

Photosynthetic and morphological functional types from different steppe communities in Inner Mongolia, North China

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Abstract

Morphological functional types and photosynthetic pathway types were identified for the forage species from steppe communities in Inner Mongolia, China, using the data of both field survey and published papers. Seven typical steppe communities were selected to investigate the morphological functional type and photosynthetic pathway type compositions and plant functional type (PFT) diversity in steppe communities at regional scale. Morphological functional types, based on plant height and leaf type combined with life span, were optimal for comparing the community differences in the region, while photosynthetic pathway types were fairly coarse for such studies. Of the seven morphological functional types in the steppe communities, perennial forbs (PEF) were the dominant type, and 60 % of species belonged to this type. Each of the high perennial grass (HPG), short perennial grass (SPG), and annual grass (ANG) types represented less than 10 % of the total, even though the grass species were dominant in the seven steppe communities. The differences of PFTs between the steppe communities were remarkable, and the PFT richness and diversity increased from the communities with moist conditions to the ones with dry environments.

Additional key words: C₃, C₄, and CAM plants; morphology; photosynthetic pathway; plant diversity; plant functional types.

Introduction

Plant functional types (hereafter PFTs) refer to groups or types of plant species that share morphological and physiological attributes and play a similar role in ecosystems (Chapin 1993, Paruelo and Lauenroth 1996, Wang 2003). They are derived from plant attributes based on morphology, physiology, regenerative growth, life form, and bioclimatic tolerance according to the aims and scales of the study (Smith *et al.* 1993, Duckworth *et al.* 2000, Wang 2003). PFTs are popular means for studying the logical links between physiological and life-history strategies at plant level and ecological processes at ecosystem and global levels (Chapin 1993, Paruelo and Lauenroth 1996). Most of studies were focused on PFT classification (Waller and Lewis 1979, Smith *et al.* 1993, Paruelo and Lauenroth 1996, Scholes *et al.* 1997, Pillar 1999, Duckworth *et al.* 2000, Wang 2003) and their relations with local and global change (Teeri and Stowe 1976, Collins and Jones 1985, Chapin 1993, Cramer 1997, Collatz *et al.* 1998). Some researchers tested the PFT diversity and abundance in grasslands, shrublands, and woodlands (Paruelo and Lauenroth 1996). These works provide

strong evidence that the abundance of PFTs correlates with vegetation differences, land use, and climatic change at local, regional, and global scales. Nevertheless, a comparative study on the differences in PFT composition and diversity at community level in steppe vegetation in China was lacking.

Steppe, distributed at the eastern end of the Eurasian steppe zone, covers large continuous areas in Mongolia plateau, and the main location in China is Inner Mongolia and the western Manchuria plain. The usual growing conditions on the steppe produce herbage superior both in quality and in quantity, so that this type of grassland is one of the best suited in northern China for the rangeland industry (Jiang *et al.* 1985, Li *et al.* 1988). Studies on local flora, grassland ecosystems, plant biomass, and their responses to climatic variables have been conducted (Jiang *et al.* 1985, Li *et al.* 1988, Zhao *et al.* 1988, Liu 1993), but differences in PFT composition and diversity between steppe communities remain unclear. The objective of the present study was to investigate the compositions of morphological functional type and photosynthetic

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pathway type and PFT diversity in steppe communities at regional scale. This study brings new information for better understanding the plant functional types in Inner

Mongolian steppe and their relations with ecosystems, climate changes, and land use.

Materials and methods

Study site: The study was conducted in the Xilin river basin (41°35'–46°46' N, 111°09'–119°58' E) in the Mid-Inner Mongolia. The typical steppe in the region, which covers about 40 000 km² area, is part of the Mongolian Plateau of average 1 200 m a.s.l., varying from 900 m in the northeast to 1 500 m in the southwest. Most of the region has dark chernozem and chestnut soils. Because of the variability in topography (elevation and landscape), annual precipitation, and annual temperature, the community types and soils vary significantly from upland to lowland or from high southwest to low northeast. There are seven main communities in the region, e.g. *Filifolium sibiricum* (FS), *Stipa baicalensis* (SB), *Festuca ovina* (FO), *Leymus chinensis* (LC), *Stipa grandis* (SG), *Stipa krylovii* (SK), and *Artemisia frigida* (AF). Floristic composition varied among the seven community types, 184 species were identified in LC, 121 in FS, 104 in SG, 99 in FO, 96 in SB, 62 in AF, and 40 in SK. These variations resulted in the differences in PFT composition and diversity.

Climate: The main determinants of the climate in the region are the Mongolian anticyclone and moist Pacific air mass. In winter, northeastern China is dominated by an intense Mongolian anticyclone. The steep pressure gradient between this high and the Aleutian low-pressure system produces a strong westerly flow of cold, dry continental air at the surface over north China. As the anticyclone breaks down in spring, the region comes increasingly under the influence of moist Pacific air mass, reaching a climax in the summer monsoon, which lasts two months. The mean annual air temperature ranges from –1.4 °C to +2 °C, varying from –21 °C in January to +21 °C in July. Moisture gradient is very steep, with annual precipitation varying from 250 mm in the northwest to 450 mm in the southeast. Precipitation is not distributed evenly over growing season, of which 70 % falls between June and August in this region.

