural Resource* 24:1-7 .
- Livshultz, T., D.J. Middleton, M.E. Endress & J.K. Williams. 2007. Phylogeny of Apocynoideae and the APSA clade (*Apocynaceae* s.l.). *Annals of the Missouri Botanical Garden* 94: 324–359.
- Lopes, E.A., & J.W. Stuth. 1984. Dietary selection and nutrition of Spanish goats as influenced by brush management. *Journal of Range Management* 37: 554–560.
- Lohmann, L.G. 2006. Untangling the phylogeny of neotropical lianas (Bignoniaceae, Bignoniaceae). *American Journal of Botany* 93: 304–318.
- Ludewig, H.A., Bowyer R.T. 1985. Overlap in winter diets of sympatric moose and white-tailed deer in Maine. *Journal of Mammalogy* 66: 390–392.
- Luginbuhl, J. M., J.T. Green, M.H. Poore & A.P. Conrad. 2000. Use of goats to manage vegetation in cattle pastures in the Appalachian region of North Carolina. *Sheep & Goat Research Journal* 16: 124–135.
- Machado, I.C., A.V. Lopes & M. Sazima. 2006. Plant sexual systems and a review of the breeding systems in the Caatinga, a Brazilian tropical dry forest. *Annals of Botany* 97: 277–287.

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- Maddumage, R., N.J. Nieuwenhuizen, S.M. Bulley, J.M. Cooney, S.A. Green & R.G. Atkinson. 2013. Diversity and relative levels of actinidin, kiwellin and thaumatin-like allergens in fifteen varieties of kiwifruit (*Actinidia*). *Journal of Agricultural and Food Chemistry*, in press.
- Magarey, R.D., D.M. Borchert & J.W. Schlegel. 2008. Global plant hardiness zones for phytosanitary risk analysis. *Scientia Agricola* 65: 54–59.
- Mahady, G.B. 2004. Cascada Sagrada (*Rhamnus purshiana*). Pages 89–94 in: P.M. Coates et al. (eds). *Encyclopedia of Dietary Supplements*. Marcel Dekker, New York.
- Manchester S.R. 1994 . Inflorescence bracts of fossil and extant *Tilia* in North America, Europe, and Asia—patterns of morphologic divergence and biogeographic history. *American Journal of Botany* 81: 1176–1185 .
- Manchester SR. 2001. Leaves and fruits of *Aesculus* (Sapindales) from the Paleocene of North America. *International Journal of Plant Sciences* 162: 985–996.
- Manchester, S.R., & R.A. Dillhoff. 2004. *Fagus* (Fagaceae) fruits, foliage and pollen from the Middle Eocene of Pacific Northwestern North America. *Canadian Journal of Botany* 82: 1509–1517.
- Manos, P., J.J. Doyle & K.C. Nixon. 1999. Phylogeny, biogeography and processes of molecular differentiation in *Quercus* subgenus *Quercus* (Fagaceae). *Molecular Phylogenetics and Evolution* 12: 333–349.
- Manos, P.S., & A.M. Stanford. 2001. The biogeography of Fagaceae: tracking the Tertiary history of temperate and subtropical forests of the Northern Hemisphere. *International Journal of Plant Science* 162: S77–S93.
- Manos, P.S., P.S. Soltis, D.E. Soltis, S.R. Manchester, Oh S-H., C.D. Bell, D.L. Dilcher & D.E. Stone. 2007. Phylogeny of extant and extinct Juglandaceae inferred from the integration of molecular and morphological data sets. *Systematic Biology* 56: 412–430.

198

- Margesin, R., G. Neuner, K.B. Storey. 2007. Cold-loving microbes, plants, and animals—fundamental and applied aspects. *Naturwissenschaften* 94: 77–99.
- Martin, B.H., M. Woods & A.R. Diamond. 2002. The vascular flora of Coffee County, Alabama. *Castanea* 67:227–246.
- Martinez-Cabrera, H.I., & S.R.S. Cevallos-Ferriz. 2006. *Maclura* (Moraceae) wood from the Miocene of the Baja California Peninsula, Mexico: Fossil and biogeographic history of its closer allies. *Review of Palaeobotany and Palynology* 140: 113–122.
- Masters, R., P. Mitchell & S. Dobbs. 1991. Ornamental and garden plants: controlling deer damage. Oklahoma Cooperative Extension Service Fact Sheet F-6427. Oklahoma State University, Stillwater. 8 pages.
- Masters, R.E., T.G. Bidwell & D.R. Elmore. 2004. White-tailed deer habitat evaluation and management guide. Oklahoma Cooperative Extension Service Fact Sheet E-979. Oklahoma State University, Stillwater. 32 pages.
- Masters, R.A., & R. L. Sheley. 2001. Principles and practices for managing rangeland invasive plants. *Journal of Range Management* 54: 502–517.
- Matallana, G., T. Wendt, D.S.D. Araujo & F.R. Scarano. 2005. High abundance of dioecious plants in a tropical coastal vegetation. *American Journal of Botany* 92: 1513–1519.
- Mathiasen, R.M., D.L. Nickrent, D.C. Shaw & D.M. Watson. 2008. Mistletoes: pathology, systematics, ecology, and management. *Plant Disease* 92: 988–1006.
- McClure, M.S. 1989. Importance of weather to the distribution and abundance of introduced adelgid and scale insects. *Agricultural and Forest Meteorology* 47: 291–302.
- McDaniel, C.A. 1992. Major antitermitic components of the heartwood of southern catalpa. *Journal of Chemical Ecology* 18: 359–369.

- McEvoy, P.M., M. Flexen & J.H. McAdam. 2006. The effects of livestock grazing on ground flora in broadleaf woodlands in Northern Ireland. *Forest Ecology and Management* 225: 39–50.
- McEwan, R.W., & B.C. McCarthy. 2008. Anthropogenic disturbance and the formation of oak savanna in central Kentucky, USA. *Journal of Biogeography* 35: 965–975.
- McKenney, D.W., J.H. Pedlar, K. Lawrence, K. Campbell & M.F. Hutchinson. 2007. Beyond traditional hardiness zones: using climate envelopes to map plant range limits. *Bioscience* 57: 929–937.
- Meerts, P. 1997. Foliar macronutrient concentrations of forest understorey species in relation to Ellenberg's indices and potential relative growth rate. *Plant and Soil* 189: 257–265.
- Melick, D., X. F. Yang & J. C. Xu. 2007. Seeing the wood for the trees: how conservation policies can place greater pressure on village forests in southwest China. *Biodiversity and Conservation* 16: 1959–1971.
- Mengel, K., & E.A. Kirkby. 2001. *Principles of Plant Nutrition*. 5th edition. Kluwer Academic Publishers, Dordrecht, The Netherlands.
- Merriam, R.W. 2003. The abundance, distribution and edge associations of six non-indigenous, harmful plants across North Carolina. *Journal of Torrey Botanical Society* 130: 283–291
- Metcalf, D.J. 2005. Biological Flora of the British Isles: *Hedera helix* L. *Journal of Ecology* 93: 632–648.
- Miller, J.H., E.B. Chambliss & C.M. Oswalt. 2008. Maps of occupation and estimates of acres covered by nonnative invasive plants in southern forests using SRS FIA data. U.S.D.A. Forest Service. Available at <http://www.invasive.org/fiamaps/summary.pdf>.
- Milne, R.I., 2006. Northern hemisphere plant disjunctions: a window on Tertiary land bridges and climate change. *Annals of Botany* 98: 465–472.

- Milne, R.I., & R.J. Abbott. 2002. The origin and evolution of Tertiary relict floras. *Advances in Botanical Research* 38: 281–314.
- Mitaine-Offer, A.-C., M. Sauvain, E. Deharo, V. Muñoz & M. Zèches-Hanrot. 2002. A new diterpene from *Tanaecium jaroba*. *Planta Medica* 68: 568–569.
- *Mitchell, F.J.G., & K.J. Kirby. 1990. The impact of large herbivores on the conservation of semi-natural woods in the British uplands. *Forestry* 63: 333–353.
- Mitchell, F.J.G. 2005. How open were European primeval forests? Hypothesis testing using palaeoecological data. *Journal of Ecology* 93: 168–177.
- Mohan, J.E., L.H. Ziska, R.B. Thomas, R.C. Sicher, K. George, J.S. Clark & W.H. Schlesinger. 2008. Biomass and toxicity responses of poison ivy (*Toxicodendron radicans*) to elevated atmospheric CO₂. *Ecology* 89: 585–587.
- Monk, C.D. 1966. An ecological significance of evergreenness. *Ecology* 47:504–505.
- Moore, M.O. 1991. Classification and systematics of eastern North American *Vitis* L. (Vitaceae) north of Mexico. *Sida* 14: 339–367.
- Moreland, D. 2003. Checklist of woody and herbaceous deer food plants of Louisiana. Louisiana Department of Wildlife and Fisheries. 174 pages.
- Morrissey, R.C., M.M. Gautier, J.A.J. Kershaw, D.F. Jacobs, J.R. Seifert & B.C. Fischer. 2009. Grapevine (*Vitis* spp.) dynamics in association with manual tending, physiography, and host tree associations in temperate deciduous forests. *Forest Ecology and Management* 8: 1839–1846.
- Moser, B., M. Schütz, K.E. Hindenlang. 2006. Importance of alternative food resources for browsing by roe deer on deciduous trees: the role of food availability and species quality. *Forest Ecology and Management* 226: 248–255
- Moser, B., M. Schütz & K.E. Hindenlang. 2008. Resource selection by roe deer: are windthrow gaps attractive feeding places? *Forest Ecology and Management* 255: 1179–1185

201

- *Motta, R. 1996. Impact of wild ungulates on forest regeneration and tree composition of mountain forests in the Western Italian Alps. *Forest Ecology and Management* 88: 93–98
- Mountford, E.P., & G.E. Peterken. 2003. Long-term change and implications for the management of woodpastures: experience over 40 years from Denny Wood, New Forest. *Forestry* 76: 19–43.
- Mu X.-Y., Zhao L.-C. & Zhang Z.-X. 2012. Phylogeny of *Celastrus* L. (Celastraceae) inferred from two nuclear and three plastid markers. *Journal of Plant Research* 125: 619–630.
- Mudrak, E.L., S.E. Johnson, D.M. Waller, 2009. Forty-seven year changes in vegetation at the Apostle Islands: effects of deer on the forest understory. *Natural Areas Journal* 29: 167–176.
- Muenschler, W.C. 1961. Poisonous plants of the United States, 2nd edition. Macmillan Co., New York. 277 pages.
- Muller, J. 1981. Fossil pollen records of extant angiosperms. *Botanical Review* 47: 1–140.
- Muller, R.N. 1978. The phenology, growth and ecosystem dynamics of *Erythronium americanum* in the northern hardwood forest. *Ecological Monographs* 48: 1–20.
- Muller, R.N., P.J. Kalisz & T.W. Kimmerer. 1987. Intraspecific variation in production of astringent phenolics over a vegetation-resource availability gradient. *Oecologia* 72: 211–215.
- Mulligan, G.A., & B.E. Junkins. 1977. The biology of Canadian weeds. 23. *Rhus radicans* L. *Canadian Journal of Plant Science* 57: 515–523.
- Munger, G.T. 2002. *Lonicera japonica*. In: Fire Effects Information System. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fire Sciences Laboratory [<http://www.fs.fed.us/database/feis/>].
- Murie, A. 1934. The Moose of Isle Royale. University of Michigan Museum of Zoology Miscellaneous Publication 25. 44 pages.

202

- Mysterud, A., L.-B. Lian & D.O. Hjermmann. 1999. Scale-dependent trade-offs in foraging by European roe deer (*Capreolus capreolus*) during winter. *Canadian Journal of Zoology* 77: 1486–493.
- Nagarkoti, A., & T.B. Thapa. 2007. Food habits of barking deer (*Muntiacus muntjac*) in the Middle Hills of Nepal. *Hystrix, International Journal of Mammalogy (new series)* 18: 77–82.
- Nakagawa, N. 1989. Bioenergetics of Japanese Monkeys (*Macaca fuscata*) on Kinkazan Island during winter. *Primates* 30: 441–460.
- Nakamura T., E. Okuyama & M. Yamazaki. 1996. Neurotropic components from star anise (*Illicium verum* Hook. fil.). *Chemical & Pharmaceutical Bulletin* 44:1908-1914.
- National Park Service. 2007. Record of Decision: Rock Creek Park and the Rock Creek and Potomac Parkway Final General Management Plan, Environmental Impact Statement. U.S. Department of the Interior, National Park Service. Washington, D.C.
- National Park Service. 2012. Rock Creek Park, White-tailed Deer Management Plan and Final Environmental Impact Statement. U.S. Department of the Interior, National Park Service. Washington, D.C. 25 pages.
- NatureServe. 2012. NatureServe Explorer: an online encyclopedia of life [<http://www.natureserve.org/explorer/>].
- Nelle, S.A. 1996. Management and use of browse in the Edwards Plateau of Texas. Pages 151–155 in: J.R. Barrow, E.D. McArthur, R.E. Sosebee & R.J. Tausch. *Shrubland Ecosystem Dynamics in a Changing Environment*. U.S.D.A. Forest Service, General Technical Report INT-GTR-338. Ogden, Utah.
- Nicotra, A.B., A. Leigh, C.K. Boyce, C.S. Jones, K.J. Niklas, D.L. Royer & H. Tsukaya. 2011. The evolution and functional significance of leaf shape in the angiosperms. *Functional Plant Biology* 38: 535–552.

203

- Nie, Z.-L., Wen J., H. Azuma, Qiu Y-L., Sun H., Meng Y., Sun W-B. & E.A. Zimmer. 2008. Phylogenetic and biogeographic complexity of Magnoliaceae in the northern hemisphere inferred from three nuclear data sets. *Molecular Phylogenetics and Evolution* 48: 1027–1040.
- Nie Z.-L., Sun H., Meng Y. & Wen J. 2009. Phylogenetic analysis of *Toxicodendron* (Anacardiaceae) and its biogeographic implications on the evolution of north temperate and tropical intercontinental disjunctions. *Journal of Systematics and Evolution* 47: 416–430.
- Nie, Z-L., Sun H., Chen Z-D., Meng, Y., S.R. Manchester & Wen J. 2010. Molecular phylogeny and biogeographic diversification of *Parthenocissus* (Vitaceae) disjunct between Asia and North America. *American Journal of Botany* 97: 1342–1353.
- Niering, W.A., & G.D. Dreyer. 1989. Effects of prescribed burning on *Andropogon scoparius* in postagricultural grasslands in Connecticut. *American Midland Naturalist* 122: 88–102.
- Niesenbaum, R.A. 1992. The effects of light environment on herbivory and growth in the dioecious shrub *Lindera benzoin* (Lauraceae). *American Midland Naturalist* 128: 270–275.
- Nikolai, V. 1988. Phenolic and mineral content of leaves influences decomposition in European forest ecosystems. *Oecologia* 75: 575–579.
- Ningen, S.S., J.C. Cole, M.W. Smith, D.E. Dunn & K.E. Conway. 2005. Increased shade intensity and afternoon irrigation decrease anthracnose severity on three *Euonymus fortunei* cultivars. *Hortscience* 40: 111-113.
- Nixon, C.M., M.W. McClain & K.R. Russell. 1970. Deer food habits and range characteristics in Ohio. *Journal of Wildlife Management* 34: 870–886.
- Noland, J. E., & S.H. Morrison. 1954. The digestibility of honeysuckle (*Lonicera japonica*) for yearling dairy bulls. *Journal of Dairy Science* 37: 173–175.
- Nomiya, H., W. Suzuki, T. Kanazashi, M. Shibata, H. Tanaka & T. Nakashizuka. 2003. The Response of forest floor vegetation and tree regeneration to deer exclusion and disturbance in a Riparian Deciduous Forest, Central Japan. *Plant Ecology* 164: 263–276.