Methods: Community floristic composition was obtained from the field survey in the growing seasons of 2000 and

2001, as well as from literature published from 1988 to 2003 (Li *et al.* 1988, Zhao *et al.* 1988, Liu 1993, Wang 2003). In the field survey, plants were sampled from 60–80 randomly located 1×1 m quadrats in each community type. Considering the species-area relationship, 8–10 communities with different area were used as community replicates in each community type. Plant species, morphology (e.g. plant height, leaf type, life span), landscape, elevation, and soil type were recorded. The data on photosynthetic pathway types were compiled from references published between 1985 and 2002 (e.g. Takeda and Hakoyama 1985, Li *et al.* 1988, Li 1993, Redmann *et al.* 1995, Pyankov *et al.* 2000, Wang 2002a,b, 2003). Plants identified in the region were classified into photosynthetic pathway types (C₄, C₃, and CAM) by physiological attributes and into morphological functional types: shrubs (SHR), high perennial grasses (HPG), short perennial grasses (SPG), annual grasses (ANG), annual forbs (ANF), perennial forbs (PEF), and succulents (SUC) by morphological attributes (Zhao *et al.* 1988, Li *et al.* 1988, Wang 2003) (Table 2).

Mean species number and proportion of PFTs in each community type were statistically analyzed with the *MINITAB 13.1*, and regressions of PFT richness and diversity from community types were performed using *EXCEL for Windows 98*. PFT richness (R) was calculated using Margalef's (1959) index:

$$R = \frac{S - 1}{\ln N} \quad (1)$$

where *N* is the number of species in the community and *S* is the number of PFTs.

PFT diversity (H) was calculated using the Shannon-Wiener index (Barbour *et al.* 1987):

$$H = - \sum_{i=1}^S P_i \ln P_i \quad (2)$$

where *P_i* is the PFT proportion of the total PFT number in each community, and *S* is the number of PFTs.

Results

PFT: Of the total 257 species, 126 species were identified with C₃ (97 species), C₄ (26 species), and CAM (3 species) photosynthetic pathways (Table 2) which is 77, 21, and 2 %, respectively. The occurrence of C₄ species was common in *Gramineae* (12 species) and *Chenopodiaceae* (6 species). *Gramineae* was the leading C₄ family, followed by *Chenopodiaceae*. The differences in

the occurrence and percentage of photosynthetic pathway types among the seven communities were significant in the steppe region (Fig. 1A,B). LC and SG communities had the greatest number of species in both C₃ and C₄, followed by AF, FS, and SK. In general, the numbers for both C₃ and C₄ species were reduced significantly from LC to SK communities, the species numbers of C₃ and C₄

for SK community being only about 37 and 29 % of that for LC community, respectively. There were more CAM species among the relatively dry vegetation types (e.g. AF, FO, and SG). The percentages of C₃ and C₄ species in LC, SG, AF, and SK were fairly high (Fig. 1B).

Relatively more plants of the C₄ type in the drier communities (e.g. AF and SK) indicated that C₄ plants are generally more tolerant to drought and have greater ability to maintain intense photosynthesis.

Table 1. The community types, topography, soil types, annual precipitation, and annual temperature of the Xilingol steppe in Inner Mongolia, North China.

Community type	Landscape	Elevation [m]	Soil type	Annual precipitation [mm]	temperature [°C]
<i>Filifolium sibiricum</i>	Upland	1300~1500	Dark chernozem	>450	-1.4
<i>Stipa baicalensis</i>	Upland	1270~1500	Dark chernozem	<450	-1.4
<i>Festuca ovina</i>	Upland	1300~1500	Dark chernozem	<450	-1.4
<i>Leymus chinensis</i>	Lowland	900~1300	Chernozem	350~450	-0.3~1
<i>Stipa grandis</i>	Lowland	1100~1300	Chernozem	350~450	-0.3~1
<i>Stipa krylovii</i>	Lowland	900~1100	Light chernozem	250~350	1~2
<i>Artemisia frigida</i>	Lowland	900~1100	Light chernozem	250~350	1~2

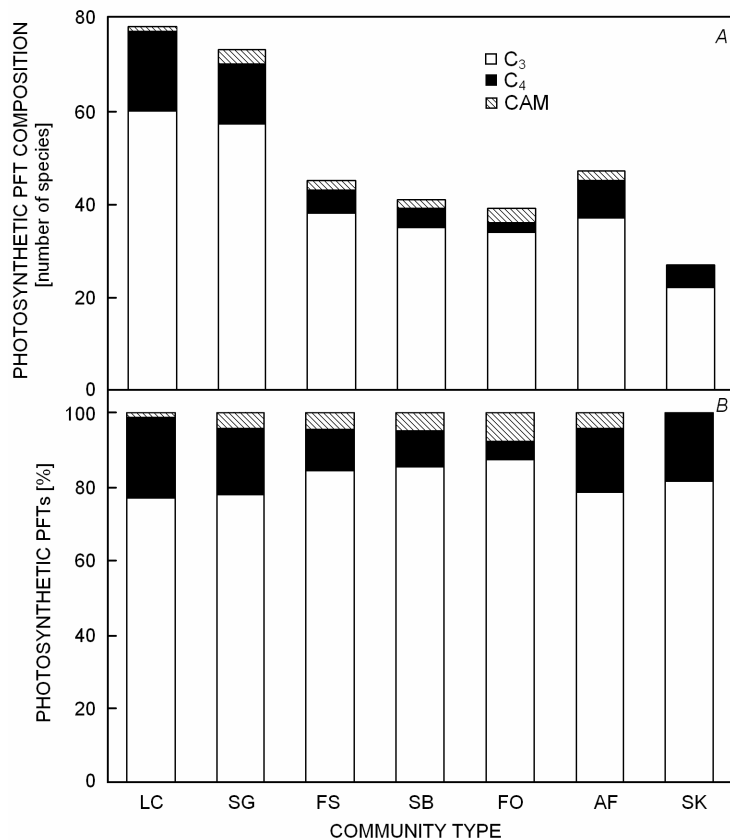


Fig. 1. The photosynthetic pathway types of the present plant species (A) and their composition (B) in different community types (PFTs) in Inner Mongolia steppe, north China.