204

- Nostro, A., M.P. Germano, V. D'Angelo, A. Marino & M.A. Cannatelli. 2000. Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Letters in Applied Microbiology* 30: 379–384.
- Nuzzo, V. 1997. Element stewardship abstract for *Lonicera japonica*, Japanese Honeysuckle The Nature Conservancy, Arlington, Virginia. 22 pages.
- Nyahangare, E.T., T. Hove, B.M. Mvumi, H. Hamudikuwanda, S.R. Belmain, J. Madzimure & P.C. Stevenson. 2012. Acute mammalian toxicity of four pesticidal plants. *Journal of Medicinal Plants Research* 6: 2674-2680.
- Oberhuber, W., & H. Bauer. 1991. Photoinhibition of photosynthesis under natural conditions in ivy (*Hedera helix* L.) growing in an understory of deciduous trees. *Planta* 185: 545–553.
- *Ogura, S-I. 2011. Diet selection and foraging behavior of cattle in species-rich vegetation. *Advanced Studies on Sustainable Animal Production: Interrelationships among Human, Animal and Environment*, 8th International Symposium of Integrated Field Science. Abstract 2011-03.
- Ohi-toma T., T. Sugawara, H. Murata, S. Wanke, C. Neinhuis & J. Murata. 2006. Molecular phylogeny of *Aristolochia* sensu lato (Aristolochiaceae) based on sequences of *rbcl*, *matK*, and *phyA* genes, with special reference to differentiation of chromosome numbers. *Systematic Botany* 31: 481–492.
- Ohwi, J. 1965. *Flora of Japan*. Edited by F.G. Meyer & E.H. Walker. Smithsonian Institution, Washington, D.C. 1067 pages.
- Okerman, A. 2000. Combating the “Ivy Desert”: The Invasion of *Hedera helix* (English Ivy) in the Pacific Northwest United States. *Restoration and Reclamation Review* 6-4, 10 pages. Department of Horticultural Science, University of Minnesota, St. Paul.

205

- Olien, W.C. 2001. Introduction to the muscadines. Pages 1–13 in: F.M. Basiouny & D.G. Himelrick (eds). *Muscadine Grapes*. American Society of Horticultural Science Press, Alexandria, Virginia.
- Olmstead, R.G., M.L. Zjhra, L.G. Lohmann, S.O. Grose & A.J. Eckert 2009. A molecular phylogeny and classification of Bignoniaceae. *American Journal of Botany* 96: 1731–1743
- Olson, L. 2006. The economics of terrestrial invasive species: a review of the literature. *Agricultural Resource Economics Review* 35: 178–194.
- Öquist, G., & N.P.A. Huner. 2003. Photosynthesis of overwintering evergreen plants. *Annual Reviews of Plant Biology* 54: 329–355.
- Ordoñez, J.C., P.M. van Bodegom, J.M. Witte, I.J. Wright, P.B. Reich & R. Aerts. 2009. A global study of relationships between leaf traits, climate and soil measures of nutrient fertility. *Global Ecology and Biogeography* 18: 138–149.
- Ortiz, R.D.C., E.A. Kellogg & H.V.D. Werff. 2007. Molecular phylogeny of the moonseed family (Menispermaceae): implications for morphological diversification. *American Journal of Botany* 94: 1425–1438.
- Palomo, E. 2012. Do deer eat kiwi vines? Demand Media, Hearst Communications, Inc. Posted at <http://home.guides.sfgate.com/deer-eat-kiwi-vines-49347.html>.
- **Pan Q-H., Zhang W-H. & Lu R-Q. 2005. Analysis of components of crude protein and amino acid in leaves of *Euonymus fortunei*. *Chinese Agricultural Science Bulletin* 2005-08-058.
- Papageorgiou, N., C. Neophytou, A. Spais, C. Vavalekas, 1981. Food preferences and protein and energy requirements for maintenance of roe deer. *Journal of Wildlife Management* 45: 728-733.
- Park, D.M., S.S Hong, C. Lee, M.S Lee, S.J Kang, Y.S Shin, J-K. Jung, J.T Hong, Y. Kim, M.K. Lee, B.Y. Hwang. 2010. Naphthoquinones from *Catalpa ovata* and their inhibitory effects on the production of nitric oxide. *Archives of Pharmacological Research* 33: 381–385.

206

- Parker, J. 1962. Relationships among cold hardiness, water-soluble protein, anthocyanins and free sugars in *Hedera helix* L. *Plant Physiology* 37: 809–813.
- Parkes, J.P. 2008. Management of Himalayan thar (*Hemitragus jemlahicus*) in New Zealand: the influence of Graeme Caughley. *Wildlife Research* 36: 41–47
- Paul, J.S. 1993. Goats control poison ivy, oak and sumac. *Small Farm Today*, Missouri Farm Publishing 19: 1–44.
- Pederson, B.A. and B.S. Wallis. 2004. Effects of white-tailed deer herbivory on forest gap dynamics in a wildlife preserve, Pennsylvania, USA. *Natural Areas Journal* 24: 81–94.
- Peet, R.K., J.D. Fridley & J.M. Gramling. 2003. Variation in species richness and species pool size across a pH gradient in forests of the southern Blue Ridge Mountains. *Folia Geobotanica* 38: 391–401.
- Peet, R.K., M.T. Lee, M.D. Jennings & D. Faber-Langendoen. 2012. VegBank – a permanent, open-access archive for vegetation-plot data. Pages 233–242 in: J. Dengler, J. Oldeland, F. Jansen, M. Chytrý, J. Ewald, M. Finckh, F. Glöckler, G. Lopez-Gonzalez, R.K. Peet & J.H.J. Schaminée, (eds). *Vegetation databases for the 21st century*. *Biodiversity & Ecology* 4: 233–241.
- *Pei J., Wan D-G., Tang Y., Guo J-L., Yang X-J. & Cai W-L. 2009. Study on the renal toxicity of *Caulis clematidis armandii* in rats. *West China Journal of Science* 24: 461–463.
- Peng, S.L., Chen B-M., Lin Z-G., Ye Y-H., Yu Y-N., Li J-L. & Li H-J. 2009. The status of noxious plants in lower subtropical region of China. *Acta Ecologica Sinica* 29: 79–83.
- Peñuelas, J., J. Sardans, C. Stefanescu, T. Parella & I. Filella. 2006. *Lonicera implexa* leaves bearing naturally laid eggs of the specialist herbivore *Euphydryas aurinia* have dramatically greater concentrations of iridoid glycosides than other leaves. *Journal of Chemical Ecology* 32: 1925–1933.

207

- **Perrin, P.M., F.J.G. Mitchell & D.L. Kelly. 2011. Long-term deer exclusion in yew-wood and oakwood habitats in southwest Ireland: changes in ground flora and species diversity. *Forest Ecology and Management* 262: 2328–2337.
- Petrides, G.A. 1975. Principal foods versus preferred foods and their relations to stocking rate and range condition. *Biological Conservation* 7: 161–169.
- Pettorelli, N., J.-M. Gaillard, P. Duncan, J.-P. Ouellet & G. Van Laere. 2001. Population density and small-scale variation in habitat quality affect phenotypic quality in roe deer. *Oecologia* 128: 400–405.
- Pettorelli, N., S. Dray, J.-M. Gaillard, D. Chessel, P. Duncan, A. Illius, N. Guillon, F. Klein & G.V. Laere. 2003. Spatial variation in springtime food resources influences the winter body mass of roe deer fawns. *Oecologia* 137: 363–369.
- Pigg, K.B., S.M. Ickert-Bond & J. Wen. 2004. Anatomically preserved *Liquidambar* (Altingiaceae) from the Middle Miocene of Yakima Canyon, Washington State, USA, and its biogeographic implications. *American Journal of Botany* 91: 499–509.
- Pimentel, D., R. Zuniga & D. Morrison. 2005. Update on the environmental and economic costs associated with alien–invasive species in the United States. *Ecological Economics* 52: 273–88.
- Platt, S.G., & C.G. Brantley. 1997. Canebrakes: an ecological and historical perspective. *Castanea* 62: 8–21.
- Porter, W.F., & H. B. Underwood. 1999. Of elephants and blind men: deer management in the U.S. National Parks. *Ecological Applications* 9: 3–9
- Putman, R.J., P.J. Edwards, J.C.E. Mann, R.C. How & S.D. Hill. 1989. Vegetational and faunal changes in an area of heavily grazed woodland following relief of grazing. *Biological Conservation* 47: 13–32.

208

- Putz, F.E., & H.A. Mooney. 1991. *The Biology of Vines*. Cambridge University Press, Cambridge, England. 544 pages.
- Putz, F.E. 1995. Relay ascension of big trees by vines in Rock Creek Park, District of Columbia. *Castanea* 60: 167–169.
- Pykälä, J., 2000. Mitigating human effects on European biodiversity through traditional animal husbandry. *Conservation Biology* 14: 705–712.
- Qaderi, M.M, D.R Clements & P.B. Cavers. 2009. The biology of Canadian weeds. 139. *Rhamnus cathartica* L. *Canadian Journal of Plant Science* 89: 169–189.
- *Qian Z-M., Li H-J., Qi F-F., He Q-H. & Li P. 2006. Flavonoids from the aerial parts of *Lonicera syringantha* Maxim. *Chemistry and Industry of Forest Products* 2006-03.
- *Qin S-J., Li H-J., Li P. & Tang D. 2008 Studies on chemical constituents of aerial parts of *Lonicera dasystyla* Rehd. *Chinese Pharmacology Journal* 43: 662–664.
- Qu F-L., Ding Q-L. & Zhang H-M. 2001. Study on the chemical composition in *Euonymus fortunei*. *Journal of Nanjing Military Medical College* 2001-04.
- Rackham, O. 1990. *Trees & Woodland in the British Landscape*. Revised edition. Phoenix Press, London, England. 234 pages.
- Rackham, O. 2003. *Ancient Woodland: its History, Vegetation and Uses in England*. 2nd edition. Edward Arnold, London. 624 pages
- Rackham, O. 2006. *Woodlands*. Collins, London. 609 pages.
- Raulston, J.C. 1992. Evergreen vines for use in the southeastern United States. *Friends of the Raulston Arboretum Newsletter* 23. North Carolina State University, Raleigh. [Original version written for 1992 Southern Nurserymen's Association Research Workers Proceedings.]
- Reich, P.B. M.B. Walters & D.S. Ellsworth. 1992. Leaf lifespan in relation to leaf, plant, and stand characteristics among diverse ecosystems. *Ecological Monographs* 62: 365–392.

209

- Renton, J.C. 2010. The impact of cattle grazing on aspen regeneration on Crown Lands in western Manitoba. M.Sc. thesis, University of Manitoba, Winnipeg.
- Renner, S.S., & R.E. Ricklefs. 1995. Dioecy and its correlates in the flowering plants. *American Journal of Botany* 82: 596–606
- Renner, S.S., G.W. Grimm, G.M. Schneeweiss, T.F. Stuessey & R.E. Ricklefs. 2008. Rooting and dating maples (*Acer*) with an uncorrelated-rates molecular clock: implications for north American/Asian disjunctions. *Systematic Biology* 57: 795–808.
- Retuerto, R., B. Fernández-Lema & J. R. Obeso, 2006. Changes in photochemical efficiency in response to herbivory and experimental defoliation in the dioecious tree *Ilex aquifolium*. *International Journal of Plant Science* 167: 279–289.
- Richardson, J.E., L.W. Chatrou, J.B. Mols, R.H.J. Erkens & M.D. Pirie. 2004. Historical biogeography of two cosmopolitan families of flowering plants: Annonaceae and Rhamnaceae. *Philosophical Transactions of the Royal Society of London Series B* 359: 1495–1508.
- Rinkes, Z.L. 2004. Eastern wild turkey (*Meleagris gallopavo silvestris*) habitat preference and the impact of scratch foraging on hardwood regeneration in mixed oak forests. M.Sc. thesis, Ohio University, Athens. 34 pages.
- Rock Creek Conservancy. 2012. Help save the trees in Rock Creek Park! Available at <http://www.rockcreekconservancy.org/index.php/component/content/article/280-save-trees>.
- Roe, F.G. 1951. *The North American buffalo*. University of Toronto Press, Toronto, Ontario, Canada. 957 pages.
- Rogosic, J., R.E. Estell, D. Skobic, A. Martinovic & S. Maric. 2006. Role of species diversity and secondary compound complementarity on diet selection of Mediterranean shrubs by goats. *Journal of Chemical Ecology* 32: 1279–1287.

- *Rogosic, J., R.E. Estell, D. Skobic & S. Stanic. 2007. Influence of secondary compound complementarity and species diversity on consumption of Mediterranean shrubs by sheep. *Applied Animal Behaviour Science* 107: 58–65.
- Rooney, T.P. 2009. High white-tailed deer densities benefit graminoids and contribute to biotic homogenization of forest ground-layer vegetation. *Plant Ecology* 202: 103–111.
- *Rossell, C.R., S. Patch & S. Salmons. 2007. Effects of deer browsing on native and non-native vegetation in a mixed oak-beech forest on the Atlantic Coastal Plain. *Northeastern Naturalist* 14: 61–72.
- Ruberson, J.R., K. Takasu, G.D. Buntin, J.E. Eger, W.A. Gardner, J.K. Greene, T.M. Jenkins, W.A. Jones, D.M. Olson, R.M. Roberts, D.R. Suiter, M.D. Toews. 2012. From Asian curiosity to eruptive American pest: *Megacopta cribraria* (Hemiptera: Plataspidae) and prospects for its biological control. *Applied Entomology and Zoology* DOI 10.1007/s13355-012-0146-2
- Rudis, V.A., A. Gray, W. McWilliams, R. O'Brien, C. Olson, S. Oswalt & B. Schulz. 2005. Regional monitoring of nonnative plant invasions with the forest inventory and analysis Program. Pages 49–64 in: R.E. McRoberts, G.A. Reams, P.C. Van Deusen & W.H. McWilliams (eds). *Proceedings of the Sixth Annual Forest Inventory and Analysis Symposium*. U.S.D.A. Forest Service, General Technical Report WO-70.
- Rule, L.C., J.P. Colletti, T.R. Liu, S.E. Jungst, C.W. Mize & R.C. Schultz. 1994. Agroforestry and forestry-related practices in the midwestern United States. *Agroforestry Systems* 27: 79–88.
- Saima, S., A.A. Dasti, F. Hussain, S.M. Wazir & S.A. Malik. 2009. Floristic compositions along an 18 km long transect in Ayubia National Park, District Abbottabad, Pakistan. *Pakistan Journal of Botany* 41: 2115–2127.