Morphological functional types: A detailed description of the floristic composition in each community is presented in Table 2. Of the 257 species identified in the seven communities, 7 functional types, e.g. shrubs (SHR), high perennial grasses (HPG), short perennial grasses (SPG), annual grasses (ANG), annual forbs (ANF), perennial

forbs (PEF), and succulents (SUC) were grouped by morphological traits, e.g. height, leaf morphology, and life span. There were six PFTs in the SB and FO communities, but seven PFTs occurred in the other five communities. Of the total identified species in the region, 60 % was of the PEF type, followed by ANF (15 %),

HPG (10%), SHR (6%), SPG (4%), ANG (4%), and SUC (2%). The grass PFTs represented about 18% of all, even though the species were dominant (*e.g.* *Stipa grandis* P. Smirn., *S. baicalensis* Roshev., *Leymus chinensis* Trin. Tzvel.), and produced 90% of herbage in the steppe vegetations. This indicated that the abundance of PFTs was not consistent with the community composition.

The morphological PFT compositions of the communities varied significantly due to the differences in precipitation, soil, mean annual temperature, and elevation (Table 1). Species abundance of PEF, ANF, and HPG dropped significantly from the LC to SK community (Fig. 2A). Number of PEF species in the SK community was only 25% of that in LC community, and the numbers of ANG and HPG species in SK were about 20 and 35%

of those in LC, respectively. Species numbers in the other two grass PFTs also declined from LC to SK. The numbers of SHR and SUC species did not vary considerably, except that they were a little lower in the LC and SK communities. PFT percentages varied remarkably in the seven communities (Fig. 2B): PEF percent exhibited relatively little variation among the seven communities, being significantly low in the AF community, while the ANF percent was twice the average value. The SHR percent increased from LC to SK, *i.e.* from moist to drier. Annual grass percentages were rather low in the seven communities, and no annual grasses were identified in both SB and FO communities, indicating that human disturbances of the communities were not common in the region.

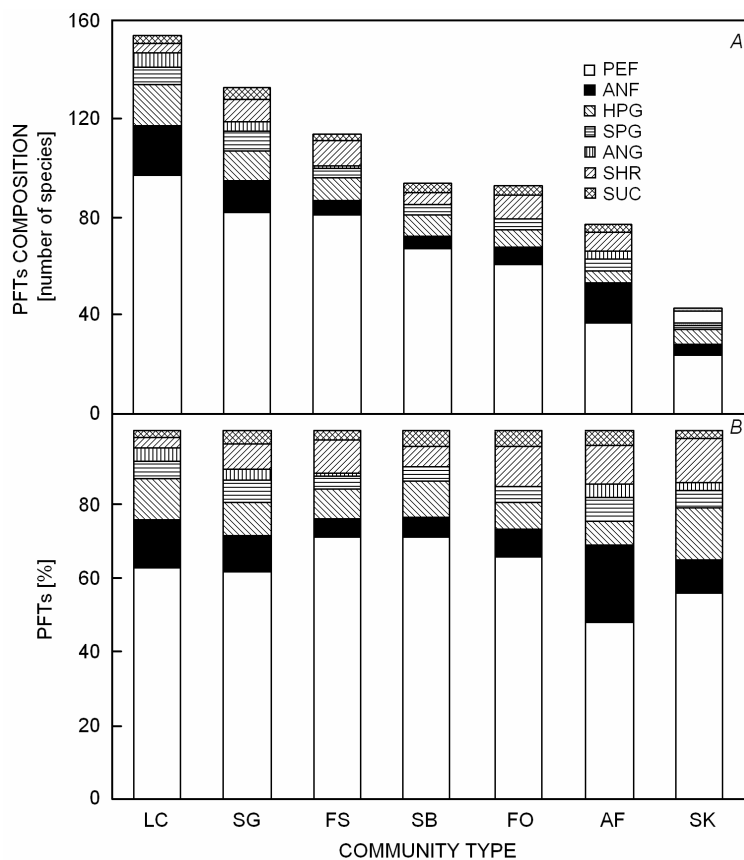


Fig. 2. Species numbers in each plant functional type, PFT (A) and their percentages (B) in steppe communities in Inner Mongolia, north China.

PFT richness and diversity: Variations in numbers of species, the simplest measure of PFT diversity in the seven communities, were described above. But the richness index of Margalef (Eq. 1) and diversity index of Shannon-Wiener (Eq. 2) can comprehensively reflect the PFT differences among the communities. PFT richness index (R) incorporating both the number of PFTs and the number of species in PFT was the highest in SK, even if the total number of species in this community was consi-

derable lower. In general, the R index increased from the moist community to the drought one (Fig. 3). This suggested that the PFT richness was not consistent with the community species abundance in the steppe vegetation.

The Shannon-Wiener diversity index H (Eq. 2) includes not only richness, but also evenness of the PFT distribution. The latter is the ratio of diversity to maximum possible diversity for the given numbers of species and PFT. This index also increased significantly

Table 2. Photosynthetic pathways (C₃, C₄, and CAM) and plant functional types from different communities in Inner Mongolian steppe, North China. Nomenclature follows Kitagawa (1979) and Yin and Wang (1997). Communities: FS = *Filifolium sibiricum* community, SB = *Stipa baicalensis* community, FO = *Festuca ovina* community, LC = *Leymus chinensis* community, SG = *Stipa grandis* community, SK = *Stipa krylovii* community, AF = *Artemisia frigida* community. Plant functional types (PFTs): SHR = shrubs, HPG = high perennial grasses, SPG = short perennial graminaceous plants (grasses and sedge), ANG = annual grasses, ANF = annual forbs, PEF = perennial forbs, SUC = succulents.