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- Sakai, A., H. Nomiya & W. Suzuki. 2002. Horizontal distribution of stolons of a temperate liana, *Wisteria floribunda* DC., and its ecological significance. *Journal of Forest Research* 7:125–130.
- Samain, M.S., S. Wanke & P. Goetghebeur. 2010. Unraveling extensive paraphyly in the genus *Hydrangea* s. l. with implications for the systematics of tribe Hydrangeae. *Systematic Botany* 35: 593–600.
- Samant, S.S. 1998. Diversity, distribution and conservation of fodder resource of west Himalaya, India. Pages 109–128 in: B. Misri (ed). *Proceedings of the Third Temperate Pasture and Fodder Network (TAPAFON)*. Pokhara, Nepal, sponsored by F.A.O. Rome.
- Samant, S.S., M. Singh, M. Lal & S. Pant. 2007. Diversity, distribution and prioritization of fodder species for conservation in Kullu District, northwestern Himalaya, India. *Journal of Mountain Science* 4: 259–274
- Sasek, T.W., & B.R. Strain. 1991. Effects of CO₂ enrichment on the growth and morphology of a native and an introduced honeysuckle vine. *American Journal of Botany* 78: 69–75.
- Sawian, J.T., S. Jeeva, F.G. Lyndem, B.P. Mishra & R.C. Laloo. 2007. Wild edible plants of Meghalaya, North-east India. *Natural Product Radiance* 6: 410–426.
- Schaetzl, R.J., F.J. Krist, K. Stanley & C.M. Hupy. 2009. The natural soil drainage index: an ordinal index of long-term soil wetness. *Physical Geography* 30: 383–409.
- Schaffner, J.H. 1904. Poisonous and other injurious plants of Ohio. *The Ohio Naturalist* 4: ??–35, 69–73.
- **Schaller, G.B., Teng Q-T., Pan W-S. & Qiang Z-S. 1986. Feeding behavior of Sichuan takin (*Budorcas taxicolor*). *Mammalia* 50: 311–322.
- Schanbel, A., & J.F. Wendel. 1998. Cladistic biogeography of *Gleditsia* (Leguminosae) based on ndhF and rpl16 chloroplast gene sequences. *American Journal of Botany* 85: 1753–1765.

212

- Schierenbeck, K.A., & J.D. Marshall. 1993. Seasonal and diurnal patterns of photosynthetic gas exchange for *Lonicera sempervirens* and *L. japonica* (Caprifoliaceae). *American Journal of Botany* 80: 1292–1299
- Schierenbeck, K.A., R.N. Mack & R.R. Sharitz. 1994. Effects of herbivory on growth and biomass allocation in native and introduced species of *Lonicera*. *Ecology* 75: 1661–1672.
- Schnitzer, S.A., & F. Bongers. 2002. The ecology of lianas and their role in forests. *Trends in Ecology and Evolution* 17: 223–230.
- Schnitzler, A., & P. Heuzé. 2006. Ivy (*Hedera helix* L.) dynamics in riverine forests: effects of river regulation and forest disturbance. *Forest Ecology and Management* 236: 12–17.
- Schoch, W., I. Heller, F.H. Schweingruber & F. Kienast. 2004. Wood Anatomy of Central European Species. Available at <http://www.woodanatomy>.
- Schönenberger, J., & E.M. Friis. 2001. Fossil flowers of ericalean s.l. affinity from the Late Cretaceous of southern Sweden. *American Journal of Botany* 88: 467–480.
- Schulz, B.K., & A.N. Gray. 2012. The new flora of northeastern USA: quantifying introduced plant species occupancy in forest ecosystems. *Environmental Monitoring and Assessment* DOI 10.1007/s10661-012-2841-4. 27 pages.
- Schwärzel, K., K-H. Feger, J. Häntzschel, A. Menzer, U. Spank, F. Clausnitzer, B. Köstner & C. Bernhofer. 2009. A novel approach in mode-based mapping of soil water conditions at forest sites. *Forest Ecology and Management* 258: 2163–2174.
- Schweitzer, J.A., & K.C. Larson. 1999. Greater morphological plasticity of exotic honeysuckle species may make them better invaders than native species. *Journal of the Torrey Botanical Society* 126: 15–23.
- SE-EPPC [Southeast Exotic Pest Plants Council]. 2012 (ongoing). Early Detection and Distribution Mapping System [<http://www.eddmaps.org/southeast/distribution/>]. Center for Invasive Species and Ecosystem Health, University of Georgia, Athens.

213

- Segelquist, C.A., & R.E. Pennington. 1968. Deer browse in the Ouachita forest in Oklahoma. *Journal of Wildlife Management* 32: 623–626.
- Segelquist, C.A., & Rogers, M.J. 1975. Response of Japanese honeysuckle to fertilization. *Journal of Wildlife Management* 39: 769–775.
- Seidelmann, K., B. Melzer & T. Speck. 2012. The complex leaves of the Monkey's Comb (*Amphilophium crucigerum*, Bignoniaceae): a climbing strategy without glue. *American Journal of Botany* 99: in press.
- Seki, Y., & M. Koganezawa. 2012. Does sika deer overabundance exert cascading effects on the raccoon dog population? *Journal of Forest Research* DOI 10.1007/s10310-011-0332-z, 7 pages.
- Setoguchi, M. 1990. Food habits of red-bellied tree squirrels on a small island in Japan. *Journal of Mammalogy* 71: 570–578.
- Shah, G.M., U. Jan, B.A. Bhat & F.A. Ahangar. 2009. Diets of hangul deer, *Cervus elaphus hanglu* (Cetartiodactyla: Cervidae), [in] Dachigam National Park, Kashmir, India. *Journal of Threatened Taxa* 1: 398–400.
- Shaw, C.E. 2008. An Evaluation of Quality Deer Management Programs in Tennessee. M.Sc. thesis, University of Tennessee, Knoxville. 219 pages.
- Shaw, C.E., C.A. Harper, M.W. Black, A.E. Houston. 2010. Initial effects of prescribed burning and understory fertilization on browse production in closed-canopy hardwood stands. *Journal of Fish and Wildlife Management* 10: 64–72.
- Shi J-P., & Zhu H. 2009. Tree species composition and diversity of tropical mountain cloud forest in the Yunnan, southwestern China. *Ecological Research* 24: 83–92.
- Shi W., Wang G. & Han W. 2012. Altitudinal variation in leaf nitrogen concentration on the eastern slope of Mount Gongga on the Tibetan Plateau, China. *PLOS ONE* 7: e44628. 6 pages.

214

- Shinde, A.K., S.K. Sankhyan, R. Bhatta & D.L. Verma. 2000. Seasonal changes in nutrient intake and its utilization by range goats in a semi-arid region of India. *Journal of Agricultural Science, Cambridge* 135: 429–436.
- Siegler, H.R. 1937. Winter rodent damage to game cover. *Journal of Mammalogy* 18: 57–61.
- Simmons, M.P., V. Savolainen, C.C. Clevinger, R.H. Archer & J.I. Davus. 2001. Phylogeny of the Celastraceae inferred from 26S nuclear ribosomal DNA, phytochrome B, *rbcL*, *atpB*, and morphology. *Molecular Phylogenetics and Evolution* 19: 353–366.
- Simon, S.A., T.K. Collins, G.L. Kauffman, W.H. McNab & C.J. Ulrey. 2005. Ecological zones in the Southern Appalachians: first approximation. U.S.D.A., Forest Service, Southern Research Station Research Paper No. SRS-41.
- Skulman, B.W., J.D. Mattice, M.D. Cain & E.E. Gbur. 2004. Evidence for allelopathic interference of Japanese honeysuckle (*Lonicera japonica*) to loblolly and shortleaf pine regeneration. *Weed Science* 52: 433–439.
- Smiley, C.J., J. Gray & L.M. Huggins. 1975. Preservation of Miocene fossils in unoxidized lake deposits, Clarkia, Idaho; with a section on fossil insecta by W. F. Barr and J. M. Gillespie. *Journal of Paleontology* 49: 833–844.
- Smith, J. 2010. The history of temperate agroforestry. 17 pages. Organic Research Centre, Newbury, England. Available at http://orgprints.org/18173/1/History_of_agroforestry_v1.0.pdf.
- Smith, L.M., & H.L. Reynolds. 2012. Positive plant-soil feedback may drive dominance of a woodland invader, *Euonymus fortunei*. *Plant Ecology* 213: 853–860.
- Smith, M.W. 1982. Effects of selected silvicultural practices on swamp rabbit (*Sylvilagus aquaticus*) habitat. M.Sc. thesis. Mississippi State University, Starkville. 176 pp.

215

- Smith, S.A. 2009. Taking into account phylogenetic and divergence time uncertainty in a parametric biogeographic analysis: an example using the Northern Hemisphere plant clade Caprifoliaceae. *Journal of Biogeography* 36: 2324–2337.
- Smyth, B.B. 1903. Preliminary list of medicinal and economic Kansas plants, with their reputed therapeutic properties. *Transactions of the Kansas Academy of Science* 18: 191–209.
- *Son, K.H., J.O. Park, K.C. Chung, H.W. Chang, H.P. Kim, J.S. Kim & S.S. Kang. 1992. Flavonoids from the aerial part of *Lonicera japonica*. *Archives of Pharmacological Research* 15: 365–370.
- Sotala, D.J., & C.M. Kirkpatrick. 1973. Foods of white-tailed deer, *Odocoileus virginianus*, in Martin County, Indiana. *American Midland Naturalist* 89: 281–286.
- Sousa, D.P., & R.N. Almeida. 2002. Neuroleptic-like properties of the chloroform extract of *Maytenus obtusifolia* Mart. roots. *Biological and Pharmacological Bulletin* 28: 224–225.
- Sparks, D. 2003. *Environmental Soil Chemistry*. Academic Press, London, England. 352 pages.
- Sparks, J.C., R.E. Masters, D.M. Engle, M.W. Palmer & G.A. Bukenhofer. 1998. Effects of late growing-season and late dormant-season prescribed fire on herbaceous vegetation in restored pine-grassland communities. *Journal of Vegetation Science* 9: 133–142.
- Spivey, A.C., M. Weston & S. Woodhead. 2001. Celastraceae sesquiterpenoids: biological activity and synthesis. *Chemistry Society Reviews* 31: 43–59.
- *Srećec, S., V. Zechner-Krpan, V. Petravić-Tominac, L. Kozačinski, M. Popović, A. Čerenak. 2011. Hop (*Humulus lupulus* L.) secondary metabolites and possibilities of using hop in nutrition of ruminants. *Hmeljarski Bilten* 18: 5–13.
- Stafne, R.A., A.E. Einert, G.L. Klingaman. 2005. Fertilizer applications on establishment and growth of three groundcover species in sun and shade. *Journal of Environmental Horticulture* 23: 157–161

216

- Staines, B.W., & D. Welch. 1981 Deer and their woodland habitats. Pages 138–142 in: F.T. Last & A.S. Gardiner (eds.). *Forest and woodland ecology: an account of research being done in ITE*. Symposium No. 8, NERC, Institute of Terrestrial Ecology, Cambridge.
- Stamp, N. 2003. Out of the quagmire of plant defense hypotheses. *Quarterly Review of Biology* 78: 23–55.
- Stanford, A.M., R. Harden & C.R. Parks. 2000 Phylogeny and biogeography of *Juglans* (Juglandaceae) based on matK and ITS sequence data. *American Journal of Botany* 87: 872–882.
- **Stapf, O. 1926. *Actinidia kolomikta* (East Asia). *Curtis' Botanical Magazine* 151 (“1925”) Tab. 9093. [Quoted in: J.M. Condon. 1991. Ph.D. thesis, University of Canterbury.]
- Stevens, M.T., & S.M. Esser. 2009. Growth-defense tradeoffs differ by gender in dioecious trembling aspen (*Populus tremuloides*). *Biochemical Systematics and Ecology* 37:567–573.
- Stevens, P.F. 2012 [continuing]. Angiosperm Phylogeny Website. Version 9, June 2008: <http://www.mobot.org/MOBOT/research/AP/>.
- Stransky, J. J. 1984. Forage yield of Japanese honeysuckle after repeated burning or mowing. *Journal of Range Management* 37: 237–238.
- Sun, Q., K. Yoda, M. Suzuki & H. Suszuki. 2003. Vascular tissue in stem and roots of woody plants can conduct light. *Journal of Experimental Botany* 54: 1627–1635.
- Sung T.V., N.H. Cuong, T.T. Thuy, P.T. Ninh & L.T.H. Nhung. 2008. Isolation and structural characterization of phenolic glycoside and triterpenes in *Celastrus hindsii* Benth. *Journal of Chemistry* 46: 224–228.
- Surette, S.B., & J.S. Brewer. 2008. Inferring relationships between native plant diversity and *Lonicera japonica* in upland forests in north Mississippi, USA. *Applied Vegetation Science* 11: 205–214.

217

- Swearingen, J.M. & S. Diedrich. 2004. English Ivy. U.S. National Park Service, Washington, DC. Available at: <http://www.nps.gov/plants/alien/fact/hehe1.htm>.
- Swihart, R.K., & R.H. Yahner. 1983. Browse preferences of jackrabbits and cottontails for species used in shelterbelt plantings. *Journal of Forestry* 8: 92–94.
- Takahashi, H., & K. Kaji. 2001. Fallen leaves and unpalatable plants as alternative foods for sika deer under food limitation. *Ecological Research* 16: 257–262.
- Takatsuki, S. 1986. Food habits of Sika deer on Mt. Goyo, Northern Honshu. *Ecological Research* 1: 119–128.
- Takatsuki, S. 1988. Rumen contents of Sika Deer on Tsushima Island, western Japan. *Ecological Research* 3: 181–183.
- Takatsuki, S. 2008. What is “natural” vegetation? A reconsideration of herbivory by wild ungulates. Pages 239–247 in: D.R. McCullough, S. Takatsuki & K. Kaji (eds.). *Sika Deer: Biology and Management of Native and Management of Native and Introduced Populations*. Springer, Tokyo.
- Takatsuki, S. 2009. Effects of sika deer on vegetation in Japan: a review. *Biological Conservation*. 142: 1922–1929.
- Takatsuki, S., & Y. Hirabuki. 1997. Effects of Sika deer browsing on the structure and regeneration of the *Abies firma* forest on Kinkazan Island, Northern Japan. *Journal of Sustainable Forestry* 6: 203–221.
- Talley, S.M., R.O. Lawton & W.N. Setzer. 1996. Host preferences of *Rhus radicans* (Anacardiaceae) in a southern deciduous hardwood forest. *Ecology* 77: 1271–1276
- Tanentzap, A.J., L.E. Burrows, W.G. Lee, G. Nugent, J.M. Maxwell & D.A. Coomes. 2009. Landscape-level vegetation recovery from herbivory: progress after four decades of invasive red deer control. *Journal of Applied Ecology* 46: 1064–1072.

218

- Tang, C.Q. 2006. Forest vegetation as related to climate and soil conditions at varying altitudes on a humid subtropical mountain, Mount Emei, Sichuan, China. *Ecological Research* 21: 174–180.
- Tansley, A.G. 1935. *The British Isles and their Vegetation*. Cambridge University Press, Cambridge. 474 pages.
- Taverna, K., R.K. Peet & L.C. Phillips. 2005. Long-term change in ground-layer vegetation of deciduous forests of the North Carolina Piedmont, USA. *Journal of Ecology* 93: 202–213.
- Telfer, E.S. 1967. Comparison of a deer yard and a moose yard in Nova Scotia. *Canadian Journal of Zoology* 45: 485–490.
- Thanabhorn, S., K. Jaijoy, S. Thamaree, K. Ingkaninand & A. Panthong. 2006. Acute and subacute toxicity study of the ethanol extract from *Lonicera japonica* Thunb. *Journal of Ethnopharmacology* 107: 370–373.
- Thill, R.E. 1984. Deer and cattle diets on Louisiana pine-hardwood sites. *Journal of Wildlife Management* 48: 788–798.
- Thill, R.E., & A. Martin. 1989. Deer and cattle diets on heavily grazed pine-bluestem range. *Journal of Wildlife Management* 53: 540–548.
- Thimonier, A., J.L. Dupouey, F. Bost, & M. Becker. 2006. Simultaneous eutrophication and acidification of a forest ecosystem in North-East France. *New Phytologist* 126: 533–539.
- Thomas, L.K. 1980. *The Impact of Three Exotic Plant Species on a Potomac Island*. National Park Service Scientific Monograph Series, no. 13. 179 pp.
- Thomas, L.K. 1998. Topographic alterations, forest structure, and invasion by English ivy (*Hedera helix* L.) in the Rock Creek Floodplain, Washington, D.C. *Natural Areas Journal* 18: 164–168.
- Thomas, P.A., M. El-Barghathi & A. Polwart. 2011. Biological Flora of the British Isles: *Euonymus europaeus* L. *Journal of Ecology* 99: 345–365.

219

- Thompson, K., Parkinson, J.A., Band, S.R. & Spencer, R.E. 1997. A comparative study of leaf nutrient concentrations in a regional herbaceous flora. *New Phytologist* 136: 679–689.
- Thrift, J.H. 2007. *Effects of white-tailed deer herbivory on forest plant communities*. M.Sc. thesis, Clemson University. Clemson, South Carolina. 107 pages.
- Tibbets, T.J., & F.W. Ewers. 2000. Root pressure and specific conductivity in temperate lianas: exotic *Celastrus orbiculatus* (Celastraceae) vs. native *Vitis riparia* (Vitaceae). *American Journal of Botany* 87:1272–1278.
- Tixier, H., P. Duncan, J. Scehovic, A. Yant, M. Gleizes & M. Lila. 1997. Food selection by European roe deer (*Capreolus capreolus*): effects of plant chemistry, and consequences for the nutritional value of their diets. *Journal of Zoology* 242: 229–245.
- Todd, J.B. 1927. Winter food of cottontail rabbits. *Journal of Mammalogy* 8: 222–228.
- Toll, J.E., T.S. Baskett & C.S. Conaway. 1960. Home range, reproduction, and foods of the swamp rabbit in Missouri. *American Midland Naturalist* 63: 398–412.
- Torgenson, O., & W.H. Pfander. 1971. Cellulose digestibility and chemical composition of Missouri deer foods. *Journal of Wildlife Management* 35: 221–231.
- Trisel, D.E. 1997. *The invasive shrub *Lonicera maackii* (Rupr.) Herder (Caprifoliaceae); factors contributing to its success and its effect on native species*. Dissertation, Miami University. 200 pages.
- **Troels-Smith, J. 1960. Ivy, mistletoe and elm climate indicators - fodder plants. *Danmarks Geologiske Undersøgelse Række IV*, 4 . pages 1–32.
- Tseng, Y-H., sieh C-F. & Hu J-M. 2008. Incidences and ecological correlates of dioecious angiosperms in Taiwan and its outlying Orchid Island. *Botanical Studies* 49: 261–276.
- Tsuji, Y., & S. Takatsuki. 2004. Food habits and home range use of Japanese macaques on an island inhabited by deer. *Ecological Research* 19:381–388.

220

- Turner, K.E., K.A. Cassida & D.P. Belesky. 2009. Finishing lambs and goat kids on pasture. Pages 3–14 in: M.R. Morales (ed.). *Appalachian Workshop and Research Update: Improving Small Ruminant Grazing Practices*. Mountain State University, Beckley, West Virginia.
- Úbeda, X., L.R. Outeiro & M. Sala. 2006. Vegetation regrowth after a differential intensity forest fire in a mediterranean environment, northeast Spain. *Land Degradation & Development* 17: 429–440.
- Ulrey, C.J. 2002. The relationship between soil fertility and the forests of the Southern Appalachian region. Ph.D. dissertation, North Carolina State University, Raleigh. 234 pages.
- *Umucalilar, H.D., N. Gulsen, B. Coskun, A. Hayirli, & H. Dural. 2007. Nutrient composition of mistletoe (*Viscum album*) and its nutritive value for ruminant animals. *Agroforestry Systems* 71: 77–87.
- U.S.D.A. Agricultural Research Service. 1990. U.S.D.A. Plant Hardiness Zone Map. Miscellaneous Publication No 1475. U.S. Government Printing Office, Washington, DC
- U.S.D.A. Agricultural Research Service. 2012. U.S.D.A. Plant Hardiness Zone Map. Published online in cooperation with Oregon State University, at <http://planthardiness.ars.usda.gov/PHZMWeb/>.
- U.S.D.A. Forest Service. 2010. Southern Invasive Plant Maps. Available at http://srsfia2.fs.fed.us/data_center/data_mapping.shtml.
- Uytvanck, J.V., & M. Hoffmann. 2009. Impact of grazing management with large herbivores on forest ground flora and bramble understorey. *Acta Oecologica* 35: 523–532.
- VanDriesche, R.G., P. Kingsley, M. Rose, M. Bryan. 1998. Effect of euonymus scale (Homoptera: Diaspididae) on *Euonymus* spp. survival in southern New England, with estimates of economic costs of pest damage. *Environmental Entomology* 27: 217–220.
- VanDulmen, A. 2001. Pollination and phenology of flowers in the canopy of two contrasting rain forest types in Amazonia, Columbia. *Plant Ecology* 153: 73–85.

221

- VanOmmen-Kloeke, A.E.E., J.C. Douma, J.C. Ordoñez, P.B Reich & P.M. Van Bodegom. 2011. Global quantification of contrasting leaf life span strategies for deciduous and evergreen species in response to environmental conditions. *Global Ecology and Biogeography* 21: 224–235.
- VanWieren, S.E. 1995. The potential role of large herbivores in nature conservation and extensive land use in Europe. *Biological Journal of the Linnean Society* 56, supplement: 11–23.
- Vanguelova E.I., Y. Hirano, T.D. Eldhuset, L. Sas-Paszt, M.R. Bakker & U. Püttsepp. 2007. Tree fine root Ca/Al molar ratio—indicator of Al and acidity stress. *Plant Biosystematics* 141: 460–480.
- Vega-Frutis, R., M.A. Munguía-Rosas, S. Varga, M-M. Kytöviita. 2012. Sex-specific patterns of antagonistic and mutualistic biotic interactions in dioecious and gynodioecious plants. *Perspectives in Plant Ecology, Evolution and Systematics*, in press.
- Vera, F.W.M. 2000. *Grazing Ecology and Forest History*. CABI Publishing, Wallingford, England. 506 pages.
- Vera, F.W.M. 2009. Large-scale nature development—the Oostvaardersplassen. *British Wildlife*, June 2009, pages 28–36.
- Veit, M., L. Jungbauer, K.R. Wendler & E. Zentner. 2011. Effects of phytogetic feed additives containing *Quillaja saponaria* on ammonia in fattening pigs. Pages 1255–1257 in: J. Köfer & H. Schobesberger (eds). *Animal hygiene and sustainable livestock production. Proceedings of the XVth International Congress of the International Society for Animal Hygiene*, Vienna, Austria 3: 1255–1257.
- Verheyden-Tixier, H., & P. Duncan. 2000. Selection for small amounts of hydrolysable tannins by a concentrate selecting mammalian herbivore. *Journal of Chemical Ecology* 26: 351–358.

222

- Verheyen, K., L. Baeten, P. DeFrenne, M. Bernhardt-Römermann, J. Brunet, J. Cornelis, G. Decocq, H. Dierschke, O. Eriksson, R. Hédl, T. Heinken, M. Hermy, P. Hommel, K. Kirby, T. Naaf, G. Peterken, P. Petrik, J. Pfadenhauer, H. VanCalster, G-R. Walther, M. Wulf & G. Verstraeten. 2012. Driving factors behind the eutrophication signal in understory plant communities of deciduous temperate forests. *Journal of Ecology* 100: 352–365.
- Verhoeven, A.S., W.W. Adams & B. Demmig-Adams. 1998. Two forms of sustained xanthophyll cycle-dependent energy dissipation in overwintering *Euonymus kiautschovicus*. *Plant, Cell & Environment* 21: 893–903
- Verloover, F. 2006. Catalogue of neophytes in Belgium (1800–2005). Volume 39 in: E. Robbrecht (ed.). *Scripta Botanica Belgica*. Miscellaneous documentation. Published by the National Botanic Garden, Belgium.
- Villar, A., M.C. Terencio & M. Paya. 1986. Hypotensive effect of *Rhamnus lycioides* extracts. *Journal of Ethnopharmacology* 16: 269–273.
- Vogl, O., & J.D. Mitchell. 1996. Oriental lacquer. 11. Botany and chemistry of the active components of poisonous Anacardiaceae. *Journal of Macromolecular Science, Pure & Applied Chemistry* 33: 1581–1599.
- VonPoser, G.L., J. Schripsema, A.T. Henriques & S.R. Jensen. 2000. The distribution of iridoids in Bignoniaceae. *Biochemical Systematics and Ecology* 28: 351–366.
- Wade, G.L., & M.T. Mengak. 2010. Deer-tolerant ornamental plants. University of Georgia Cooperative Extension & Warnell Circular 285. 8 pages.
- Waggy, M.A. 2010. *Hedera helix*. In: Fire Effects Information System. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fire Sciences Laboratory (Producer). Available at <http://www.fs.fed.us/database/feis/>.
- Wallach, A.D., M. Inbar & U. Shanas. 2009. Roe deer and decapitated *Anemone* flowers. *Israel Journal of Plant Sciences* 57: 103–106.

223

- Wallander, E. 2008. Systematics of *Fraxinus* (Oleaceae) and evolution of dioecy. *Plant Systematics and Evolution* 273: 25–49.
- Wambui, C.C., S. Ando, S.A. Abdulrazak, I.M. Osuga & T. Ichinohe. 2012. In vitro assessment of ruminal fermentation characteristics of tropical browse mixtures supplemented with yeast. *Japanese Society of Grassland Science* 58: 53–57.
- Wamelink, G.W.W., V. Joosten, H.F. VanDobben, & F. Berendse. 2002. Validity of Ellenberg indicator values judged from physico-chemical field measurements. *Journal of Vegetation Science* 13: 269–278.
- **Wang F., Li D-Z. & Yang J-B. 2002. Molecular phylogeny of the Lardizabalaceae based on trnL-F sequence and combined Chloroplast data. *Acta Botanica Sinica* 44: 971–977.
- *Wang G-W., Hu W-T., Huang B-K. & Qin L-P. 2011. *Illicium verum*: A review on its botany, traditional use, chemistry and pharmacology. *Journal of Ethnopharmacology* 136: 10–20.
- Wang H-H., C.L. Wonkka, W.E. Grant & W.E. Rogers. 2012. Potential range expansion of Japanese honeysuckle (*Lonicera japonica* Thunb.) in southern U.S. forestlands. *Forests* 3: 573–590.
- *Wang, Q., D.L. Dilcher, X.Y. Zhu, Y.L. Zhou & T.A. Lott. 2006. Fruit and leaflets of *Wisteria* (Leguminosae, Papilionoideae) from the Miocene of Shandong Province, eastern China. *International Journal of Plant Sciences* 167: 1061–1074.
- *Wang Q., S.R. Manchester & D.L. Dilcher. 2010. Fruits and foliage of *Pueraria* (Leguminosae, Papilionoideae) from the Neogene of Eurasia and their biogeographic implications. *American Journal of Botany* 97: 1982–1998.
- *Wang Z-Q., Wua L-H. & Liu T-T. 2009. Revegetation of steep rocky slopes: Planting climbing vegetation species in artificially drilled holes. *Ecological Engineering* 35: 1079–1084.