	Species	C ₃ /C ₄	Community	PFTs
<i>Gymnospermae</i>				
<i>Ephedraceae</i>	<i>Ephedra sinica</i> Stapf.		FS SG SK AF	SHR
	<i>E. monosperma</i> Gmel. ex Mey.		FO	SHR
<i>Angiospermae</i>				
<i>Dicotyledoneae</i>				
<i>Polygonaceae</i>	<i>Polygonum angustifolium</i> Pall.		FS FO LC	PEF
	<i>P. aviculare</i> L.	C ₃	LC AF	ANF
	<i>P. divaricatum</i> L.		FS SB FO LC SG	PEF
	<i>P. sibiricum</i> Laxm.	C ₃	FS LC	PEF
<i>Chenopodiaceae</i>	<i>Agriophyllum pungens</i> (Vahl) Link A. Dietr.	C ₄	SG	ANF
	<i>Atriplex sibirica</i> L.	C ₄	LC	ANF
	<i>Axyris amaranthoides</i> L.	C ₃	AF	ANF
	<i>Bassia dasyphylla</i> O. Kuntze	C ₄	SG	ANF
	<i>Chenopodium acuminatum</i> Willd.	C ₃	LC	ANF
	<i>Ch. glaucum</i> L.	C ₃	LC	ANF
	<i>Kochia prostrata</i> Schrad.	C ₄	FS FO LC SG SK AF	SHR
	<i>K. sieversiana</i> (Pall.) C.A. Mey.	C ₄	LC	SUC
	<i>Salsola collina</i> Pall.	C ₄	SB SG SK AF	SUC
<i>Amaranthaceae</i>	<i>Amaranthus retroflexus</i> L.	C ₄	AF	ANF
<i>Portulacaceae</i>	<i>Portulaca oleracea</i> L.	C ₄	AF	ANF
<i>Caryophyllaceae</i>	<i>Arenaria capillaris</i> Poir.		FS FO	PEF
	<i>Dianthus superbus</i> L.		FO	PEF
	<i>D. versicolor</i> Fisch. ex Link		FS SB FO SG AF	PEF
	<i>Gypsophila davurica</i> Turcz. ex Fenzl		FS LC SG	PEF
	<i>Melandrium apricum</i> (Turcz.) Rhrb.		FS SB FO LC SG	ANF
	<i>Silene jenissensis</i> Willd.		FS SB FO LC SG SK	PEF
	<i>S. repens</i> Patr.		LC	PEF
	<i>Stellaria cherleriae</i> Willd.		FS	PEF
<i>Ranunculaceae</i>	<i>Clematis hexapetala</i> Pall.	C ₃	FS FO LC	PEF
	<i>Delphinium grandiflorum</i> L.		SB LC	PEF
	<i>Pulsatilla ambigua</i> Turcz. ex Pritz.		FS SB FO LC SG	PEF
	<i>P. turczaniovii</i> Kryl. ex Sery.		LC	PEF
	<i>Ranunculus japonicus</i> Thumb.	C ₃	SG	PEF
	<i>Thalictrum petaloedum</i> L. var. <i>supr.</i> (Nakai) Kitag.	C ₃	SG SB	PEF
	<i>Th. simplex</i> L.	C ₃	SB FO SG SK	PEF
	<i>Th. squarrosum</i> Steph.	C ₃	FS SB FO LC SG AF	PEF
<i>Papaveraceae</i>	<i>Papaver nudicaule</i> L.		LC	PEF
<i>Cruciferae</i>	<i>Dontostemon integrifolium</i> (L.) Ledeb.		FO LC SG SK AF	ANF
	<i>D. micranthus</i> C.A. Mey.		FS	ANF
	<i>Draba nemoroda</i> var. <i>leiocarpa</i> Linded.		FS SB FO LC SG	ANF
	<i>Lepidium apetalum</i> Willd.	C ₃	AF	ANF
	<i>Ptilotrichum elongatum</i> C.A. Mey.		FS SB FO SG AF	SHR
	<i>Thlaspi cochleariforme</i> D.C.		SB FO LC	PEF
<i>Crassulaceae</i>	<i>Orostachys fimbriatus</i> (Turcz.) Berger	CAM	FS SB FO SG AF	SUC
	<i>O. malacophyllus</i> (Pall.) Fisch.	CAM	SB FO SG AF	SUC
	<i>Sedum aizoon</i> L.	CAM	FS FO LC SG	SUC
<i>Rosaceae</i>	<i>Chamaerhodos canescens</i> J. Krause		FS	PEF
	<i>Ch. erecta</i> (L.) Bunge		FO SG SK AF	ANF
	<i>Geum aleppicum</i> Jacq.	C ₃	LC	PEF

Table 2 (continued)