224

- Ward, J.S. 2000. Limiting deer browse damage to landscape plants. Bulletin of the Connecticut Agricultural Experiment Station No. 968 pp. 15 pp.
- **Ward, L.K., A. Hackshaw, R.T. Clarke. 1995. Food-plant families of British insects and mites: the influence of life form and plant family. Biological Journal of the Linnean Society 55: 109–127.
- Waters, S.B., R.D. Bray & G.F. Levy. 1974. A taxonomic survey of the spring vascular flora of the Nansemond County, Virginia, portion of the Great Dismal Swamp. Castanea 39: 82–95.
- Watson, D.M. 2001. Mistletoe—A keystone resource in forests and woodlands worldwide. Annual Review of Ecology and Systematics 32: 219–250.
- Weakley, A.S. 2011. Flora of the Southern and Mid-Atlantic States. Working draft of 15 May 2011. University of North Carolina Herbarium, Chapel Hill. Available at <http://www.herbarium.unc.edu/flora.html>.
- Weatherill, R.G., & L.B. Keith. 1969. The effect of livestock grazing on an aspen forest community. Alberta Department of Lands and Forests, Fish and Wildlife Division Technical Bulletin No. 1. Calgary, Alberta.
- Weber E. 2005. *Lonicera henryi* Hemsl.—a potential exotic forest weed in Switzerland. Botanica Helvetica 115: 77–81.
- Webster, C.R., M.A. Jenkins & J.H. Rock. 2005. Twenty years of forest change in the woodlots of Cades Cove, Great Smoky Mountains National Park. Journal of the Torrey Botanical Society 132: 280–292.
- Webster, C.R., J.H. Rock, R.E. Froese, M.A. Jenkins. 2008. Drought–herbivory interaction disrupts competitive displacement of native plants by *Microstegium vimineum*, 10-year results. Oecologia 2008. 157: 497–508.
- Wen, J., & T.F. Stuessy. 1993. Phylogeny and biogeography of *Nyssa* (Cornaceae). Systematic Botany 18: 68–79.

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- Wen J. 1999. Evolution of Eastern Asian and Eastern North American disjunct distributions in flowering plants. Annual Review of Ecology and Systematics 30: 421–455.
- Wen, J. 2001 Evolution of eastern Asian and eastern North American biogeographic disjunctions: a few additional issues. International Journal of Plant Science 162: S117–S122.
- Whitehead, William. 1777. The Goat's Beard. A Fable. Second Edition. J. Dodsley, London.
- Williams, A., J. Antonovics & J. Rolff. 2011. Dioecy, hermaphrodites and pathogen load in plants. Oikos 120: 657–660.
- Williams, A.H. 1997. In praise of grazing. Restoration and Management Notes 15: 116–118.
- Williams, R., & T. Baxley. 2008. Managing native vegetation for wildlife. University of Florida, IFAS Extension Publication FOR186. 5 pages.
- Willson, A. 2006. Forest conversion and land use changes in rural northwest Yunnan, China: implications for the 'big picture'. Mount Research and Development 26: 227–236.
- Wink, M., & O. Schimmer. 2010. Molecular modes of action of defensive secondary metabolites. Chapter 2 in: M. Wink (ed.) Functions and Biotechnology of Plant Secondary Metabolites, Vol. 39, 2nd edition. Annual Plant Reviews. Wiley & Blackwell, Chichester.
- Winkler, D. 1998. The forests of the eastern part of the Tibetan Plateau. A case study from Jiuzhaigou (Zitsa Degu; NNW Sichuan). Forest Management and Sustainability 47:184–210.
- Wirdlechner, M.P. 1997. China Hardiness Zones in China. U.S.D.A.-Agricultural Research Service. North Central Regional Plant Introduction Station, Ames, Iowa, U.S.A.
- Wittig, R., & H. Neite. 1986. Acid indicators around the trunk base of *Fagus sylvatica* in limestone and loess beechwoods: distribution pattern and phytosociological problems. Plant Ecology 64: 113–199.
- Wolfe, J.A. 1987. Late Cretaceous-Cenozoic history of deciduousness and the terminal Cretaceous event. Paleobiology 13: 215–226.

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- Wolters, G.L., A. Martin & W.P. Clary. 1977. Timber, browse, and herbage on selected loblolly-shortleaf pine-hardwood forest stands. USDA Forest Service Publ. SO-223. 9 pages.
- Woodcock D.W. 1994. Occurrence of woods with a gradation in vessel diameter across a ring. International Association of Wood Anatomists Journal 15: 377–385.
- Wright, K. & K. Long. 2002. Browse preferences of white-tailed deer (*Odocoileus virginianus*) at UNDERC, Vilas/Gogebic Counties, Wisconsin/Michigan. Student Paper (advisor G. Belovsky), University of Michigan, Ann Arbor. 10 pages.
- Wright, I.J. et al. [GLOPNET]. 2004 The world-wide leaf economics spectrum. Nature 428: 821–827.
- Wright, I.J., P.B. Reich, J.H.C. Cornelissen, D.S. Falster, P.K. Groom, K. Hikosaka, W. Lee, C.H. Lusk, U. Niinemets, J. Oleksyn, N. Osada, H. Poorter, D.I. Warton & M. Westoby. 2005. Modulation of leaf economic traits and trait relationships by climate. Global Ecology and Biogeography 14: 411–421.
- Wright, R.G. 1999. Wildlife management in the National Parks: questions in search of answers. Ecological Applications 9: 30–36
- **Xia H-P., Cai X-A. & Peng C-X. 2007. Comparison of five lianas for vertical greening. Acta Prataculturae Sinica 2007-03.
- Xiang Q-Y., D.E. Soltis, P.S. Soltis, S.R. Manchester & D.J. Crawford. 2000. Timing the Eastern Asian-Eastern North American floristic disjunction: molecular clocks confirm paleontological estimates. Molecular Phylogenetics and Evolution 15: 462–472.
- Xiang Q-Y., D.T. Thomas & Xiang Q-P. 2011. Resolving and dating the phylogeny of Cornales—effects of taxon sampling, data partitions, and fossil calibrations. Molecular Phylogenetics and Evolution 59: 123–138.

227

- Xiang, Z-F., Huo S., Xiao W., Quan R-C. & C.C. Grueter. 2007. Diet and feeding behavior of *Rhinopithecus bieti* at Xiaochangdu, Tibet: adaptations to a marginal environment. American Journal of Primatology 69: 1–18.
- **Xiao Z., Wang Y-P., Wang X-W., Xing W-B., He S-Q. & Zhang Q-L. 2010. Structural botany characteristics of *Actinidia arguta* and *Actinidia kolomikta* in the Changbai Mountains. Journal of Northwest Normal University (Natural Science) 2010-05.
- Xie L., Wen J. & Li L-Q. 2011. Phylogenetic analyses of *Clematis* (Ranunculaceae) based on sequences of nuclear ribosomal ITS and three plastid regions. Systematic Botany 36: 907–921.
- **Yan L-H. 2007. Sexual system and environmental adaptability of vines in Hupingshan Mountain, Hunan Province. Journal of Northeast Forestry University 2007-07.
- Yang, J., Cai Y-L., Sun S-C. & Wang. L. 2006. Eco-physiological modelling of leaf photosynthesis and adaptation analysis of Chinese ivy (*Hedera nepalensis* var. *sinensis*) in an evergreen broad-leaved forest in eastern China. Photosynthetica 44: 579-585.
- *Yao, X., G. Gongyu & G. Chen. 2006. Determination of active constituents in *Lonicera confusa* DC. by capillary electrophoresis with amperometric detection. Biomedical Chromatography 20: 1192-9.
- *Ying D-Y., Guang Z-Y., Mao C-L. & Zhi N-M. 2011. Three new sesquiterpene pyridine alkaloids from *Euonymus fortunei*. Helvetica Chimica Acta 94: 1139–1145.
- Yokoyama, M., K. Kaji & M. Suzuki. 2000. Food habits of sika deer and nutritional value of sika deer diets in eastern Hokkaido, Japan. Ecological Research 15:345–355.
- Young, S., R.H. Simmons & C. Hamblin-Katnik. 2012. Instructions for removing English ivy and discussion of safety. City of Alexandria Dept. Recreation, Parks, and Cultural Activities. Alexandria, Virginia. Available at <http://alexandriava.gov/48838>.

228

- Yuan C-M, Liu W-Y., Tang C-Q. & Li X-S. 2009. Species composition, diversity, and abundance of lianas in different secondary and primary forests in a subtropical mountainous area, SW China. *Ecological Research* 24: 1361–1370
- Zanne, A.E., K. Sweeney, M. Sharma & C.M. Orians. 2006. Patterns and consequences of differential vascular sectoriality in 18 temperate tree and shrub species. *Functional Ecology* 20: 200–206.
- Zaya, D. & Howe H. F. 2009. The anomalous Kentucky Coffee Tree: megafaunal fruit sinking to extinction? *Oecologia* 161: 221–226.
- Zbidah, M., A. Lupescu, K. Jilani, A. Fajol, D. Michael, S.M. Qadri & F. Lang. 2012. Apigenin-induced suicidal erythrocyte death. *Journal of Agriculture and Food Chemistry* 60: 533–538.
- Zerega, N.J.C., W.L. Clement, S.L. Datwyler, G.D. Weiblen. 2005. Biogeography and divergence times in the mulberry family (Moraceae). *Molecular Phylogenetics and Evolution* 37: 402–416.
- Zecca, G., J.R. Abbott, Sun W-B., A. Spada, F. Sala & F. Grassi. 2012. The timing and the mode of evolution of wild grapes (*Vitis*). *Molecular Phylogenetics & Evolution* 62: 736–747.
- *Zhang C., Yin Z., Ye W., Guo L., Zhang J. & Shen W. 2009. Chemical constituents from stems of *Lonicera japonica*. *China Journal of Chinese Materia Medica* 34: 3051–3053.
- *Zhang C-L., Feng S-X., Zhang L-X., & Ren Z-J. 2012. A new cytotoxic steroidal saponin from the rhizomes and roots of *Smilax scobinicaulis*. *Natural Product Research* 1: 1–6.
- Zhang Y., J.L. Hanula & S. Horn. 2012. The biology and preliminary host range of *Megacopta cribraria* (Heteroptera: Plataspidae) and its impact on kudzu growth. *Environmental Entomology* 41:40–50.
- Zotz., G., N. Cueni & C. Körner. 2006. *In situ* growth stimulation of a temperate zone liana (*Hedera helix*) in elevated CO₂. *Functional Ecology* 20: 763–769

Zouhar, K. 2009. *Euonymus fortunei*. In: Fire Effects Information System. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fire Sciences Laboratory (Producer). Available at <http://www.fs.fed.us/database/feis/>. 27 pages.



APPENDIX ONE

Six-letter abbreviations for woody species used in Figures 2 and 3

T1 = large tree; T2 = small tree; S1 = large shrub; S2 = small shrub; V1 = large vine; V2 = small vine. See Campbell & Medley (2012) and Weakley (2012) for distributions, nomenclature, synonyms, etc.

Abbrev	Full Binomial Name	LF	Abbrev	Full Binomial Name	LF
Aceflo	<i>Acer floridanum</i> (Chapman) Pax	T1	Aruapp	<i>Arundinaria appalachiana</i> Triplett, Weakley, & L.G. Clark	S2
Aceneg	<i>Acer negundo</i> L.	T1	Arugig	<i>Arundinaria gigantea</i> (Walt.) Muhl.	S2
Acenig	<i>Acer nigrum</i> Michx. f.	T1	Arutec	<i>Arundinaria tecta</i> (Walt.) Muhl.	S2
Acepen	<i>Acer pensylvanicum</i> L.	T2	Asitri	<i>Asimina triloba</i> (L.) Dunal	T2
Acepla	<i>Acer platanoides</i> L.	T1	Bersca	<i>Berchemia scandens</i> (Hill) K. Koch	V2
Acertr	<i>Acer rubrum</i> L. var. <i>trilobum</i> Torr. & Gray ex K. Koch	T1	Betall	<i>Betula alleghaniensis</i> Britt.	T1
Acerub	<i>Acer rubrum</i> L. var. <i>rubrum</i>	T1	Betlen	<i>Betula lenta</i> L.	T1
Acesac	<i>Acer saccharum</i> Marsh. var. <i>saccharum</i>	T1	Betnig	<i>Betula nigra</i> L.	T1
Acesch	<i>Acer saccharum</i> Marsh. var. <i>schneckii</i> Rehd.	T1	Bigcap	<i>Bignonia capreolata</i> L.	V1
Acesnm	<i>Acer saccharinum</i> L.	T1	Brbcan	<i>Berberis canadensis</i> P. Mill.	S2
Acespi	<i>Acer spicatum</i> Lam.	T2	Bropap	<i>Broussonetia papyrifera</i> (L.) L'Hér. ex Vent.	T1
Aesfla	<i>Aesculus flava</i> Ait.	T1	Calflo	<i>Calycanthus floridus</i> L. var. <i>glaucus</i> (Willd.) Torr. & Gray	S1
Aesgla	<i>Aesculus glabra</i> Willd.	T1	Callyo	<i>Calycocarpum lyonii</i> (Pursh) Gray	V2
Aespav	<i>Aesculus pavia</i> L.	T2	Camrad	<i>Campsis radicans</i> (L.) Seem. ex Bureau	V1
Ailalt	<i>Ailanthus altissima</i> (P. Mill.) Swingle	T1	Caraqu	<i>Carya aquatica</i> (Michx. f.) Nutt.	T1
Albjul	<i>Albizia julibrissin</i> Durazz.	T1	Carcar	<i>Carya caroliniae-septentrionalis</i> (Ashe) Engl. & Graebn.	T1
Alnser	<i>Alnus serrulata</i> (Ait.) Willd.	S1	Carcor	<i>Carya cordiformis</i> (Wangenh.) K. Koch	T1
Amearb	<i>Amelanchier arborea</i> (Michx. f.) Fern.	T2	Cargla	<i>Carya glabra</i> (P. Mill.) Sweet	T1
Amelae	<i>Amelanchier laevis</i> Wieg.	T2	Carill	<i>Carya illinoensis</i> (Wangenh.) K. Koch	T1
Amesan	<i>Amelanchier sanguinea</i> (Pursh) DC.	S2	Carlac	<i>Carya laciniosa</i> (Michx. f.) G. Don	T1
Amespi	<i>Amelanchier spicata</i> (Lam.) K. Koch	S2	Carova	<i>Carya ovata</i> (P. Mill.) K. Koch	T1
Amocro	<i>Amorpha croceolanata</i> Wats.	S2	Carovl	<i>Carya X ovalis</i> (Wangenh.) Sarg.	T1
Amofru	<i>Amorpha fruticosa</i> L.	S1	Carpall	<i>Carya pallida</i> (Ashe) Engl. & Graebn.	T1
Amonit	<i>Amorpha nitens</i> Boynt.	S2	Cartom	<i>Carya tomentosa</i> (Lam. ex Poir.) Nutt.	T1
Amparb	<i>Ampelopsis arborea</i> (L.) Koehne	V1	Casden	<i>Castanea dentata</i> (Marsh.) Borkh.	T1
Ampcor	<i>Ampelopsis cordata</i> Michx.	V1	Caspum	<i>Castanea pumila</i> (L.) P. Mill.	T2
Araspi	<i>Aralia spinosa</i> L.	T2	Catbig	<i>Catalpa bignonioides</i> Walt.	T1
Aroarb	<i>Aronia arbutifolia</i> (L.) Pers.	S1	Catspe	<i>Catalpa speciosa</i> (Warder) Warder ex Engelm.	T1
Aromel	<i>Aronia melanocarpa</i> (Michx.) Ell.	S1	Ceame	<i>Ceanothus americanus</i> L.	S2
			Cellae	<i>Celtis laevigata</i> Willd.	T1