	Species	C ₃ /C ₄	Community	PFTs
Rosaceae (cont.)	<i>Potentilla acaulis</i> L.	C ₃	SB FO LC SG SK AF	PEF
	<i>P. anserina</i> L.	C ₃	FS	PEF
	<i>P. bifurca</i> L.	C ₃	FS SB FO LC SG SK AF	PEF
	<i>P. flagellaris</i> Willd.	C ₃	LC	PEF
	<i>P. fragaricoides</i> L.	C ₃	SG SK	PEF
	<i>P. longifolia</i> Willd. ex Schlech.		SB	PEF
	<i>P. multifida</i> L. var. <i>nubigena</i> Wolf.		AF	PEF
	<i>P. sericea</i> L.		FS SG AF	PEF
	<i>P. tanacetifolia</i> Willd. ex Schlech.		FS SB FO LC SG SK AF	PEF
	<i>P. verticillaris</i> Steph. ex Willd.		FS SB FO LC SG SK AF	PEF
	<i>P. viscosa</i> Willd. ex Schlech.		SB FO LC SG	PEF
	<i>Sanguisorba officinalis</i> L.	C ₃	FS SB FO LC SG	PEF
	<i>Sibbardia adpressa</i> Bunge		FS	PEF
	<i>S. bijugs</i> Bunge		FS SG	PEF
<i>Spiraea aquilegifolia</i> Pall.		FS FO	SHR	
Fabaceae	<i>Astragalus adsurgens</i> Pall.		FS SB FO LC SG	PEF
	<i>A. dahuricus</i> (Pall.) D.C.		AF	ANF
	<i>A. galactites</i> Pall.		FS FO LC SG SK AF	PEF
	<i>A. melilotoides</i> var. <i>tenuis</i> Turcz.		FS SB FO LC SG	PEF
	<i>A. mongholicus</i> Bunge.		LC	PEF
	<i>A. scaberrimus</i> Bunge.		FO SG	PEF
	<i>Caragana microphylla</i> Lam.	C ₃	FS SB FO LC SG SK AF	SHR
	<i>C. stenophylla</i> Pojark.	C ₃	SG SK AF	SHR
	<i>Glycyrrhiza uralensis</i> Fisch.		LC SG	PEF
	<i>Gueldensfaedtia stenophylla</i> Bunge.	C ₃	SB FO SG SK AF	PEF
	<i>G. verna</i> (Georgi) A. Bor.		LC	PEF
	<i>Hedysarum gmelini</i> Ldb.		FS SB FO LC SG AF	PEF
	<i>H. lignosum</i> Ldb.		FO	SHR
	<i>Lathyrum palustris</i> L.		LC	PEF
	<i>L. quinquenervius</i> Miq.		SB	PEF
	<i>Lespedeza davurica</i> Schindler	C ₃	FS SG	SHR
	<i>L. hedysaroides</i> Kitag.		FS	SHR
	<i>Medicago falcata</i> L.		LC	PEF
	<i>M. lupulina</i> L.		LC	ANF
	<i>Melilotoides ruthenica</i> (L.) Sojak	C ₃	LC FS	PEF
	<i>Melissitus ruthenica</i> (L.) Peschkova		FS SB LC SG AF	PEF
	<i>Oxytropis hirta</i> Bunge	C ₃	LC SG	PEF
	<i>O. filiformis</i> D.C.	C ₃	FS SK AF	PEF
	<i>O. grandiflorus</i> Pall.		FS	PEF
	<i>O. leptophylla</i> (Pall.) D.C.		FS SB SG AF	PEF
	<i>O. myriophylla</i> D.C.	C ₃	FS SB FO LC SG AF	PEF
	<i>O. ochrantha</i> Turcz.		FS LC	PEF
	<i>Swainsona salsula</i> Pall.		LC	PEF
	<i>Thermopsis lanceolata</i> R.Br.	C ₄	FS SB LC SG	PEF
	<i>Trifolium lupinaster</i> L.		FS SB FO LC SK	PEF
	<i>Vicia ampena</i> Fisch. var. <i>oblongifolia</i> Reg.		FS SB LC	PEF
	<i>V. cracca</i> L.		SB LC	PEF
	<i>V. unijuga</i> R.Br.		FS LC	PEF
Geraniaceae	<i>Erodium stephanianum</i> Willd.	C ₃	AF	ANF
Linaceae	<i>Linum baicalense</i> Juz.		LC	ANF
	<i>L. perenne</i> L.		FS SB	PEF
Zygophyllaceae	<i>Tribulus terrestris</i> L.	C ₄	LC	ANF
Rutaceae	<i>Haplophyllum dauricum</i> Juss.	C ₃	FS SB FO LC SG AF	PEF
Polygalaceae	<i>Polygala tenuifolia</i> Willd.	C ₃	FS SB FO LC SG	PEF
Euphorbiaceae	<i>Euphorbia esula</i> L.	C ₃	FS SB FO LC SG	PEF
	<i>E. hirtella</i> Maxim.		LC	ANF

Table 2 (continued)

	Species	C ₃ /C ₄	Community	PFTs
<i>Euphorbiaceae</i> (cont.)	<i>Euphorbia humifusa</i> Willd.	C ₄	AF	ANF
<i>Violaceae</i>	<i>Viola dissecta</i> Ledeb.	C ₃	LC SG	PEF
<i>Thymelaeaceae</i>	<i>Diarthron linifolium</i> Turcz.	C ₃	SG	ANF
	<i>Stellera chamaejasme</i> L.	C ₃	FS SB FO LC SG AF	SUC
<i>Umbelliferae</i>	<i>Bupleurum scorzonerifolium</i> Willd.	C ₃	FS SB FO LC SG AF	PEF
	<i>Saposhnikovia divaricatum</i> (Turcz.) Schischk.	C ₃	FS SB FO LC SG SK AF	PEF
<i>Primulaceae</i>	<i>Androsace longifolia</i> Turcz.		FS SB FO LC SG SK AF	PEF
	<i>A. septentrionalis</i> L.		FO LC	ANF
	<i>Glaux maritima</i> L.	C ₃	LC	ANF
	<i>Primula sibirica</i> Jacq.		LC	PEF
<i>Plumbaginaceae</i>	<i>Limonium bicolor</i> O. Kuntze	C ₃	FS LC SG AF	PEF
<i>Gentianaceae</i>	<i>Gentiana dahurica</i> Fisch.		FS FO LC	PEF
	<i>G. squarrosa</i> Ldb.		FS SB LC SG AF	ANF
	<i>Gentianopsis barbata</i> (Froel.) Ma		FS SB	ANF
<i>Convolvulaceae</i>	<i>Convolvulus ammannii</i> Desr.		AF	ANF
<i>Asclepiadaceae</i>	<i>Cynanchum amplexicaule</i> Hemsl.	C ₃	SG	PEF
<i>Boraginaceae</i>	<i>Cynoglossum divaricatum</i> Steph.	C ₃	AF SG	ANF
	<i>Eritrichium mandshuricum</i> M. Pop.		FS	PEF
	<i>E. rupestre</i> (Pall.) Bunge		SB FO	PEF
	<i>Lappula myosotis</i> Moench.		FO LC SG SK	ANF
<i>Labiatae</i>	<i>Phlomis mongolica</i> Turcz.		LC	PEF
	<i>Ph. tuberosa</i> L.		FS SB SG AF	PEF
	<i>Schizonepeta multifida</i> (L.) Briq.		FS SB FO LC AF	PEF
	<i>Scutellaria baicalensis</i> Georgi	C ₃	FS SB FO LC SG	PEF
	<i>S. ikonnikovii</i> Juz.	C ₃	LC	PEF
	<i>S. scordiifolia</i> Fisch. ex Schrank		SB FO LC SG	PEF
	<i>S. viscidula</i> Bunge		FS	PEF
	<i>Thymus serpyllum</i> L.	C ₃	FS FO	SHR
	<i>Th. mongolicum</i> Ronn.		SG AF	SHR
	<i>Plantaginaceae</i>	<i>Plantago asiatica</i> L.	C ₃	LC AF
<i>P. depressa</i> Willd.		C ₃	LC AF	PEF
<i>Scrophulariaceae</i>	<i>Cymbaria dahurica</i> L.		FS SB FO LC SG SK AF	PEF
	<i>Linaria vulgaris</i> Mill.		LC	PEF
	<i>Pedicularis striata</i> Pall.		FS SB FO LC SG	PEF
	<i>Veronica incana</i> L.		FS FO LC	PEF
	<i>V. linarifolia</i> Pall. ex Link		SB LC	PEF
	<i>V. sibirica</i> L.		LC SG	PEF
	<i>V. tubiflora</i> L.		LC	PEF
<i>Rubiaceae</i>	<i>Galium verum</i> L.	C ₃	FS SB FO LC SG	PEF
<i>Valerianaceae</i>	<i>Patrinia rupestris</i> Juss.		FS	PEF
<i>Dipsacaceae</i>	<i>Scabiosa comosa</i> Fisch. ex Roem		FS SB FO LC	PEF
<i>Campanulaceae</i>	<i>Adenophora coronopifolia</i> Fisch.		FS SG	PEF
	<i>A. paniculata</i> Nannfeldt		SB SG	PEF
	<i>A. stenanthina</i> (Ldb.) Kitag.		LC	PEF
<i>Compositae</i>	<i>Arctogeron gramineum</i> L.		FS	PEF
	<i>Artemisia anethifolia</i> Weber	C ₃	LC	ANF
	<i>A. annua</i> L.	C ₃	SG LC	ANF
	<i>A. argyi</i> Levl. et Vant.	C ₃	AF	PEF
	<i>A. commutata</i> Bess.		LC SG SK	PEF
	<i>A. desertorum</i> Spreng.	C ₃	AF SG	PEF
	<i>A. dracunculus</i> L.	C ₃	FO LC	PEF
	<i>A. eriopoa</i> Bunge		FS SB FO LC SG	PEF

Table 2 (continued)

	Species	C ₃ /C ₄	Community	PFTs	
<i>Compositae</i> (cont.)	<i>Artemisia frigida</i> Willd.	C ₃	FS SB FO LC SG SK AF	SHR	
	<i>A. gmelinii</i> Web. ex Stechm.		FS SB FO LC SG	SHR	
	<i>A. irtramongolica</i> H.C. Fu		SB FO AF	SHR	
	<i>A. laciniata</i> Willd.		FS SB FO LC	PEF	
	<i>A. japonica</i> Thunb. var. <i>manshurica</i> Kom.	C ₃	LC	ANF	
	<i>A. mongolica</i> Fisch. ex Bess.		LC	PEF	
	<i>A. pectinata</i> Pall. (<i>Neopallasia pectinata</i>) Polsak.	C ₃	SG	ANP	
	<i>A. scoparia</i> Waldst. et Kit.	C ₃	FS SB FO LC SK AF	ANF	
	<i>A. selengensis</i> Turcz. ex Bess.	C ₃	LC	PEF	
	<i>A. sieversiana</i> Willd.	C ₃	AF	ANF	
	<i>Aster alpinus</i> L.	C ₃	FS SB FO LC SG	PEF	
	<i>Crepis crocea</i> (Lamk.) Back.		FS FO	PEF	
	<i>Echinops latifolius</i> Tausch		FS SG	PEF	
	<i>Filifolium sibiricum</i> Kitam.	C ₃	FS SB FO LC SG SK	PEF	
	<i>Heteropappus altaicus</i> (Willd.) Novopokr.	C ₃	FS SB FO LC SG SK AF	PEF	
	<i>Hypochaeris grandiflora</i> Ledeb.	C ₄	LC	PEF	
	<i>Inula britannica</i> L. var. <i>sublanata</i> Kom.	C ₃	LC	PEF	
	<i>I. japonica</i> Thunb.	C ₃	LC	PEF	
	<i>Ixeris chinensis</i> Nakai subsp. <i>graminifolia</i> Kitag.	C ₃	SG	PEF	
	<i>I. sonchifolia</i> Bunge	C ₃	AF SG	PEF	
	<i>Leontopodium leontopodioides</i> Reauv.	C ₃	FS SB LC SG	PEF	
	<i>Ligularia mongolica</i> (Turcz.) D.C.		SB	PEF	
	<i>Olgaea leucophylla</i> Iljin		FS SB LC SG	PEF	
	<i>Picris japonica</i> Thunb.	C ₃	LC SG SK	ANF	
	<i>Rhaponticum uniflorum</i> (L.) D.C.		FS FO LC SG	PEF	
	<i>Saussurea japonica</i> (Thunb.) D.C.	C ₃	SB SG	PEF	
	<i>S. runcinata</i> D.C.	C ₃	SB SG	PEF	
	<i>Scorzonera austriaca</i> Willd.	C ₃	LC SG	PEF	
	<i>S. sinensis</i> Lipsch.		SG	PEF	
	<i>S. sinensis</i> Lipsch. et Krasch.		FS FO	PEF	
	<i>Senecio integrifolius</i> Glairv.		FS SB LC SG	PEF	
	<i>Serratula centauroides</i> L.		FS SB FO LC SG AF	PEF	
	<i>Sonchus brachyotus</i> D.C.		LC	PEF	
	<i>Taraxacum mongolicum</i> Hand.	C ₃	FO	PEF	
	<i>T. ohwianum</i> Kitam.	C ₃	LC SG SK FS FO	PEF	
	<i>Xanthium strumarium</i> L.	C ₃	AF	ANF	
	<i>Youngia stenoma</i> (Turcz.) Ledeb.		FO	PEF	
	<i>Y. tenuifolia</i> (Willd.) Babcret et Stebb.		FS	PEF	
	<i>Monocotyledoneae</i>				
	<i>Cyperaceae</i>	<i>Carex dahurica</i> Kukenth.	C ₃	LC SG SK FS FO AF	SPG
<i>C. duriuscula</i> C.A.M.		C ₃	SG AF	SPG	
<i>C. enervis</i> C.A.M.		C ₄	LC	SPG	
<i>C. korshinskyi</i> Kom.			FS SB FO LC SG AF	SPG	
<i>C. lanceolata</i> Boott			SB LC	SPG	
<i>C. pediformis</i> C.A.M.		C ₄	LC SG	SPG	
<i>Gramineae</i>		<i>Achnatherum sibiricum</i> (L.) Keng		FS SB FO SG SK	HPG
	<i>A. splendens</i> (Trin.) Nevski	C ₄	LC	HPG	
	<i>Agropyron cristatum</i> (L.) Gaertner	C ₃	FS SB FO LC SG SK AF	HPG	
	<i>Aneurolepidium secalinum</i> (Georgi) Kitag.		LC	HPG	
	<i>Avena fatua</i> L.	C ₃	AF	ANG	
	<i>Beckmannia syzigachne</i> (Steud.) Fernald	C ₃	LC	ANG	
	<i>Bromus inermis</i> Leyss.	C ₃	FS SB LC	HPG	
	<i>Calamagrostis epigeios</i> (L.) Roth	C ₃	LC	HPG	
	<i>C. purpurea</i> (Trin.) Trin.		LC SG	HPG	
	<i>Chloris virgata</i> Sw.	C ₄	LC AF	ANG	
	<i>Cleistogenes squarrosa</i> (Trin.) Keng	C ₄	FS SB FO LC SG SK AF	SPG	
	<i>Digitaria ischaemum</i> (Schreb.) Schreb. ex Muhl.	C ₄	LC SG	ANG	
	<i>Echinochloa crus galli</i> (L.) Beauv.	C ₄	LC	ANG	