Celocc	<i>Celtis occidentalis</i> L.	T1	Crapun	<i>Crataegus punctata</i> Jacq.	T2
Celten	<i>Celtis tenuifolia</i> Nutt.	T2	Craspa	<i>Crataegus spathulata</i> Michx.	T2
Cepocc	<i>Cephalanthus occidentalis</i> L.	S1	Crasuc	<i>Crataegus succulenta</i> Schrad. ex Link	T2
Cercan	<i>Cercis canadensis</i> L.	T2	Crauni	<i>Crataegus uniflora</i> Muenchh.	T2
Chivir	<i>Chionanthus virginicus</i> L.	T2	Cravir	<i>Crataegus viridis</i> L.	T2
Claken	<i>Cladrastis kentukea</i> (Dum.-Cours.) Rudd	T1	Crflame	<i>Corylus americana</i> Walt.	S1
Cleacu	<i>Clethra acuminata</i> Michx.	S1	Crpcar	<i>Carpinus caroliniana</i> Walt.	T2
Clecat	<i>Clematis catesbyana</i> Pursh	V2	Cyrrac	<i>Cyrilla racemiflora</i> L.	T2
Cleter	<i>Clematis terniflora</i> DC.	V2	Decver	<i>Decodon verticillatus</i> (L.) Ell.	S2
Clevir	<i>Clematis virginiana</i> L.	V2	Diopub	<i>Diospyros virginiana</i> L. var. <i>virginiana</i>	T1
Clorb	<i>Celastrus orbiculatus</i> Thunb.	V2	Diovir	<i>Diospyros virginiana</i> L. var. <i>pubescens</i> (Pursh) Dippel	T1
Clssca	<i>Celastrus scandens</i> L.	V2	Dirpal	<i>Dirca palustris</i> L.	S2
Coccar	<i>Cocculus carolinus</i> (L.) DC.	V2	Elamul	<i>Elaeagnus multiflora</i> Thunb.	T2
Comper	<i>Comptonia peregrina</i> (L.) Coult.	S2	Eleumb	<i>Elaeagnus umbellata</i> Thunb.	S1
Coralt	<i>Cornus alternifolia</i> L. f.	S1	Eubrec	<i>Eubotrys recurva</i> (Buckl.) Britt.	S2
Coramm	<i>Cornus amomum</i> P. Mill.	S1	Euoame	<i>Euonymus americanus</i> L.	S2
Cordru	<i>Cornus drummondii</i> C.A. Mey.	S1	Euoafr	<i>Euonymus atropurpureus</i> Jacq.	S1
Corflo	<i>Cornus florida</i> L.	T2	Euofor	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz.	V1
Corobl	<i>Cornus obliqua</i> Raf.	S1	Faggra	<i>Fagus grandifolia</i> Ehrh.	T1
Corrac	<i>Cornus racemosa</i> Lam.	S1	Foracu	<i>Forestiera acuminata</i> (Michx.) Poir.	S1
Corsto	<i>Cornus stolonifera</i> Michx.	S1	Forlig	<i>Forestiera ligustrina</i> (Michx.) Poir.	T2
Corstr	<i>Cornus stricta</i> Lam.	S1	Fraame	<i>Fraxinus americana</i> L.	T1
Cracal	<i>Crataegus calpodendron</i> (Ehrh.) Medik.	T2	Frabil	<i>Fraxinus biltmoreana</i> Beadle	T1
Crachr	<i>Crataegus chrysoarpa</i> Ashe ?	T2	Frapen	<i>Fraxinus pennsylvanica</i> Marsh var. <i>pennsylvanica</i>	T1
Cracoc	<i>Crataegus coccinea</i> L. ?	T2	Frasub	<i>Fraxinus pennsylvanica</i> Marsh. var. <i>subintegerrima</i> (-) Fern.	T1
Cracol	<i>Crataegus collina</i> Chapman	T2	Frapro	<i>Fraxinus profunda</i> (Bush) Bush	T1
Cracru	<i>Crataegus crus-galli</i> L.	T2	Fraqua	<i>Fraxinus quadrangulata</i> Michx.	T1
Craeng	<i>Crataegus engelmannii</i> Sarg.	T2	Frasma	<i>Fraxinus smallii</i> Beadle	T1
Cragat	<i>Crataegus gattingeri</i> Ashe	T2	Frncar	<i>Frangula caroliniana</i> (Walt.) Gray	T2
Crainc	<i>Crataegus incaeua</i> Sarg. ?	T2	Gaybac	<i>Gaylussacia baccata</i> (Wangenh.) K. Koch	S2
Craint	<i>Crataegus intricata</i> Lange	T2	Gaybra	<i>Gaylussacia brachycera</i> Michx.	S2
Craira	<i>Crataegus iracunda</i> Beadle	T2	Gleaqu	<i>Gleditsia aquatica</i> Marsh.	T1
Cramac	<i>Crataegus macrosperma</i> Ashe	T2	Gletri	<i>Gleditsia triacanthos</i> L.	T1
Cramar	<i>Crataegus marshallii</i> Egglest.	T2	Gymdio	<i>Gymnocladus dioica</i> (L.) K. Koch	T1
Cramol	<i>Crataegus mollis</i> Scheele	T2	Haltet	<i>Halesia tetraptera</i> Ellis	T1
Crafer	<i>Crataegus persimilis</i> Sarg. ?	T2	Hamvir	<i>Hamamelis virginiana</i> L.	S1
Crapru	<i>Crataegus pruinosa</i> (Wendl. f.) K. Koch	T2	Hedhel	<i>Hedera helix</i> L.	V1

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Humlup	<i>Humulus lupulus</i> L.	V2	Malpum	<i>Malus pumila</i> P. Mill.	T1
Hydarb	<i>Hydrangea arborescens</i> L.	S2	Mencan	<i>Menispermum canadense</i> L.	V2
Hyplob	<i>Hypericum lobocarpum</i> Gattinger	S2	Moralb	<i>Morus alba</i> L.	T1
Hypro	<i>Hypericum prolificum</i> L.	S2	Morrub	<i>Morus rubra</i> L.	T1
Iledec	<i>Ilex decidua</i> Walt.	S1	Nesumb	<i>Nestronia umbellula</i> Raf.	S2
Ilemon	<i>Ilex montana</i> Torr. & Gray ex Gray var. <i>montana</i>	S1	Nysaqu	<i>Nyssa aquatica</i> L.	T1
Ileopa	<i>Ilex opaca</i> Ait.	T2	Nysbif	<i>Nyssa biflora</i> Walt.	T1
Ilever	<i>Ilex verticillata</i> (L.) Gray	S1	Nyssyl	<i>Nyssa sylvatica</i> Marsh.	T1
Isomac	<i>Isotrema macrophylla</i> (Lam.) C.F. Reed	V2	Ostvir	<i>Ostrya virginiana</i> (P. Mill.) K. Koch	T2
Isotom	<i>Isotrema tomentosa</i> (Sims) Huber	V2	Oxyden	<i>Oxydendrum arboreum</i> (L.) DC.	T2
Itevir	<i>Itea virginica</i> L.	S2	Parqui	<i>Parthenocissus quinquefolia</i> (L.) Planch.	V1
Jugcin	<i>Juglans cinerea</i> L.	T1	Perpal	<i>Persea palustris</i> (Raf.) Sarg.	T2
Jugnig	<i>Juglans nigra</i> L.	T1	Phihir	<i>Philadelphus hirsutus</i> Nutt.	S2
Juncom	<i>Juniperus communis</i> L.	S1	Phiint	<i>Philadelphus intectus</i> Beadle ?	S1
Junvir	<i>Juniperus virginiana</i> L.	T1	Phipub	<i>Philadelphus pubescens</i> Loisel.	S2
Kalbux	<i>Kalmia buxifolia</i> (Berg.) Gift, Kron, & Stevens	S2	Pholeu	<i>Phoradendron leucarpum</i> (Raf.) Reveal & M.C. Johnston	V0
Kallat	<i>Kalmia latifolia</i> L.	S1	Phyopu	<i>Phyocarpus opulifolius</i> (L.) Maxim.	S2
Lacmul	<i>Lackeya multiflora</i> (Torr. & Gray) Fortunato, Queiroz & Lewis	V2	Pinech	<i>Pinus echinata</i> P. Mill.	T1
Leufon	<i>Leucothoe fontanesiana</i> (Steud.) Sleumer	S2	Pinell	<i>Pinus elliottii</i> Engelm.	T1
Linben	<i>Lindera benzoin</i> (L.) Blume	S1	Pinpal	<i>Pinus palustris</i> P. Mill.	T1
Liqsty	<i>Liquidambar styraciflua</i> L.	T1	Pinpun	<i>Pinus pungens</i> Lam.	T1
Lirtul	<i>Liriodendron tulipifera</i> L.	T1	Pinrig	<i>Pinus rigida</i> P. Mill.	T1
Londio	<i>Lonicera dioica</i> L.	V2	Pinser	<i>Pinus serotina</i> Michx.	T1
Lonpro	<i>Lonicera prolifera</i> (Kirchn.) Rehd.	V2	Pinstr	<i>Pinus strobus</i> L.	T1
Lonsem	<i>Lonicera sempervirens</i> L.	V2	Pintae	<i>Pinus taeda</i> L.	T1
Lyolig	<i>Lyonia ligustrina</i> (L.) DC.	S1	Pinvir	<i>Pinus virginiana</i> P. Mill.	T1
Macpom	<i>Maclura pomifera</i> (Raf.) Schneid.	T1	Plaaqu	<i>Planera aquatica</i> J.F. Gmel.	T2
Magacu	<i>Magnolia acuminata</i> (L.) L.	T1	Plaocc	<i>Platanus occidentalis</i> L.	T1
Magfra	<i>Magnolia fraseri</i> Walt.	T1	Popdel	<i>Populus deltoides</i> Bartr. ex Marsh.	T1
Magmac	<i>Magnolia macrophylla</i> Michx.	T1	Popgra	<i>Populus grandidentata</i> Michx.	T1
Magpyr	<i>Magnolia pyramidata</i> Bartr.	T1	Pophet	<i>Populus heterophylla</i> L.	T1
Magtri	<i>Magnolia tripetala</i> (L.) L.	T1	Pruame	<i>Prunus americana</i> Marsh.	T2
Magvir	<i>Magnolia virginiana</i> L.	T2	Pruang	<i>Prunus angustifolia</i> Marsh.	T2
Malang	<i>Malus angustifolia</i> (Ait.) Michx.	T2	Pruavi	<i>Prunus avium</i> (L.) L.	T1
Malbac	<i>Malus baccata</i> (L.) Borkh.	T2	Pruhor	<i>Prunus hortulana</i> Bailey	T2
Malcor	<i>Malus coronaria</i> (L.) P. Mill.	T2	Prumah	<i>Prunus mahaleb</i> L.	T2
Malioe	<i>Malus ioensis</i> (Wood) Britt.	T2	Prumex	<i>Prunus mexicana</i> S. Wats.	T2

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Prumun	<i>Prunus munsoniana</i> W. Wight & Hedrick	T2	Rhalan	<i>Rhamnus lanceolata</i> Pursh	S1
Pruser	<i>Prunus serotina</i> Ehrh.	T1	Rhoarb	<i>Rhododendron arborescens</i> (Pursh) Torr.	S1
Pruvir	<i>Prunus virginiana</i> L.	S1	Rhocal	<i>Rhododendron calendulaceum</i> (Michx.) Torr.	S1
Ptetri	<i>Ptelea trifoliata</i> L.	S1	Rhocal	<i>Rhododendron canadense</i> (Michx.) Sweet	S1
Puemon	<i>Pueraria montana</i> (Lour.) Merr.	V2	Rhopat	<i>Rhododendron catawbiense</i> Michx.	S1
Pyrca	<i>Pyrus calleryana</i> Dcne.	T1	Rhocum	<i>Rhododendron cumberlandense</i> E.L. Braun	S1
Pyrcom	<i>Pyrus communis</i> L.	T1	Rhomax	<i>Rhododendron maximum</i> L.	T2
Pyrpub	<i>Pyralia pubera</i> Michx.	S1	Rhomin	<i>Rhododendron minus</i> Michx.	S1
Quealb	<i>Quercus alba</i> L.	T1	Rhoper	<i>Rhododendron periclymenoides</i> (Michx.) Shinnors	S1
Quebic	<i>Quercus bicolor</i> Willd.	T1	Rhopri	<i>Rhododendron prinophyllum</i> (Small) Millais	S1
Quecoc	<i>Quercus coccinea</i> Muenchh.	T1	Rhuaro	<i>Rhus aromatica</i> Ait.	S2
Quefal	<i>Quercus falcata</i> Michx.	T1	Rhucop	<i>Rhus copallinum</i> L. var. <i>latifolia</i> Engl.	S1
Quehem	<i>Quercus hemisphaerica</i> Bartram ex Willd.	T1	Rhugla	<i>Rhus glabra</i> L.	S1
Queili	<i>Quercus ilicifolia</i> Wangerh.	T2	Rhutyp	<i>Rhus typhina</i> L.	T2
Queimb	<i>Quercus imbricaria</i> Michx.	T1	Ribame	<i>Ribes americanum</i> P. Mill.	S2
Queinc	<i>Quercus incana</i> Bartram	T2	Ribcyn	<i>Ribes cynosbati</i> L.	S2
Quelae	<i>Quercus laevis</i> Walt.	T2	Ribmis	<i>Ribes missouriense</i> Nutt.	S2
Quelau	<i>Quercus laurifolia</i> Michx.	T1	Robboy	<i>Robinia boyntonii</i> Ashe	S2
Quelyr	<i>Quercus lyrata</i> Walt.	T1	Robhis	<i>Robinia hispida</i> L.	S2
Quemac	<i>Quercus macrocarpa</i> Michx.	T1	Robpse	<i>Robinia pseudoacacia</i> L.	T1
Quemar	<i>Quercus marilandica</i> Muenchh.	T1	Roscar	<i>Rosa carolina</i> L.	S2
Quemic	<i>Quercus michauxii</i> Nutt.	T1	Rospal	<i>Rosa palustris</i> Marsh.	S2
Quemon	<i>Quercus montana</i> Willd.	T1	Rosset	<i>Rosa setigera</i> Michx.	S2
Quemrg	<i>Quercus margareta</i> Ashe ex Small	T2	Ruball	<i>Rubus allegheniensis</i> Porter	S2
Quemue	<i>Quercus muehlenbergii</i> Engelm.	T1	Rubalu	<i>Rubus alumnus</i> Bailey	S2
Quenig	<i>Quercus nigra</i> L.	T1	Rubarg	<i>Rubus argutus</i> Link	S2
Quepag	<i>Quercus pagoda</i> Raf.	T1	Rubcan	<i>Rubus canadensis</i> L.	S2
Quepal	<i>Quercus palustris</i> Muenchh.	T1	Rubdep	<i>Rubus depavitus</i> Bailey	S2
Quephe	<i>Quercus phellos</i> L.	T1	Rubfro	<i>Rubus frondosus</i> Bigelow	S2
Quepri	<i>Quercus prinoides</i> Willd.	T2	Rublau	<i>Rubus laudatus</i> Berger	S2
Querub	<i>Quercus rubra</i> L.	T1	Rubocc	<i>Rubus occidentalis</i> L.	S2
Quesch	<i>Quercus shumardii</i> Buckl. var. <i>schneckii</i> (Britt.) Sarg.	T1	Rubodo	<i>Rubus odoratus</i> L.	S2
Queshu	<i>Quercus shumardii</i> Buckl.	T1	Rubpen	<i>Rubus pensilvanicus</i> Poir.	S2
Quesin	<i>Quercus sinuata</i> Walt.	T1	Rubror	<i>Rubus roribaccus</i> (Bailey) Rydb.	S2
Queste	<i>Quercus stellata</i> Wangerh.	T1	Salamy	<i>Salix amygdaloides</i> Anderss.	T2
Quetex	<i>Quercus texana</i> Buckl.	T1	Salcar	<i>Salix caroliniana</i> Michx.	S1
Quevel	<i>Quercus velutina</i> Lam.	T1	Saldis	<i>Salix discolor</i> Muhl.	S1