Table 2 (continued)

	Species	C ₃ /C ₄	Community	PFTs	
<i>Gramineae</i>	<i>Elymus dahuricus</i> Turcz.	C ₃	LC SG	HPG	
	<i>Enneapogon borealis</i> (Griseb.) Honda	C ₄	SG SK	ANG	
	<i>Eragrostis minor</i> Host.	C ₄	LC SG	ANG	
	<i>Festuca ovina</i> L.	C ₃	FS SB FO	HPG	
	<i>F. dahurica</i> L.	C ₃	SG	HPG	
	<i>Helictotrichon schellianum</i> (Hack.) Kitag.		FS SB LC	HPG	
	<i>Hierochloa glabra</i> Trin.	C ₄	FS SB LC SG	HPG	
	<i>Hordeum brevisubulatum</i> (Trin.) Link	C ₃	LC	HPG	
	<i>Koeleria cristata</i> (L.) Pers.	C ₃	FS SB FO LC SG AF	HPG	
	<i>Leymus chinensis</i> (Trin.) Tzvel.	C ₃	FS SB FO LC SG SK AF	HPG	
	<i>Pennisetum centrasiaticum</i> Tzvel.	C ₄	LC SG SK	HPG	
	<i>Phragmites communis</i> Trin.	C ₃	LC	HPG	
	<i>Poa attenuata</i> Trin.		FS SB FO LC SG AF	SPG	
	<i>P. pratensis</i> L.	C ₃	SG	SPG	
	<i>P. sphondylodes</i> Trin. ex Bunge		SG	SPG	
	<i>Psammochloa villosa</i> Trin.		FO	HPG	
	<i>Puccinellia tenuiflora</i> (Turcz.) Scrib. et Merr.	C ₃	LC	HPG	
	<i>Roegneria gmelini</i> Kitag.		LC	HPG	
	<i>R. turczaninowii</i> (Drob.) Nevski.	C ₃	LC SG	HPG	
	<i>Setaria arenaria</i> Kitag.	C ₄	LC	ANG	
	<i>S. glauca</i> (L.) Beauv.	C ₄	FS	ANG	
	<i>S. viridis</i> (L.) Beauv.	C ₄	AF SG	ANG	
	<i>Stipa baicalensis</i> Roshev.	C ₃	FS SB	HPG	
	<i>S. gobica</i> Roshev.	C ₃	SG SK	HPG	
	<i>S. grandis</i> P. Smirn.	C ₃	FO SG AF	HPG	
	<i>S. krylovii</i> Roshev.	C ₃	SK AF	HPG	
	<i>Liliaceae</i>	<i>Allium anisopodium</i> Ldb.		FS SB FO LC SG AF	PEF
		<i>A. bidentatum</i> Fisch. ex Prokh.		SG	PEF
		<i>A. condensatum</i> Turcz.		FS SB LC SG	PEF
		<i>A. neriniflorum</i> (Herbert.) Baker		SG SK	PEF
<i>A. polyrrhizum</i> Turcz.		C ₃	SB FO LC SG AF	PEF	
<i>A. ramosum</i> L.			FS FO SG	PEF	
<i>A. senescens</i> L.		C ₃	FS SB FO LC SG SK AF	PEF	
<i>A. tenuissimum</i> L.		C ₃	FS SB FO SG SK AF	PEF	
<i>A. unisopodium</i> L.			SK	PEF	
<i>Anemarrhena asphodeloides</i> Bunge			FS FO SG SK	PEF	
<i>Asparagus davuricus</i> Fisch. ex Link.			SG SK	PEF	
<i>A. gibbus</i> Ivan. ex Grub.			FO SG AF	PEF	
<i>Hemerocallis minor</i> Mill.			FS SB LC	PEF	
<i>Lilium concolor</i> Salisbury			LC SG	PEF	
<i>L. tenuifolium</i> Fisch.			FS	PEF	
<i>Iridaceae</i>	<i>Iris dichotoma</i> Pall.	C ₃	FS SB LC	PEF	
	<i>I. ensata</i> Pall.		SB SG AF	PEF	
	<i>I. ruthenica</i> Ker-Gawl.	C ₃	LC SG	PEF	
	<i>I. tenuifolia</i> Pall.		FS LC SG SK AF	PEF	
	<i>I. ventricosa</i> Pall.		SB	PEF	

from the moist community to the drier community or from FS to AF, peaked in AF, and was much lower in SB (Fig. 3). The H value of AF was 43 and 45 % greater than those of FS and SB, respectively. The strong linear

regression of PFT diversity with community types (from FS to AF) suggested a tight relationship between PFT diversity, community changes, and precipitation.

Discussion

The use of PFT is a popular means for studying the logical links between physiological and life-history strategies at plant level and community structure at population

level, for predicting the consequences of global change on vegetation and ecosystem processes at both ecosystem and global levels (Chapin 1993, Paruelo and Lauenroth

1996, Wang 2003). Williams and Markley (1973) and Wang (2002c, 2003) tested the relationships between PFT and land management, because PFT can reflect the tendency of vegetation dynamics. One of the crucial questions is how to define the optimal and comparable PFTs for the regional vegetation types (Pillar 1999, Wang 2003). To define PFT, relevant traits must be selected and an appropriate method must be used to classify plants into types. Morphological and photosynthetic pathway types are the common means for PFT classification at different levels, but these two different traits were misused

in many studies (Wang 2003). Morphological functional types brought more details for studying the linkage between species, communities, and ecosystems at local scale, while classification for PFTs by photosynthetic traits (C_3 , C_4 , and CAM) was coarse and not fit for both the studies (*e.g.* community classification and community structure analysis) and land management (*e.g.* grazing and mowing) at plant community, landscape, and local levels in the steppe region (Wang 2003). This is supported by the observations of this study (Table 2, Figs. 1 and 2).

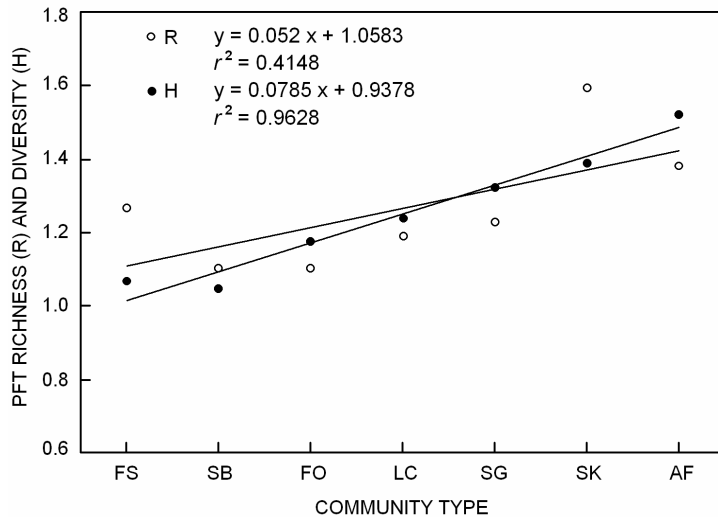


Fig. 3. Plant functional type (PFT) richness (R) index and diversity (D) index in different community types of Inner Mongolia steppe, north China.

Morphological functional types were not only more appropriate for studying the linkage between species, communities, and ecosystems (Smith *et al.* 1993, Wang 2003), but also for steppe management decisions (Wang 2003). Relatively complex geo-relief and landscape in the Xilingol steppe result in vegetation complexities of communities of *Leymus chinensis*, *Filifolium sibiricum*, *Stipa baicalensis*, and *Artemisia frigida* (Li *et al.* 1988, Zhao *et al.* 1988). Morphological functional types are well fit for depicting the differences between these community types (Li *et al.* 1988, Zhao *et al.* 1988) and reflect their seasonal dynamics in the steppe. These may be supported by the observations that species numbers of both PEF and HPG dropped significantly from LC to SK community and from moist to dry seasons, while the species numbers of SPG and SHR types increased remarkably (Fig. 2). Species of ANF, SHR, and SUC types were the main plants composing the dry steppe communities, and probably due to these types the PFTs can use the limited water resource in rainy seasons and survive the dry seasons in the form of dormant seeds (Li *et al.* 1988, Wang

2002a,b,c, 2003). PFT differences among the communities can also be supported by the variables of the R and H indexes observed in this study (Fig. 3). The increase of PFT diversity index from the moist communities to the drier ones was mainly due to the increasing evenness index of the drier communities that indicates that the drier communities are capable to adapt to the drought. This suggests that morphological functional types in the steppe region can indicate both spatial and temporal changes in scales of steppe ecosystem communities.

Photosynthetic pathway type is a too coarse trait for defining PFTs at both the community level and local scale, because of the high C_3 presence and only few families in which C_4 species occur (Wang 2003). This is supported by the observations of this study (Fig. 1) and of the previous research (Wang 2002a,b,c, 2003), but photosynthetic pathway types may be more fit for detecting the PFTs changes at global scale (Wang 2004). This study and the previous ones (McIntyre *et al.* 1999, Pillar 1999) suggest that the identification of optimal PFTs depends on the plant attribute selection and the study scales.

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