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Saleri	<i>Salix eriocephala</i> Michx.	S1	Ulmrub	<i>Ulmus rubra</i> Muhl.	T1
Salhum	<i>Salix humilis</i> Marsh.	S2	Ulmser	<i>Ulmus serotina</i> Sarg.	T1
Salint	<i>Salix interior</i> Rowlee	T2	Ulmtho	<i>Ulmus thomasi</i> Sarg.	T1
Salnig	<i>Salix nigra</i> Marsh.	T1	Vacarb	<i>Vaccinium arboreum</i> Marsh.	S1
Salocc	<i>Salix occidentalis</i> Walt.	S2	Vaccoc	<i>Vaccinium constablaei</i> Gray	S1
Salser	<i>Salix sericea</i> Marsh.	S1	Vaccor	<i>Vaccinium corymbosum</i> L.	S1
Samcan	<i>Sambucus canadensis</i> L.	S1	Vacery	<i>Vaccinium erythrocarpum</i> Michx.	S2
Sampub	<i>Sambucus pubens</i> Michx.	S1	Vacfus	<i>Vaccinium fuscum</i> Ait.	S1
Sasalb	<i>Sassafras albidum</i> (Nutt.) Nees	T1	Vacpal	<i>Vaccinium pallidum</i> Ait.	S2
Sidlyc	<i>Sideroxylon lycioides</i> L.	T2	Vacsim	<i>Vaccinium simulatum</i> Small	S1
Smibon	<i>Smilax bona-nox</i> L.	V1	Vacsta	<i>Vaccinium stamineum</i> L.	S2
Smigla	<i>Smilax glauca</i> Walt.	V1	Vibace	<i>Viburnum acerifolium</i> L.	S1
Smihis	<i>Smilax hispida</i> Muhl. ex Torr.	V1	Vibcas	<i>Viburnum cassinoides</i> L.	S1
Smirot	<i>Smilax rotundifolia</i> L.	V1	Vibdea	<i>Viburnum deamii</i> Rehd.	S1
Spialb	<i>Spiraea alba</i> Du Roi	S2	Viblan	<i>Viburnum lantanoides</i> Michx.	S1
Spitom	<i>Spiraea tomentosa</i> L.	S2	Vibmol	<i>Viburnum molle</i> Michx.	S1
Sprvir	<i>Spiraea virginiana</i> Britt.	S1	Vibnud	<i>Viburnum nudum</i> L.	S1
Statri	<i>Staphylea trifolia</i> L.	S1	Vibpru	<i>Viburnum prunifolium</i> L.	T2
Steova	<i>Stewartia ovata</i> (Cav.) Weatherby	T2	Vibra	<i>Viburnum rafinesquianum</i> J.A. Schultes var. <i>affine</i> (-) House	S1
Styame	<i>Styrax americanus</i> Lam.	S2	Vibrec	<i>Viburnum recognitum</i> Fern.	S1
Symalb	<i>Symphoricarpos albus</i> (L.) Blake	S2	Vibruf	<i>Viburnum rufidulum</i> Raf.	T2
Symorb	<i>Symphoricarpos orbiculatus</i> Moench	S2	Vitaes	<i>Vitis aestivalis</i> Michx. var. <i>aestivalis</i>	V1
Taxasc	<i>Taxodium ascendens</i> Brong.	T1	Vitbai	<i>Vitis baileyana</i> Munson	V1
Taxdis	<i>Taxodium distichum</i> (L.) L.C. Rich.	T1	Vitcin	<i>Vitis cinerea</i> (Engelm.) Millard	V1
Thuocc	<i>Thuja occidentalis</i> L.	T1	Vitlab	<i>Vitis labrusca</i> L.	V1
Thydif	<i>Thyrsanthella difforme</i> (Walter) Pichon	V2	Vitpal	<i>Vitis palmata</i> Vahl	V1
Tilame	<i>Tilia americana</i> L.	T1	Vitrip	<i>Vitis riparia</i> Michx.	V1
Tilhet	<i>Tilia heterophylla</i> Vent.	T1	Vitrot	<i>Vitis rotundifolia</i> Michx.	V1
Toxrad	<i>Toxicodendron radicans</i> (L.) Kuntze	V1	Vitrop	<i>Vitis rupestris</i> Scheele	V2
Toxver	<i>Toxicodendron vernix</i> (L.) Kuntze	S1	Vitvul	<i>Vitis vulpina</i> L.	V1
Tsucan	<i>Tsuga canadensis</i> (L.) Carr.	T1	Wismac	<i>Wisteria macrostachya</i> (Torr. & Gray) Nutt. ex Rbns. & Fern.	V2
Txscan	<i>Taxus canadensis</i> Marsh.	S2	Xansim	<i>Xanthorhiza simplicissima</i> Marsh.	S2
Ulmala	<i>Ulmus alata</i> Michx.	T1	Zaname	<i>Zanthoxylum americanum</i> P. Mill.	S1
Ulmame	<i>Ulmus americana</i> L.	T1			
Ulpum	<i>Ulmus pumila</i> L.	T1			

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APPENDIX TWO

List of plots in VegBank with occurrences of evergreen vines for Figure 2 (Dec 2012).

Figure 2c: *Hedera helix*

- #1 <http://vegbank.org/cite/VB.Ob.4058.ROCR53>
- #2 <http://vegbank.org/cite/VB.Ob.4059.ROCR54>
- #3 <http://vegbank.org/cite/VB.Ob.4066.ROCR61>
- #4 <http://vegbank.org/cite/VB.Ob.3930.ROCR24>
- #5 <http://vegbank.org/cite/VB.Ob.4056.ROCR51>
- #6 <http://vegbank.org/cite/VB.Ob.4057.ROCR52>

Figure 2d: *Euonymus fortunei*

- # 1 <http://vegbank.org/cite/VB.Ob.4056.ROCR51>

Figure 2e: *Bignonia capreolata*

- # 1 <http://vegbank.org/cite/VB.Ob.25891.COSW115>
- #2 <http://vegbank.org/cite/VB.Ob.25992.COSW212>
- #3 <http://vegbank.org/cite/VB.Ob.25920.COSW248>
- #4 <http://vegbank.org/cite/VB.ob.26507.CHAT43>
- #5 <http://vegbank.org/cite/VB.Ob.25985.COSW205>
- #? see *Lonicera japonica* #14 and *Smilax bona-nox* # 8;

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Figure 2f: *Lonicera japonica*

- #1 <http://vegbank.org/cite/VB.Ob.3623.HAFE11>
- #2 <http://vegbank.org/cite/VB.Ob.26027.027020553>
- #3 <http://vegbank.org/cite/urn:lsid:cvs.bio.unc.edu:observation:7349-{\CCCB9791-7B32-4428-B3D9-4F5BCB0FA01D}>
- #4 <http://vegbank.org/cite/VB.Ob.26114.027090002>
- #5 <http://vegbank.org/cite/VB.Ob.4081.ROCR79>
- #6 <http://vegbank.org/cite/urn:lsid:cvs.bio.unc.edu:observation:8161-{\7102FDB7-0DA9-4A1B-8D1C-7F81A39B2769}>
- #7 <http://vegbank.org/cite/VB.Ob.4727.VAFO60>
- #8 <http://vegbank.org/cite/VB.Ob.26108.027080400>
- #9 <http://vegbank.org/cite/VB.Ob.3553.HAFE21>
- #10 <http://vegbank.org/cite/VB.Ob.26028.027020554>
- #11 <http://vegbank.org/cite/urn:lsid:cvs.bio.unc.edu:observation:8197-{\A00CBEE5-2994-4DDC-9E2F-3A25B2AEC71E}>
- #12 <http://vegbank.org/cite/urn:lsid:cvs.bio.unc.edu:observation:8162-{\691FD52B-FE14-4293-8842-BA2CC63134C3}>
- #13 <http://vegbank.org/cite/VB.Ob.4699.VAFO32>
- #14 <http://vegbank.org/cite/VB.ob.27093.HOLL2>; also has Bigcap
- #15 <http://vegbank.org/cite/VB.Ob.4711.VAFO44>
- #16 <http://vegbank.org/cite/VB.Ob.4069.ROCR66>
- #17 <http://vegbank.org/cite/VB.Ob.3952.ROCR74>

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Figure 2g: *Lonicera sempervirens*.

With zero plot data, typical associates are gleaned from miscellaneous literature; see also caption to Figure 1 for some sources.

Figure 2h: *Smilax bona-nox*.

- #1 <http://vegbank.org/cite/VB.ob.26361.CHAT96>
- #2 <http://vegbank.org/cite/VB.ob.26694.CHAT38>
- #4 <http://vegbank.org/cite/VB.ob.27160.HOLL22>
- #5 <http://vegbank.org/cite/VB.ob.26497.BANK37>
- #6 <http://vegbank.org/cite/VB.ob.26751.CHAT51>
- #7 <http://vegbank.org/cite/VB.Ob.25961.COSW121>
- #8 <http://vegbank.org/cite/VB.Ob.25893.COSW117>; also has Bigcap
- #9 <http://vegbank.org/cite/VB.Ob.25992.COSW212>

Figure 2i: *Smilax glauca*.

- #1 <http://vegbank.org/cite/VB.Ob.4565.FIIS44>
- #2 <http://vegbank.org/cite/VB.Ob.4566.FIIS45>
- #3 <http://vegbank.org/cite/VB.ob.26485.CHER29>
- #4 <http://vegbank.org/cite/VB.ob.26474.CHAT3>
- #5 <http://vegbank.org/cite/VB.ob.26688.CHAT32>
- #6 <http://vegbank.org/cite/VB.ob.26505.CHAT41>
- #7 <http://vegbank.org/cite/VB.ob.26654.BANK61>
- #8 <http://vegbank.org/cite/VB.ob.26695.CHAT39>
- #9 <http://vegbank.org/cite/VB.ob.26468.CHAT24>
- #10 <http://vegbank.org/cite/VB.ob.26905.APAL74>

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- #11 <http://vegbank.org/cite/VB.Ob.4570.FIIS1>
- #12 <http://vegbank.org/cite/VB.Ob.4608.FIIS39>
- #13 <http://vegbank.org/cite/VB.ob.26793.APAL25>

APPENDIX THREE

Modal positions of dioecious trees, shrubs and vines along hydrological gradients

The following figures compare typical habitats of dioecious woody species with all woody species for Kentucky or other eastern states, following the format of Figure 2.

Among trees, shrubs and vines, there appear to be concentrations in the broad zone between:

(a) deeper woods on better drained terraces and hills; versus (b) wetlands, thin woods, shrubland and grassland on drier ground. It is also notable that among dominant herbaceous families typical of open lands, relatively few are dioecious or even dioecious-tending.

Following is an incomplete initial listing for Kentucky; * most/all alien. In deeper woods, especially mesic woods of various types, there are virtually no typical dioecious herbaceous species, with the notable exception of those marked @.

Ranunculaceae: *Thalictrum*@ (all?). Saxifragaceae: *Astilbe*@.

Rosaceae: *Fragaria*. Cannabaceae: *Cannabis*. Cucurbitaceae: *C. foetidissima* (check others).

Urticaceae: *Urtica* (esp. *dioica**); see also flexible monoecy in *Laportea*@.

Caryophyllaceae: some *Silene**. Polygonaceae: *Rumex acetosella*+*, *P. amphibium* (?).

Amaranthaceae: *Amaranthus tuberculatus* group (partly adventive).

Solanaceae: *S. carolinianum* (andromonoecious). Ericaceae: *Epigaea* (Conn et al. 1980).

Araliaceae: *Aralia nudicaulis*@; note also sex-changing *Panax trifolium*.

Asteraceae: *Antennaria*, *Anaphalis*.

Naijadaceae: *Naijas marina**; see also sex-changing *Arisaema*.

Liliales: *Asparagus**; *Chamaelirium*@, *Dioscorea*@

Poales—Cyperaceae: only *Carex picta*; ?no Juncaceae (but see *Distichia*, *Oxychloe*);

?no Poaceae (but to west see *Buchloe* and allies, *Distichlis*, *Gynerium*).

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(a) Trees of east-central states (same as in Figure 2a).

		CLIFFS	CLIFFS	CLIFFS	PINVIR JUNVIR ULMSER	PINRIG QUEPRI ULMALA	QUELAE QUEILI CELTEN	XERIC EXTREME
MESIC SLOPES (below)	CLIFFS	QUECOC QUEVEL QUESCH	QUEMON CARGLA FRAQUA	VACARB QUEFAL (ULMTHO)	PINECH (POPGRA) FRABIL	QUEMAR MALCOR GLETRI	QUEMRG MALANG (RHUTYP)	GRASS- LAND
TSUCAR HALTET CLAKEN	MAGMAC QUERUB TILAME	CASDEN CAROVA ULMRUB	OXYARB QUEALB QUEMUE	CARPAL CARTOM CARCAR	PINTAE SASALB PRUSER	PINPAL QUESTE GYMDIO	QUEINC (QUEIMB) (QUEMAC)	GRASS- LAND
TSUCAN FAGGRA ACESAC	BETLEN LIRTUL AESFLA	MAGACU ACERUB CARCOR	ILEOPA NYSSYL FRAAME	(PINSTR) (JUGCIN) JUGNIG	QUENIG ROBPSE MORRUB	QUEHEM DIOVIR MACPOM	PINSER QUEPHE QUESIN	GRASS- LAND
PLAOCC	MAGTRI BETNIG ACENEG	QUEMIC ULMAME	QUESHU CARLAC	QUEPAG CELSPP	QUELAU ILEDEC FRAPEN	(QUEPAL) CHATHY (QUEBIC)	PINELL TAXASC QUELYR	BOG or MARSH or FEN
POPDEL	ACESNM CATSPE	ACETRI CARILL	LIQSTY FRASUB	NYSBIF PLAAQU	MAGVIR FRACAR FORACU	PERPAL TAXDIS POPHET	CYRRAC NYSQU GLEAQU	STAGNANT WATER
RHEIC EXTREME	SALINT	SALNIG				STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(b) Dioecious or polygamo-dioecious trees of east-central states.

		CLIFFS	CLIFFS	CLIFFS	JUNVIR			XERIC EXTREME
MESIC SLOPES (below)	CLIFFS				POPGRA FRABIL	GLETRI	(RHUTYP)	GRASS-LAND
					SASALB	GYMDIO		GRASS-LAND
		ACERUB	ILEOPA NYSSYL FRAAME		DIOVIR MORRUB	MACPOM		GRASS-LAND
	ACENEG				ILEDEC			BOG or MARSH or FEN
POPDEL	ACESNM	ACETRI	FRAPEN	NYSBIF PLAAQU	FRACAR FORACU	POPHET	NYSAQU GLEAQU	STAGNANT WATER
RHEIC EXTREME	SALINT	SALNIG				STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(c) Small trees and shrubs of Kentucky (rare/local species in parentheses).

		CLIFFS (PHIPUB)	CLIFFS PHIHIR	CLIFFS VIBRAF	(RHOMIN) (JUNCOM) HYPFRO	(GAYBRA) CORRAC RHALAN	(AMESAN) ROBHIS RHUARO	(AMESPI) (SYMALB) PHYOPU
MESIC SLOPES (below)	(RHOCAT) PHIINT	STEOVA VIBACE DIRPAL	VACCSTA CORFLO OSTVIR	VACCOR AMEARB VIBRUF	KALLAT ROSCAR (RIBMIS)	GAYBAC CHIVIR PTETRI	(AMELAE) CRLAME PRUANG	(EUBREC) CRASPP CRACRU+
	(LEUFON) (TAXCAN) (VIBMOL)	RHOMAX CORALT RIBCYN	PYRPUB (NESUMB) STATRI	RHOPRI HAMVIR EUOATR	RHOCAL (STYGRA) SIDLYC	AROMEL PRUAME CORDRU	RHUCOP RHUGLA ZANAME	(VACERY) HYPPRO (BRBCAN)
	(VIBLAN)	CLEACU (ACEPEN) (ACESPI)	ILEMON LINBEN CRPCAR	RHOPER EUOAME VIBPRU	RHOCUM ASITRI FRACAR	ARUAPP ARASPI SYMOCC	AROPRU SALHUM ROSSET	(AMECAN) HYPLOB PRUMUN
	(SAMPUB)	CALFLO VIBDEA HYDARB	XANSIM VIBCAS AESPVA	RHOCAN ILEDEC SAMCAN	VACFUS VIBNUD ARUGIG	ARUTEC ROSPAL (RIBEAME)	SPITOM SALOCC (SPIALB)	(CLEALN) (TOXVER) (COROBL)
	(COMPER) (SPIVIR) (SALERI)	RHOARB VIBREC COROBL	(SALDIS) CORAMM AMOFRU	ILEVER ITEVIR AMONIT	LYOLIG STYAME AMOCRO	CRAVIR+		STAGNANT WATER
RHEIC EXTREME	SALSER SALCAR SALINT	ALNSER	CORSTR	CEPOCC	DECVER	STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(d) Dioecious or polygamo-dioecious small trees and shrubs of Kentucky.

		CLIFFS	CLIFFS	CLIFFS		RHALAN	RHUARO	XERIC EXTREME
MESIC SLOPES (below)						CHIVIR PTETRI		GRASSLAND
(TAXCAN)		PYRPUB (NESUMB)				RHUCOP RHUGLA ZANAME		GRASSLAND
		ILEMON LINBEN				SALHUM ROSSET		GRASSLAND
						SALOC		BOG or MARSH or FEN
(COMPER) (SALERI)		(SALDIS)	ILEVER					STAGNANT WATER
RHEIC EXTREME	SALSER SALCAR SALINT					STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(e) Woody vines of Kentucky (same as in Figure 7).

		CLIFFS	CLIFFS	CLIFFS				XERIC EXTREME
					(LONRET)			
MESIC SLOPES (below)	LONDIO					VITAES SMIBON		GRASSLAND
(SCHGLA)				SMIROT	SMIGLA COCCAR	LONSEM CELSCA		GRASSLAND
ISOMAC		BIGCAP	PARQUI	VITROT VITBAIL VITVUL	CLEVIR MENCAN	(LACMUL) ROSSET		GRASSLAND
ISOTOM		(HUMLUP)	[DECBAR] TOXRAD SMIHIS	VITLAB VITCIN CAMRAD	AMPARB (BERSCA)			BOG or MARSH or FEN
SCoured SHORES	VITRIP	AMPCOR CALLYO	WISFRU VITPAL	THYDIF WISMAC				STAGNANT WATER
RHEIC EXTREME	(VITRUP)					STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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(f) Dioecious or polygamo-dioecious woody vines of Kentucky.

		CLIFFS	CLIFFS	CLIFFS				XERIC EXTREME
MESIC SLOPES (below)	CLIFFS				VITAES SMIBON			GRASSLAND
				SMIROT	SMIGLA COCCAR	CELSCA		GRASSLAND
				VITROT VITBAIL VITVUL	CLEVIR MENCAN	ROSSET		GRASSLAND
		HUMLUP	TOXRAD SMIHIS	VITLAB VITCIN				BOG or MARSH or FEN
SCoured SHORES	VITRIP	CALLYO	VITPAL					STAGNANT WATER
RHEIC EXTREME	VITRUP					STAGNANT WATER	STAGNANT WATER	HYDRIC EXTREME

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APPENDIX FOUR

Table of *Lonicera* spp., with notes on range, habit, browsing by ruminants and chemistry.

Species	Range; approx. hardness zone	Habit	Ruminant Use; with references	Chemistry; with references
CAPRIFOLIUM GROUP				
<i>ciliata</i>	W North America HZ 4-6?	vine; DEC		+ apigenin + luteolin (Glennie 1969)
<i>hirsuta</i>	E North America HZ 3-4	vine; DEC	?medium-low use (Beals+ 1960, Weatherill+ 1969, Wright+ 2002)	low phenolics (Lieurance 2012)
<i>dioica</i>	E North America HZ 4-6	vine; DEC	??high use (JC)	
<i>reticulata</i> (= <i>prolifera</i>)	E North America HZ 5-6	vine; DEC	??high use (JC)	low phenolics (Lieurance 2012)
<i>flava</i>	E North America HZ 7	vine; DEC	?high use (Turner+ 2009)	low phenolics (Lieurance 2012)
<i>sempervirens</i>	E North America HZ 6-9	vine; SEG or delayed deciduous	high-medium on average but variable (Schierenbeck+ 1994, Ashton+ 2008, Jull 2001, Wade+ 2010)	low phenolics (Lieurance 2012) - apigenin + luteolin (Glennie 1969)

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<i>albifolia</i>	S North America HZ 8–9	vine; SEG- DEC?	?high use (Nelle 1996)	
<i>periclymenum</i>	N Europe HZ 4–7	vine; DEC- SEG	high use (Gonzalez-H+ 1996*, 1999*, Gill+ 2001, McEvoy+ 2006)	?low phenolics; (Gonzalez-H.+ 2003) pr = 10-11%*
<i>caprifolium</i>	S Europe HZ 7–8	vine; SEG- DEC?	high use (Mitchell+ 1990, Kirby 2001)	
× <i>heckrottii</i> , etc.	(horticultural) HZ 6–8?	vine; DEC- SEG	?medium-high use (Fargione+ 2001)	
<i>implexa</i> (<i>capri- folium</i> ssp. <i>implexa</i>)	SW Europe HZ 8–9?	vine; SEG- DEC?	high use (Cabbidu+ 2000, Decandia+ 2000*)	?low phenolics (Cabbidu+ 2000) + apigenin + luteolin (Flamini+ 1997) ?pr = 7%*
<i>etrusca</i>	S Europe HZ 8–9?	vine; DEC- SEG	high use (LeHouérou 1980, Kidjo+ 2007; NOT Papageorgiou+ 1981* in summer)	+ apigenin + luteolin (Guyen.+ 2012) pr = 8%*
<i>tragophylla</i> (<i>subaequalis</i> , <i>yunnanensis</i>)	S China HZ 6–9?	vines; DEC- SEG?		

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JAPONICA GROUP				
<i>biflora</i>	Mediterranean HZ 6–9?	shrub <2m? DEC-SEG?	(grows with Nerium oleander, L. <i>implexa</i>)	
<i>acuminata</i> or allies (<i>henryi</i>)	Sino-Himalaya HZ 6–9?	vines; SEG	??medium-high use (see refs in Table 5)	
<i>japonica</i> or allies (<i>affinis</i> , <i>hypoglauca</i> , etc.)	East Asia HZ 6–9	vines; SEG	medium-high use esp. in winter (see refs. in Table 5)	high phenolics (Lieurance 2012) + apigenin (Son+ 1992) + luteolin (Zhang+ 2009) TAS: mealy- bitter
<i>ferruginea</i> or allies	East Asia HZ 6–9?	vines; SEG?		
<i>macrantha</i> or allies (<i>confusa</i> , <i>dasystylis</i> , <i>similis</i> etc.)	East Asia HZ 6–9?	vine; SEG- DEC		+ luteolin (Li+ 2001, Yao+ 2006, Qin+ 2008)

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XYLOSTEUM GROUP				
<i>xylosteum</i>	Europe HZ 4–6	shrub; DEC	medium use (Mysterud+ 1999, Boulanger+ 2009)	high phenolics (Lieurance 2012) + apigenin + luteolin (Glennie 1969)
<i>tatarica</i> (also <i>ruprechtiana</i> , <i>chrysantha</i>)	East-Central Asia HZ 4–6	shrub; DEC	?low use (Heinrich+ 1995; see also Swihart+ 1983)	high phenolics (Lieurance 2012) + apigenin (Jiang+ 2008) - apigenin (Glennie 1969)
× <i>bella</i> , etc.	(horticultural) HZ 4–6	shrub; DEC	?low use (Mudrak+ 2009)	
<i>morrowii</i>	N East Asia HZ 4–6	shrub; DEC	?medium use or variable (Turner+ 2009, Averill 2012)	
<i>maackii</i>	East Asia HZ 5–7	shrub; DEC but delayed	low-medium use (Heinrich+ 1995, Trisel 1997; JC pers. obs.)	high phenolics; + apigenin + luteolin (Lieurance 2012) TAS: mealy- bitter(-acid)

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CAERULEA GROUP				
<i>hypoleuca</i> ? (position unclear)	Himalayas HZ 3–5?	shrub; DEC		
<i>ligustrina</i> or allies (<i>nitida</i> , <i>pileata</i>)	Sino-Himalaya HZ 5–7?	shrub; SEG- DEC		+/- apigenin + luteolin (Glennie 1969)
<i>caerulea</i> or allies	Eur Asi NAME HZ 3–5?	shrub; DEC		?high phenolics + luteolin (Chaovanalikit+ 2004, Borchard+ 2011)
<i>villosa</i> (<i>caerulea</i> var. <i>villosa</i>)	E North America HZ 3–4?	shrub; DEC		
REMAINING (regular flws)				
<i>involutrata</i> (or <i>ledbourii</i>)	W N. America HZ 3–4?	shrub; DEC	?low to medium use, esp. moose (Harry 1957, Weatherill+ 1969)	+ apigenin + luteolin (Glennie 1969)

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<i>gracilipes</i>	Japan	shrub; DEC		+ apigenin + luteolin (Kikuchi+ 1996)
<i>angustifolia</i> group (<i>myrtilus</i> <i>rupicola</i> , <i>syrinantha</i>)	Sino-Himalaya	shrub; DEC		+ luteolin (Qian+ 2006) +/- apigenin (Glennie 1969)
REMAINING (bilabiate flws)				
<i>canadensis</i>	E North America HZ 4-5	shrub; DEC	medium-high on average but very variable (Howard 1937, Atwood 1941, Krefting 1941, Beals+ 1960, Christensen 1963, Telfer 1967, Petrides 1975, Belovski 1981 (?), Cornett+ 2000, Wright+ 2002)	
<i>oblongifolia</i>	E North America HZ 3-4?	shrub; DEC	?low use (Renton 2010; increases with cattle)	+ apigenin + luteolin (Glennie 1969)

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<i>utahensis</i>	W N, America HZ 3-4?	shrub; DEC	?medium use, esp. moose (Harry 1957)	+ apigenin + luteolin (Glennie 1969)
<i>maximowiczii</i> and <i>nigra</i> groups	Eurasia HZ 3-8??	shrub; DEC		?high phenolics (Borchard+ 2011)
<i>tangutica</i> (<i>saccata</i>) and <i>albigena</i> groups	Eurasia HZ 3-8??	shrub; DEC	?low use (Winkler 1998, JC pers. obs.; increases in rough pasture)	+ apigenin + luteolin (Li+ 2001) (but Glennie 1969 did not)
<i>hispida</i> group	Eurasia HZ 3-5??	shrub; DEC	?low use (Winkler 1998)	
<i>fragrantissima</i> group (<i>microphylla</i>)	East Asia HZ 7-8	shrub; DEC- SEG	low use (Heinrich+ 1993)	high phenolics (Lieurance 2012) + apigenin + luteolin (Chumbalov+ 1978) TAS: mealy- slightly bitter

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DIERVILLA				
<i>lonicera</i>	E North America HZ 3-5	shrub; DEC	medium use but often persists/regrows (see refs. under <i>L. canadensis</i> ; also Murie 1934, Christensen 1963, Belovsky 1981, Cornett+ 2000, Crawford+ 1993)	?high phenolics (Lieurance 2012 etc.)
<i>sessilifolia</i>	E North America HZ 6?	shrub; DEC	?medium use but persists (Atwood 1941, Barden 1978; see also Todd 1927)	
<i>rivularis</i>	E North America HZ 7?	shrub; DEC		



